

PASI JOLMA

Calcium Metabolism and Vascular Tone in Experimental Hypertension and Renal Failure

ACADEMIC DISSERTATION

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University of Tampere, Medical School, Department of Pharmacological Sciences Tampere University Hospital, Department of Internal Medicine Finland

Supervised by Docent Ilkka Pörsti University of Tampere

Reviewed by Docent Ilkka Kantola University of Turku Docent Eero Mervaala University of Helsinki

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LIST OF ORIGINAL COMMUNICATIONS

This thesis is based on the following original communications, which are referred to in the text by Roman numerals I-V:

- I Jolma P, Kalliovalkama J, Tolvanen J-P, Kööbi P, Kähönen M, Hutri-Kähönen N, X Wu and Pörsti I (2000): High calcium diet enhances vasorelaxation in nitric oxide-deficient hypertension. American Journal of Physiology - Heart and Circulatory Physiology, 279 (3): H1036-H1043.
- II Kähönen M, Näppi S, Jolma P, Hutri-Kähönen N, Tolvanen J-P, Saha H, Koivisto P, Krogerus L, Kalliovalkama J and Pörsti I (2003): Vascular influences of calcium supplementation and vitamin D-induced hypercalcaemia in NaCl-hypertension. Journal of Cardiovascular Pharmacology, 42(3):319-328.
- III Kalliovalkama J, Jolma P, Tolvanen J-P, Kähönen M, Hutri-Kähönen N, Saha H, Tuorila S, Moilanen E and Pörsti I (1999): Potassium channel-mediated vasorelaxation is impaired in experimental renal failure. American Journal of Physiology, 277(4 Pt 2): H1622-H1629.
- IV Jolma P, Kalliovalkama J, Tolvanen J-P, Kööbi P, Kähönen M, Saha H and Pörsti I (2002): Preserved endothelium-dependent but impaired isoprenaline-induced vasorelaxation of the resistance vessels in experimental renal failure. Experimental Nephrology, 10(5-6): 348-354.
- V Jolma P, Kööbi P, Kalliovalkama J, Saha H, Fan M, Jokihaara J, Moilanen E, Tikkanen I and Pörsti I (2003): Treatment of secondary hyperparathyroidism by high calcium diet is associated with enhanced resistance artery relaxation in experimental renal failure. Nephrology Dialysis Transplantation, 18(12):2560-2569.

ABBREVIATIONS

AA	Arachidonic acid
ACh	Acetylcholine
Ang I	Angiotensin I
Ang II	Angiotensin II
ANOVA	Analysis of variance
$[Ca^{2+}]_i$	Intracellular free Ca ²⁺ concentration
Ca ²⁺ pump	Ca ²⁺ -ATPase
cAMP	Cyclic adenosine 3',5'-monophosphate
cGMP	Cyclic guanosine 3',5'-monophosphate
COX	Cyclooxygenase
CRF	Chronic renal failure
EDCF	Endothelium-derived contracting factor
EDHF	Endothelium-derived hyperpolarizing factor
ESRD	End-stage renal disease
ET	Endothelin
G protein	Guanosine 5'-triphosphate-binding protein
IP ₃	Inositol 1,4,5-trisphosphate
K _{ATP}	ATP-sensitive K^+ channels
K _{Ca}	Ca ²⁺ -activated K ⁺ channels
K _{IR}	Inward rectifier K ⁺ channels
L-NAME	N ^G -nitro-L-arginine methyl ester
NA	Noradrenaline
NaCl	Sodium chloride
NO	Nitric oxide
NOS	Nitric oxide synthase
NOx	Nitrite and nitrate
PG	Prostaglandin
PGI ₂	Prostacyclin
РТН	Parathyroid hormone
RAS	Renin-angiotensin system
SH	Secondary hyperparathyroidism
SHR	Spontaneously hypertensive rats
SNP	Sodium nitroprusside
SOD	Superoxide dismutase
VSMC	Vascular smooth muscle cell
WKY	Wistar-Kyoto

INTRODUCTION

The most common risk factor for myocardial infarction, cerebral stroke, and end-stage renal disease (ESRD) is elevated blood pressure. Almost one fourth of the entire population in industrialised societies and more than half of the population aged over 65 years have elevated blood pressure (Ruoff 1998). Recent guidelines for high blood pressure detection and treatment contained key messages for physicians dealing with blood pressure evaluation (Chobanian et al. 2003). The goal blood pressure should be <140/90 mmHg, or <130/80 mmHg for patients with diabetes or chronic kidney disease and in persons older than 50 years, systolic blood pressure or more than 140 mmHg is a more important cardiovascular disease risk factor than diastolic blood pressure (Chobanian et al. 2003). Extensive studies have been carried out to elucidate the complex pathogenesis of hypertension, yet the origin of high blood pressure still remains unknown in 95 % of the patients. It is generally suggested that elevated blood pressure mainly results from increased peripheral arterial resistance, while cardiac output often remains unaltered (Kaplan 1998). Moreover, the leading cause of mortality in patients with ESRD are cardiovascular disease and stroke (Slatopolsky 2003). Cardiovascular disease in uraemia includes disorders of the heart (left ventricular hypertrophy, cardiomyopathy) and disorders of the vascular system (atherosclerosis, arteriosclerosis) (London 2003). Even after stratification by age, gender, race, and the presence or absence of diabetes, cardiovascular mortality in dialysis patients is 10 to 20 times higher than in the general population (Foley et al. 1998, London 2000). This excess cardiovascular risk and mortality seems to already be demonstrable in early renal disease and chronic renal failure (CRF) (London 2003).

In the past, numerous experimental models have been used to study the cause and progression of human cardiovascular disease (Doggrell and Brown 1998). The complex pathomechanims, the individual variation in polygenetic disposition as well as the environmental factors regarding the development of essential hypertension are among the parameters that need to be considered in studying cardiovascular phenomena such as hypertension (Lindpaintner et al. 1992). Defects in the production or action of nitric oxide (NO) (Moncada and Higgs 1993), as well as elevated levels of salt intake may contribute to the pathogenesis of essential hypertension. NaCl-induced hypertension exhibits the cardiovascular effects of long-term salt administration, whereas chronic inhibition of nitric oxide synthase (NOS) represents a more novel model for animal hypertension (Baylis et al. 1992). The most common cause of secondary hypertension is renal parenchymal disease (Preston 1999) whereas, on the other hand, sustained essential hypertension predisposes to the development of renal failure (Frohlich 1997, Luke 1998, Rahn 1998). Therefore, experimental renal failure represents a fascinating model of cardiovascular disease.

The vascular endothelium regulates the contractile state of the underlying smooth muscle by releasing relaxing and contracting factors. The endothelium is therefore an

important regulator of local blood flow and peripheral arterial resistance. Small arteries are the main regulators of peripheral vascular resistance and blood pressure and are therefore often referred to as resistance vessels (lumen diameter < 400μ m) (Bohlen 1986, Schiffrin 1997). Endothelial dysfunction is a sensitive indicator of cardiovascular disease, predicts its prognosis, and is closely associated with the development of arteriosclerosis (Galle et al. 2003). Abnormal endothelium-dependent vasodilatation of small and large arteries has been observed in patients with essential hypertension, and there is accumulating evidence suggesting that CRF is also associated with impaired endothelial function.

More than two decades ago calcium supplementation was shown to lower the blood pressure of spontaneously hypertensive rats (SHR) (Ayachi 1979) and this finding sparked a burst of activity exploring the relationships between dietary calcium intake and blood pressure regulation in human and animal models of hypertension. While the question whether elevated calcium intake lowers blood pressure in humans remains controversial, data showing an inverse correlation between calcium intake and blood pressure in several models of hypertension in the rat and dog very are consistent (Bukoski 2001). Over the past two decades, multiple mechanisms have been postulated to explain how positive calcium balance results in lower blood pressure, yet the final conclusion remains unclear. Oral calcium carbonate is also used as a phosphate binding agent in renal failure to reduce the elevated circulating levels of phosphate and parathyroid hormone (PTH) (Rostand and Drüeke 1999), but the effect of high calcium diet on vascular function during impaired kidney function has not been studied.

The present study was designed to examine the reactivity of large arteries in NO deficiency, sodium chloride (NaCl)-induced hypertension and renal failure. The vascular effects of long-term calcium supplementation in N^G-nitro-L-arginine methyl ester (L-NAME)-hypertension and the influences of high calcium intake and vitamin D-induced hypercalcaemia were evaluated in NaCl fed Wistar rats. Moreover, the reactivity of small mesenteric arteries, as well as the influences of phosphate binding by high calcium intake on the control of microvascular tone in renal failure were studied.

REVIEW OF THE LITERATURE

1 Control of blood pressure

1.1 General aspects

1.1.1 Central and autonomic nervous system

Blood pressure is regulated by a myriad of mechanisms that feature complex interaction between central and peripheral regulatory mechanisms adapting the cardiovascular system to the surrounding requirements (Culman and Unger 1992, MacGregor 1998). Under normal conditions, the kidney plays the dominant role in setting long-term arterial pressure, and the central and autonomic nervous systems act primarily as short-term regulators, stabilizing perfusion pressure in the face of disturbances in circulatory homeostasis (e.g., standing, running, and stress) (Wyss and Carlson 1999). The central nervous system modulates blood pressure via adjusting heart rate and contractility as well as controlling peripheral vascular resistance. In executing the required cardiovascular adatations, the sympathetic and parasympathetic pathways of the autonomic nervous system play the key role, but neuroendocrine pathways, such as the hypothalamo-pituitary axis, are also involved (Reid 1994, de Wardener 2001). Moreover, the sympathetic nervous system also exerts long-term trophic action on the vasculature (Tsuru et al. 2002). Anatomically, areas like the medulla oblongata and its nucleus tractus solitarius are of primary importance in the cardiorespiratory reflex integration and regulation of sympathetic neuronal activity (Colombari et al. 2001).

The effects of central nervous system on blood pressure are modified by arterial baroreceptors located at the aortic arch and the carotid sinuses, which in hypertension appear to be desensitised and reset to a higher level of blood pressure (Sleight 1991, Grassi et al. 1998, Stauss 2002). Although it is generally accepted that the primary purpose of the cardiovascular baroreflexes is to keep blood pressure close to a particular set point over a relatively short period of time, the contribution of baroreceptor dysfunction to long-term essential hypertension remains unclear (Panfilov and Reid 1994, Wyss and Carlson 1999). Nevertheless, in several animal models and in subsets of hypertensive human patients, the nervous system seems to play a more significant role in the chronic elevation of arterial pressure (Wyss and Carlson 1999). SHR have consistently shown indications of a centrally mediated increased sympathetic outflow (Reid 1994). Moreover, both juvenile and adult SHR have shown decreased activity of sympatoinhibitory neurons, suggesting that enhanced sympathetic activity is involved in the development and maintenance of hypertension in SHR (Fujino 1984, Chalmers et al. 1992). However, there are reports questioning the presence of increased sympathetic tone in the development of spontaneous hypertension (Shah and Jandhyala 1995).

Methodological difficulties have obstructed research in humans on the importance of increased sympathetic activity as a cause for long-term elevation of blood pressure (Mancia et al. 1999). Nevertheless, the sympathetic nervous system could be a key factor in the genesis of essential hypertension, and additionally it may also promote the development of its complications (Guzzetti et al 1988, Mancia et al. 1999, Esler 2000). In humans, there has been a clear relationship between cardiac sympathetic activity and the progression of hypertension in its early stages (Julius 1996).

Furthermore, plasma noradrenaline (NA) levels are reported to be elevated, rate of NA spillover from sympathetic nerve terminals could be increased, and muscle sympathetic nerve activity enhanced in patients with essential hypertension as compared with normotensive control subjects (Goldstein 1983, Floras and Hara 1993, Rahn et al. 1999). Sympathetic activity appears to be particularly high in young subjects with borderline hypertension, and increased sympathetic outflow has even been found in the normotensive offspring of hypertensive patients, supporting the idea that increased sympathetic tone could be cause, rather than the consequence, of elevated blood pressure (Floras and Hara 1993, Noll et al 1996). Altogether, dysfunction of the autonomic nervous system may contribute to the development of both essential and experimental hypertension.

1.1.2 Kidneys

Long-term regulation of arterial blood pressure is closely linked to volume homeostasis through renal body fluid feedback mechanisms (Lohmeier 2001). A key feature of the renal body fluid feedback control system is the pressure natriuresis or the ability of the kidneys to respond to changes in arterial pressure by altering the renal excretion of salt and water, i.e. sodium balance, extracellular fluid volume (ECFV) and blood volume (Navar 1997, Lohmeier 2001). In addition to playing a major role in long-term blood pressure adjustment, the sodium transporters along the nephron are very dynamic, even responding quickly to normal fluctuations of blood pressure (McDonough et al. 2003). When arterial pressure is elevated, pressure natriuresis will result in the renal excretion of sodium and water until the blood volume is decreased sufficiently to return the arterial pressure to normal levels (Guyton et al. 1972). Therefore, when the relationship between sodium excretion and arterial pressure is shifted towards higher pressures, hypertension develops (Navar 1997). Moreover, any derangements that compromise the ability of the kidneys to maintain sodium balance potentially result in the kidney's need for an elevated arterial pressure to re-establish net salt and water balance (Navar 1997, Lohmeier 2001). Such resetting of the pressure-natriuresis mechanism has been linked to the development of both human and experimental hypertension (Guyton et al. 1972, Cowley and Roman 1996, Cowley 1997). Recent studies in humans have provided quite surprising evidence of how many genetic forms of hypertension or hypotension solved to date converge an a final common pathway, altering blood pressure by changing net renal salt balance (Lifton et al. 2001). These findings establish the role of altered salt homeostasis in blood pressure variation in humans and underscore the key role of the kidney in the long-term determination of blood pressure.

The sensitivity of the pressure-natriuresis mechanism can be modified by extrarenal hormonal regulatory systems, such as the renin-angiotensin system (RAS) (Lohmeier 2001). As arterial pressure or sodium intake increases, the RAS is suppressed, which enhances the ability of the kidneys to excrete salt and water (Lohmeier 2001). Moreover, the sympathetic nervous system can alter the pressure-natriuresis mechanism and contribute to long-term regulation of arterial pressure through changes in renal sympathetic nerve activity (Lohmeier 2001). Interestingly, one of the classical explanations used for the blood pressure lowering effect of dietary calcium has been the detected natriuretic action of calcium in both humans and animals (Pörsti et al. 1991, Akita et al. 2003). There is considerable evidence from acute studies that baroreflex-mediated changes in renal sympathetic nerve activity influence pressure natriuresis and contribute to short-term regulation of body fluid volumes (Van Vliet et al. 1996, Dibona and Kopp 1997). The long-term effects of the compensatory changes in renal sympathetic nerve activity on body fluid volumes and arterial pressure remain somewhat unclear. However, certain human studies have demonstrated that renal norepinephrine overflow, an index of renal sympathetic nerve activity, is increased in the early stages of essential hypertension (Esler 1995, 2000). Furthermore, experimental studies have shown that chronic renal adrenergic stimulation may result in sustained hypertension (Van Vliet et al. 1996, Dibona and Kopp 1997, Lohmeir 200). It seems that chronic decreases, as well as increases, in renal adrenergic stimulation have a sustained modulatory influence on pressurenatriuresis (Lohmeier 2001).

Renal disease in type 2 diabetes has become an important clinical problem as the prevalence of type 2 diabetes is rising in all Westernized societies. ESRD in patients with diabetes (mostly type 2) as a co-morbid condition has also risen dramatically in the past decade (Ritz and Tarng 2001). This constellation has become the single most common cause of ESRD in most countries (Ritz and Tarng 2001, Deferrari et al. 2002). Aggressive treatment of hypertension and meticulous glycaemic control are the most important clinical strategies in preventing the increase in number of diabetic patients with ESRD (Ritz and Tarng 2001).

The linkage between kidney failure and the elevation of blood pressure is complex and these conditions may coexist at least in three clinical settings (Preston 1999). First, renal parenchymal disease with impaired renal sodium excretion leads to ECFV expansion, which is the most common form of secondary hypertension, accounting for 2.5 % to 5 % of systemic hypertension cases (Preston 1999). Practically every kidney failure patient suffers from hypertension by the time of renal-replacement therapy (Rabelink and Koomans 1997, Luke 1998). Furthermore, pathologically altered hormonal profiles with increased sympathetic

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activity, activated RAS, increased levels of catecholamines, vasopressin and endothelin (ET), enhanced oxidative stress, and decreased NO activity or production may contribute to the high incidence of hypertension in renal failure patients (Bellinghieri et al. 1999, Ligtenberg et al. 1999, Schmidt and Baylis 2000, Annuk et al. 2001, Vaziri et al. 2002, Annuk et al. 2003). Second, it is agreed that sustained, uncontrolled essential hypertension damages the renal vasculature, causes nephrosclerosis and markedly predisposes to the development of renal failure (Frohlich 1997, Luke 1998, Rahn 1998). It has been suggested that hypertension has been the cause of end-state renal disease in 8 % of dialysis patients (Bellinghieri et al. 1999). However, some reports suggest that sufficient blood pressure control in patients with essential hypertension inhibits the development of progressive decline in kidney function (Herrera-Acosta 1994, Siewert-Delle et al. 1998). The third significant clinical circumstance in which hypertension and renal failure occur simultaneously, is ischemic renal disease following by bilateral or unilateral arteriosclerotic renal artery stenosis (Preston 1999). Hence, hypertension can either be a result of renal failure or the leading cause for impaired kidney function.

1.1.3 Renin-angiotensin system

In the 1970s, a series of observations demonstrated that angiotensin II (Ang II) has deleterious effects on the heart and kidney and that patients with high levels of plasma renin activity are at a higher risk of developing stroke or myocardial infarction than those with low plasma renin activity (Gavras et al. 1971, Brunner et al. 1972). Thereafter, the development of pharmacological probes that block the RAS have helped to define the contribution of this system to blood pressure control and to the pathogenesis of diseases such as hypertension, congestive heart failure and CRF (Burnier 2001).

RAS is a regulatory cascade that plays an essential role in the regulation of blood pressure, electrolyte, and volume homeostasis (Li et al. 2002). The first and rate-limiting component of this endocrine cascade is renin, a protease synthesized and secreted predominantly by the juxtaglomerular apparatus in the nephron (Li et al. 2002). Renin cleaves angiotensin I (Ang I) from liver-derived angiotensinogen, which is then converted to Ang II by the angiotensin-converting enzyme (ACE) located in the luminal surface of the vascular endothelium (Riordan 1995, Wright et al. 1995). Although there are other angiotensin peptides with biological effects, Ang II is the major end product of the system and through binding to its receptors Ang II exerts diverse actions that affect the electrolyte, volume and blood pressure homeostasis (Ballerman et al. 1991, Burnier 2001). Ang II has numerous actions on vascular smooth muscle: it modulates vasomotor tone through its potent vasoconstrictor effects, regulates cell growth and apoptosis, influences cell migration and extracellular matrix deposition, it is proinflammatory, and it stimulates production of other growth factors and vasoactive agents (Touyz 2003). Regulation of Ang II-induced vascular

contraction is generally attributed to a G protein-mediated increase in cytoplasmic $[Ca^{2+}]_i$, which is the signal activating the contractile machinery of vascular smooth muscle cells (Touyz and Schiffrin 2000). Ang II is also a major stimulus for activation of NAD(P)H oxidases, which are a predominant source of reactive oxygen species, such as superoxide, that consume NO in vascular cells resulting in endothelial dysfunction (Cai et al. 2003).

Remodeling of small arteries in hypertension is associated with increased VSMC growth (Touyz et al. 2002), and Ang II is one of the most important active agents that have been implicated in this in this vascular process (Geisterfer et al. 1988, Touyz et al. 1999). Ang II appears to have direct growth-promoting effects independent of blood pressure changes, which may contribute to the vascular remodeling in hypertension (Morishita et al. 1994, Kim et al. 1999). Signal transduction pathways underlying Ang II-mediated growth actions involve phosphorylation of tyrosine kinases, such as c-Src, and activation of mitogen-activated protein (MAP) kinases (Takahashi and Berk 1998, Touyz et al. 1999, Touyz 2003). Alterations in tyrosine and MAP kinase signaling are suggested to play a role in the pathological cellular processes that are associated with vascular remodeling in hypertension (Touyz et al. 2002, Touyz 2003).

At first, RAS was described as an endocrine system exerting its action through Ang II, but recently, tissue-based RAS, which acts through paracrine-autocrine mechanisms, has been suggested to be of more importance (Stock et al. 1995, Rothermund and Paul 1998). Local Ang II production has been demonstrated in many tissues including the brain, kidney, adrenal cortex, heart, and blood vessel wall (Cockroft et al. 1995, Mulrow and Franco-Saenz 1996, Kubo et al. 1999). The local synthesis of renin seems to be limited to small amounts and it is likely that uptake of renal renin occurs from the circulation (Stock et al. 1995, Danser 1996). Ang I and Ang II can also be generated by other enzymatic pathways (Urata et al. 1990, Dzau et al. 1993). Ang I can be formed by nonrenin enzymes such as tonin or cathepsin, and Ang I can be converted to Ang II by enzymes such as trypsin, cathepsin, or the heart chymase (Burnier 2001). Today, the quantitative contribution of these alternative pathways to the generation of Ang II remains unclear.

ACE is also called kininase II, and it participates in metabolising bradykinin to inactive peptides. The inhibition of ACE produces an increase in plasma bradykinin levels (Linz et al. 1995, Nussberger et al. 1998), which contributes to the side effects of ACE inhibitors (eg, angioedema) and might play a role in the organ-specific effects of ACE inhibitors (Nussberger et al. 1998).

The discovery of specific Ang II receptor antagonists has confirmed the existence of various subtypes of Ang II receptors (Timmermans et al. 1993). Ang II type 1 (AT₁) receptors are selectively inhibited by losartan, whereas type 2 (AT₂) receptors are inhibited by PD 123177 and related compounds (Burnier 2001). In rodents, AT₁ receptors have been further subdivided into AT_{1A} and AT_{1B} . In amphibians and in neuroblastoma cell lines, an Ang II

receptor inhibited neither by losartan nor by PD 123177 has been classified as AT_3 (Burnier 2001). AT_1 receptors have been localized in the kidney, heart, vascular smooth muscle cells (VSMCs), brain, adrenal gland, platelets, adipocytes, and placenta (Burnier 2001). AT_2 receptors are abundant in the fetus, but their number decreases in the postnatal period (Timmermans et al. 1993, Burnier 2001). In adult tissues, AT_2 receptors are present only at low levels, mainly in the uterus, the adrenal gland, the central nervous system, the heart (cardiomyocytes and fibroblasts), and the kidney (Timmermans et al. 1993, Burnier 2001). AT_2 receptors seem to be re-expressed or upregulated in experimental cardiac hypertrophy, myocardial infarction, and vascular and wound healing (Janiak et al. 1992, Nio et al. 1995, Ohkubo et al. 1997).

However, most of the known clinical effects of Ang II are mediated by the AT_1 receptor (Horiuchi et al. 1999, Burnier 2001). The actions of Ang II via the AT_1 receptor include vasoconstriction, increased sodium retention, suppressed renin secretion, increased ET secretion, increased vasopressin release, activation of sympathetic nervous system, promotion of myocyte hypertrophy, stimulation of vascular and cardiac fibrosis and increased myocardial contractility (Burnier 2001). The physiological role of the AT_2 receptor is only partially understood. In recent years, several new functions have been attributed to AT_2 receptors, including inhibition of cell growth, promotion of cell differentation, and apoptosis (Nakajima etl. 1995, Stoll et al. 1995, Meffert et al. 1996, Morrisey and Klahr 1999). Thus, AT_2 receptors could have an important role in counterbalancing some of the effects of Ang II mediated by AT_1 receptors (Horiuchi et al 1999, Siragy et al. 1999). However, this remains a matter of debate because controversial results have been reported (Li et al. 1998, Cao et al. 1999). There is also data suggesting that AT_2 receptors could mediate the production of bradykinin, NO, and perhaps prostaglandins (PGs) in the kidney (Siragy and Carey 1996).

1.2 Vascular endothelium

Vascular endothelium serves not merely as a passive barrier between flowing blood and the vascular wall but uses this strategic location to maintain vascular homeostasis. It plays a pivotal role in modulating vascular tone, calibre, and blood flow in response to humoral, neural, and mechanical stimuli by synthesizing and releasing vasoactive substances (Behrendt and Ganz 2002). The endothelium also takes part in the regulation of various physiological functions including coagulation, lipid transport, immunological reactivity and vascular structure (Busse and Fleming 1993, Haynes and Webb 1998). The degree of contraction or relaxation of VSMCs characterises the general vasomotor tone, which governs the local blood pressure level and distributes the flow according to metabolic needs. Therefore, by releasing vasoactive substances such as NO, hyperpolarizing factor, cyclooxygenase (COX) metabolites, ET, superoxide and other contracting factors the endothelium continuously adjusts the balance between vasoconstriction and vasodilatation and maintains an adequate

blood flow (Busse and Fleming 1993, Boulanger 1999).

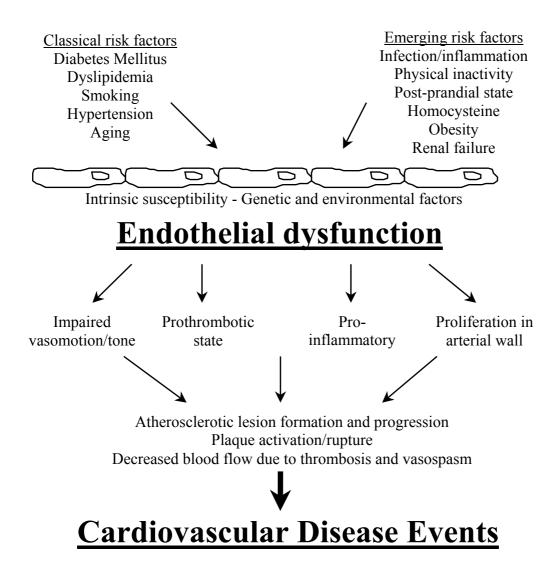
A number of studies have demonstrated that endothelium-mediated arterial relaxation is impaired in patients with essential hypertension (Panza et al. 1990, Taddei et al. 1997a, Schmieder et al. 1997) although this view is not supported by all investigators (Cockcroft et al. 1994, Hutri-Kähönen et al. 1999). Moreover, measurement of endothelial function in patients has recently emerged as a useful tool for atherosclerosis research (Widlansky et al. 2003). Clinical syndromes other than hypertension, such as stable and unstable angina, acute myocardial infarction, claudication, and stroke relate, in part, to a loss of endothelial control of vascular tone, thrombosis, and the composition of the vascular wall (Widlansky et al. 2003). Recent studies have shown that the severity of endothelial dysfunction relates to the risk for an initial or recurrent cardiovascular event, yet a growing number of interventions known to reduce cardiovascular risk also improve endothelial function (Celermaier 1997, Muiesan et al. 1999, Widlansky et al. 2003). Therefore, speculation has been prompted that endothelial function may serve as a "barometer" for cardiovascular health that could be used for patient care and evaluation of new therapeutic strategies (Vita and Keaney 2002). Given this possible causal pathway from endothelial dysfunction to atherosclerosis (Figure 1), numerous methods have been employed to measure endothelial dysfunction in humans. These in vivo methods include intracoronary agonist infusion with with quantitative angiography, brachial artery catheterization with venous occlusive plethysmography, vascular tonometry and measurements of vascular stiffness, and brachial artery ultrasound with flow-mediated dilatation (Widlansky et al. 2003).

In addition to vasomotor dysfunction, circulating blood markers of endothelial dysfunction also have prognostic value. For instance, in patients without known cardiovascular disease, elevated levels of soluble intercellular adhesion molecule (ICAM), and tissue plasminogen activator are independent predictors of future cardiovascular events (Ridker et al. 1998, Thogersen et al. 1998). In patients with known coronary disease, soluble ICAM, von Willebrand factor, tissue plasminogen activator, plasminogen activator inhibitor, and endothelin all have prognostic value (Hamsten et al. 1987, Omland et al. 1994, Thompson et al. 1995, Haim et al. 2002). Furthermore, markers of systemic inflammation, including increased levels of C-reactive protein, are also associated with endothelial dysfunction in human subjects (Fichtlscherer et al. 2000, Hingorani et al. 2000, Prasad et al. 2002, Vita and Loscalzo 2002). These studies illustrate that identifying endothelial phenotype using systemic markers carries prognostic value. However, it remains unknown which individual marker or combination of markers will prove most useful.

The impairment of endothelium-mediated vasodilatation has also been observed in various models of experimental hypertension including SHR (Cohen 1995, Küng and Lüscher 1995, Nava and Lüscher 1995). The manifestation of endothelial dysfunction has been shown to occur before the development of hypertension in SHR (Jameson et al. 1993), and

endothelial function has been reported to be impaired even in the normotensive offspring of hypertensive parents (Taddei et al. 1996). On the contrary, the endothelium-dependent vasodilatation has been found to be preserved during the developmental phase of hypertension in SHR, suggesting that endothelial dysfunction provides no significant pathogenetic contribution to the onset of hypertension (Radaelli et al. 1998). In addition, endothelial dysfunction in humans seems to be independent of the degree of vascular structural alterations and of the aetiology of hypertension, and it is probably more linked to the haemodynamic load (Rizzoni et al. 1998). The pathophysiological basis of endothelial dysfunction is still largely unknown, and some reports have even questioned whether there is any association between endothelial dysfunction and hypertension (Van Zwieten 1997).

Figure 1. The role of endothelial dysfunction in the pathogenesis of cardiovascular disease events.



1.2.1 Endothelium-derived vasodilatory factors

1.2.1.1 Nitric oxide

The pioneering experiments by Furchgott and Zawadski showed that presence of intact endothelium is essential for acetylcholine (ACh) to induce dilation of isolated arteries (Furchgott and Zawadski 1980). In contrast, if endothelium was removed, the arteries constricted in response to ACh. Subsequent studies revealed that ACh stimulated the release of a potent vasodilating substance by the endothelium, identified as NO (Ignarro et al. 1987, Furchgott 1996). When NO is lost – as after mechanical denudation of the endothelium or due to pathologic disease states affecting the endothelium – the normal vasodilator response to ACh is replaced by paradoxical constriction resulting from the direct effect of ACh on vascular smooth muscle (Ignarro et al. 1987, Furchgott 1996). In experimental models of vascular disease, increased superoxide production and the subsequent inactivation of NO, seems to be critically involved in reduced NO bioactivity and endothelial dysfunction (Ohara et al. 1993).

NO is generated by conversion of the amino acid L-arginine to NO and L-citrulline (Palmer et al. 1988) by the enzyme NOS. The isoform NOSIII (eNOS) is constitutively expressed by the endothelium (Behrendt and Ganz 2002). NO exerts its relaxing effect on vascular smooth muscle by activation of guanylate cyclase leading to increased production of cyclic guanosine 3',5'-monophosphate (cGMP) and a reduction in intracellular calcium (Behrendt and Ganz 2002). NO actively mediateds many of the functions exerted by intact endothelium. In addition to its potent vasodilating effect, NO counteracts leukocyte adhesion to the endothelium (Gauthier et al. 1995, Kubes et al. 1991), vascular smooth musle proliferation (Cornwell et al. 1994), and platelet aggregation (de Graaf etal. 1992). These biologic actions of NO emphasize its significance in protection against vascular injury, inflammation, and thrombosis, which are all key events involved throughout the process of atherosclerosis (Behrendt and Ganz 2002).

1.2.1.2 Prostacyclin

Prostacyclin (PGI₂) is a member of the PG family. PGI₂ causes vasodilatation, inhibits platelet aggregation (Busse et al. 1994) and also reduces VSMC proliferation *in vitro* (Weber et al. 1998). PG formation is initiated by the liberation of arachidonic acid (AA) from cell membrane phospholipids by phospholipase A₂. AA is converted into PGG₂ and PGH₂ by COX, and PGH₂ is then further converted into PGI₂ by PGI₂ synthase (Cohen 1995, Gryglewski 1995). PGI₂ is the major prostanoid produced in the vascular cells, although other

PGs are also synthesised in the endothelium (Busse et al. 1994). Production of vascular PGs can be blocked by non-steroidal anti-inflammatory drugs that inhibit COX, such as diclofenac (Vane and Botting 1993).

The vascular endothelial cells release PGI₂ in response to shear stress, hypoxia, and stimulation of various receptors such as muscarinic, B₂-kinin and purinergic P_{2Y} receptors (Gryglewski 1995, Lüscher and Noll 1995). The blood levels of PGI₂ are too low to have any general physiological effects, and therefore PGI₂ is regarded as a local rather than a circulating hormone (Vane and Botting 1993). PGI₂ exerts its actions by binding to membrane receptors on the smooth muscle, which activate adenylate cyclase and subsequently increase the intracellular concentration of cyclic adenosine 3',5'-monophosphate (cAMP) (Busse et al. 1994). The increased intracellular cAMP leads to hyperpolarization of cell membrane, reduction of intracellular Ca2+ concentration, and to decreased sensitivity of contractile proteins to Ca²⁺ (Bülbring and Tomita 1987, Ushio-Fukai et al. 1993, Cohen and Vanhoutte 1995). Compared with the effects of NOS inhibition, blockade of COX has negligible impact on blood pressure (Ruoff 1998). Nevertheless, the inhibition of COX seems to enhance endothelium-mediated vasodilation in SHR as well as in essential hypertensive patients (Jameson et al. 1993, Taddei et al. 1993, Takase et al. 1994, Taddei et al. 1997). Moreover, the conversion of PGH₂ into PGI₂ seems to be diminished in the hypertensive rat endothelium, which results in accumulation of vasoconstrictor PGH₂ (Cohen 1995) and consequently imbalances the endothelial production of COX-derived vasodilator and vasoconstrictor factors. In humans, vasodilator prostanoids actively contribute to the maintenance of basal vascular tone, whereas vasoconstrictor products of COX limit the endothelium-dependent vasodilations (Campia et al. 2002). However, COX products do not appear to play a major role in the endothelial dysfunction of hypertensive patients (Campia et al. 2002).

1.2.1.3 Endothelium-derived hyperpolarizing factor

In various blood vessels, endothelium-dependent relaxations can be accompanied by the endothelium-dependent hyperpolarization of smooth muscle cells (Bolton et al. 1984, Félétou and Vanhoutte 1988, Taylor et al. 1988, Chen et al. 1988, Huang et al. 1988). These endothelium-dependent relaxations and hyperpolarizations can be partially or totally resistant to inhibitors of COXs and NO synthases (Garland and McPherson 1992, Nagao and Vanhoutte 1992), and can occur without an increase in intracellular levels of cyclic nucleotides in the smooth muscle cells (Taylor et al. 1988, Cowan and Cohen 1991, Mombouli JV et al. 1992). Therefore, the existence of an additional pathway that involved smooth muscle hyperpolarization was suggested and attributed to a non-characterized endothelial factor called endothelium-derived hyperpolarizing factor (EDHF) (Mcguire et al. 2001).

Hyperpolarization of smooth muscle induces relaxation by reducing both the open probability of voltage-dependent Ca^{2+} channels, and the turnover of intracellular phosphatidylinositides, thus decreasing the intracellular Ca^{2+} concentration { $[Ca^{2+}]_i$ }(Nelson et al. 1990). As the blood vessel size decreases the contribution of EDHF-mediated responses to endothelium-dependent relaxations increases (Shimokawa et al. 1996), except for the coronary and renal vascular beds in which EDHF plays a major role in conduit arteries as well (Quilley et al 1997, Félétou and Vanhoutte 1999).

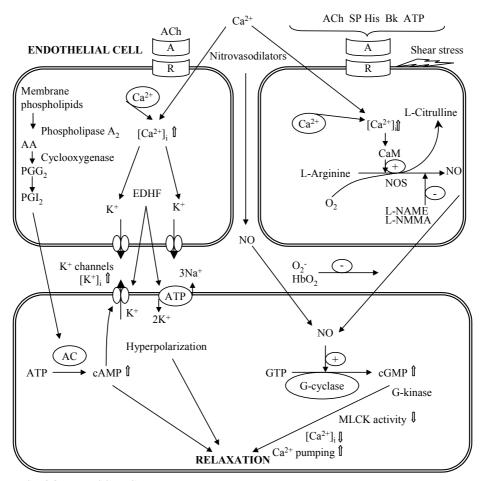
EDHF-mediated relaxations, in response to agonists that stimulate G-protein-coupled receptors, are associated with an increase in $[Ca^{2+}]_i$ in the endothelial cell (Johns et al. 1988, Lückhoff et al. 1988) and are also generated by substances that increase endothelial $[Ca^{2+}]_i$ in a receptor-independent manner (e.g. Ca^{2+} ionophores, and the sarcoplasmic reticulum Ca^{2+} -ATPase inhibitors) (Illiano et al. 1992, Fukao et al. 1995). Conversely, a decrease in the extracellular Ca^{2+} concentration attenuates EDHF responses (Chen and Suzuki 1990), suggesting that for EDHF-mediated responses, as for many other endothelial functions, the increase in endothelial $[Ca^{2+}]_i$ is a crucial step (Nilius and Droogmans 2001).

Previously, several studies have suggested that the endothelium-dependent hyperpolarization of the smooth muscle cells involved an increase in K⁺ conductance and that EDHF was an endothelium-derived K^+ channel opener (Chen et al. 1988, Taylor et al. 1988, Chen and Suzuki 1989). This interpretation has been somewhat modified by new experimental observations. A hallmark of the EDHF-mediated response is its abolition by the combination of apamin [a specific inhibitor of K_{Ca} channels of small-conductance (SK_{Ca} channels)] plus charybdotoxin [a nonselective inhibitor of large-conductactance (BK_{Ca}) and intermediate-conductance (IK_{Ca}) channels, and of some voltage-dependent K⁺-channels] (Corriu et al. 1996, Garland and Plane 1996, Zygmunt and Hoggestatt 1996). Further evidence from studies on endothelial and smooth muscle K⁺ channels suggest that the toxin combination of apamin and charybdotoxin targets two types of K_{Ca} channels (IK_{Ca} and SK_{Ca} channels) expressed on endothelial cells (i.e. prevent endothelial cell hyperpolarization) rather than K⁺ channels activated by an EDHF and located on smooth muscle cells. Iberiotoxin is a novel drug that selectively inhibits the BK_{Ca} of the VSMCs (Marchenko and Sage 1996, Cai et al. 1998, Edwards et al. 1998, Doughty et al. 1999, Edwards et al. 1999b, Edwards et al. 2000, Coleman et al. 2001, Burnham et al. 2002,).

In recent years, the most popular candidates for EDHF have been the arachinoid-acidderived products of cytochrome P450 epoxygenases (fibrates induce cypP450 in Dahl rats), namely epoxyeicosatrienoic acids (EETs) (Cohen and Vanhoutte 1995, Garland et al. 1995, Campbell et al. 1996, Vanhoutte and Mombouli 1996, Fisslthaler et al. 1999, Bolz et al. 2000, Coats et al. 2001, Archer et al. 2003). They are produced by the endothelium, released in response to vasoactive hormones and elicit vasorelaxation via stimulation of K_{Ca} (Quilley et al. 1997) and therefore appear to play an important role in the regulation of vascular homeostasis (Fleming et al. 2001). Moreover, hyperpolarization of endothelial cells might be partly regulated by activation of the cytochrome P450, because EETs might modulate endothelial Ca^{2+} influx in response to Ca^{2+} -store depletion (Hoebel et al. 1997) and might facilitate the activation of endothelial K⁺ channels by increasing their sensitivity to Ca^{2+} (Baron et al. 1997, Li and Campbell 1997). Thefore, EETs and other products of cytochrome P450 seem to be crucial for the initiation and transmission of endothelial cell hyperpolarization, and consequently to EDHF-mediated hyperpolarization and relaxation of VSMCs.

Electrotonic propagation of endothelial cell hyperpolarization via gap junctions between endothelial and smooth muscle cells has been suggested to play some role in the EDHF response in rat hepatic and mesenteric arteries, and to be the sole mechanism underlying the EDHF response in the guinea-pig internal carotid artery (Edwards et al. 1999a). It has also been suggested that the number of these heterocellular myo-endothelial gap junctions increases with the diminution in the size of the artery (Sandow and Hill 2000), a finding that would support the view of more significant contribution of EDHF to the control of vascular tone in smaller arteries (Shimokawa et al. 1996).

A moderate increase in the myo-endothelial K^+ concentration can induce the hyperpolarization of VSMCs by activating the inward rectifier K⁺ channels (K_{IR}) and the Na⁺-K⁺-ATPase (Nelson and Quayle 1995, Prior et al. 1998). Therefore, the K⁺ ion itself has also been suggested to be EDHF in small mesenteric resistance arteries of rats (Edwards et al. 1998, Dora and Garland 2001), although not all investigators are convinced of this claim (Doughty et al. 2000, Lacy et al. 2000). Thus, the nature of the responses attributed to EDHF seem still to be unsolved, but the evidence from several sources suggests that there are multiple EDHFs, and that the chemical mediator of the EDHF response may vary with the vascular bed (Edwards and Weston 1998, Campbell and Harder 1999). Previously, NO has been shown to inhibit both the production and action of EDHF (Bauersachs et al. 1996, McCulloch et al. 1997). Therefore, in pathophysiological states, such as hypertension and renal failure, which may feature decreased bioavailability of endothelium-derived NO, EDHF-mediated vasorelaxation could be enhanced and of greater importance than under physiological conditions (Bauersachs et al. 1996). However, this hypothesis has not been supported by reports where decreased endothelium-mediated hyperpolarization was observed in many forms of experimental hypertension (Fujii et al. 1992, Van de Voorde et al. 1992, Mäkynen et al. 1996, Sunano et al. 1999).



SMOOTH MUSCLE CELL

Figure 2. Schematic diagram shows major mechanisms of endothelium-dependent arterial relaxation. Abbreviations: A, agent; AA, Arachidonic acid; AC, adenylate cyclase; ACh, Acetylcholine; ATP, adenosine 5'-triphosphate; Bk, Bradykinin; $[Ca^{2+}]_i$, intracellular free calcium concentration; $[K^+]_I$, free potassium concentration; cAMP, cyclic adenosine 3',5'-monophosphate; cGMP, cyclic guanosine 3',5'-monophosphate; CaM, calmodulin; Ca²⁺ pump, Ca²⁺-Mg²⁺ ATPase; EDHF, endothelium-derived hyperpolarizing factor; G-cyclase, guanylyl cyclase; GTP, guanosine 5'-triphosphate; Hb, haemoglobin; His, histidine; L-NAME, N^G-nitro-L-arginine; L-NMMA, N^G-monomethyl-L-arginine; MLCK, myosin light chain kinase; NO, nitric oxide; NOS, nitric oxide synthase; PGH₂, prostaglandin G₂; PGI₂, prostacyclin; R, receptor; SP substance P.-, inhibition; +, stimulation; I, increase; I, decrease.

1.2.2 Endothelium-derived contractile factors

The control of blood vessel wall homeostasis is achieved via production of vasorelaxants and vasoconstrictors. In addition to being a source of relaxing factors, endothelial cells also produce contractile factors, such as vasoconstrictor prostanoids, ET-1 and Ang II. The production of endothelium-derived contractile factors (EDCFs) is induced by hypoxia, stretch, pressure, and various local and circulating hormones, and a marked heterogeneity of these responses exists among species, strains, and different vascular beds (Lüscher et al. 1992, Rubanyi 1993, Schiffrin 2001).

1.2.2.1 Cyclooxygenase-derived contractile factors

COX produces several EDCFs, the most potent of which are endothelial thromboxane A₂ and the PG endoperoxide intermediates PGG₂ and PGH₂, which all act via the same receptor (Cohen 1995). In SHR several agonists including ACh and 5-hydroxytryptamine can induce the release of COX-derived contractile factors (Lüscher and Vanhoutte 1986, Auch-Schwelk and Vanhoutte 1991, Ito and Carretero 1992), whereas in Wistar-Kyoto (WKY) rats such responses seem to be absent (Lüscher and Vanhoutte 1986, Jameson et al. 1993). Production of vascular COX-derived contractile factors can be blocked by non-steroidal anti-inflammatory drugs that inhibit COX, such as diclofenac (Auch-Schwelk and Vanhoutte 1991).

Studies in SHR and in patients with essential hypertension have suggested that endothelium-dependent vasodilatation is impaired due to the production of COX-derived factors (Jameson et al. 1993, Taddei et al. 1993, Takase et al. 1994, Küng and Lüscher 1995, Taddei et al. 1997). However, the blockade of thromboxane A₂/PG endoperoxide receptor has no or only a marginal effect on systemic blood pressure in SHR or in patients with essential hypertension (Ritter et al. 1993, Tesfamariam and Ogletree 1995, Ruoff 1998). COX inhibition has been found to enhance endothelium-mediated dilatation in essential as well as in experimental hypertension (Jameson et al. 1993, Taddei et al. 1993, Takase et al. 1994, Taddei et al. 1997). However, the production of COX-dependent EDCFs does not seem to occur in young essentially hypertensive patients, suggesting that COX-derived EDCFs do not participate in the development of human hypertension (Taddei et al. 1997).

2.2.2 Endothelin-1

ETs (ET-1, ET-2, ET-3) are vasoconstrictor peptides that are synthesised from larger precursors, preproendothelins via the formation of proendothelin, which is further cleaved to form the mature ET-1 by ET converting enzymes (Haynes and Webb 1998). Of the three forms characterized the endothelial cells only seem to produce ET-1 (Lüscher et al. 1992, Agapitov and Haynes 2002). ET is formed from the precursor preproendothelin via the formation of proendothelin (big ET). Whether the release of ET requires *de novo* intracellular

protein synthesis initiated by physiological stimuli or chemical substances remains to be elucidated (Kuchan and Frangos 1993, Lüscher et al. 1993a, Lüscher et al. 1993b, Vanhoutte 1993, Lüscher and Noll 1995). Stretching of the vascular wall, hypoxia, low shear stress and substances such as NA, Ang II and thrombin seem to induce the production of ET, whereas high shear stress and endothelial formation of cGMP and cAMP have been shown to inhibit the synthesis of ET (Kuchan and Frangos 1993, Lüscher et al. 1993b, Lüscher and Noll 1995).

Endothelium-derived ET-1 is a potent vasoconstrictor, which acts through smooth muscle ET_A and ET_B receptors that mainly mediate vasoconstriction, and endothelial ET_B receptors that oppose ET_A - and ET_B –mediated vasoconstriction by stimulating NO and PGI₂ formation (Lüscher et al. 1993a, Nava and Lüscher 1995, Taddei et al. 2000). The intracellular mechanims of action of ETs involve activation of phosphatidylinositol metabolism, mobilization of intracellular Ca²⁺ stores, promotion of calcium entry through plasmalemmal Ca²⁺ channels and protein kinase C (Schiffrin 1995a, Black et al. 2003).

Most studies in essential hypertension have reported normal or slightly elevated plasma ET levels, which may be due to the fact that most of the ET-1 is released from the endothelium abluminally towards the vascular smooth muscle rather than into the lumen and circulation (Wagner et al. 1992, Lüscher et al. 1993b, Schiffrin 1995b). Furthermore, even subthreshold concentrations of ET-1 potentiate the effect of other vasoconstrictors such as 5-hydroxytryptamine and NA (Lüscher et al. 1993c). Therefore, the concentration of circulating ET-1 may not accurately reflect its role as a local modulator of vascular tone (Vanhoutte 1993a).

In experimental hypertension the findings concergning the influence of ET-1 on both blood pressure and vascular reactivity have been somewhat inconclusive (Vanhoutte 1993, Nava and Lüscher 1995, Schiffrin 1995b). Although there is evidence for a role of ET-1 in blood pressure elevation in some experimental forms of hypertension, particularly sodiumsensitive hypertension, the significance of ET-1 may be more in accentuating rather than initiating blood pressure elevation (Schiffrin 2001a). However, some studies suggest that hypertensive rats treated with ET antagonists are protected from stroke and renal injury (Schiffrin 2001b). Furthermore, in essential hypertensive patients, the activity of exogenous ET-1 is either increased, similar or decreased as compared to normotensive subjects, depending on which vascular district or scheme of administration is considered (Taddei et al. 2001). However, recent studies suggest that essential hypertension is characterized by increased ET-1 vasoconstrictor tone (Taddei et al. 2001). ET-1 also features a range of other local actions – in the kidney, the nervous system and other hormone systems – that could play a part in the genesis of hypertension (Goddard and Webb 2000). Therefore, potentially the ET antagonists could become effective disease-modifying agents in different forms of cardiovascular disease (Schiffrin 2001b).

1.3 Vascular smooth muscle

1.3.1 Contraction

Contraction of vascular smooth muscle is a complex process that is initiated when vasoconstricting neurotransmitters or hormones activate the cell surface receptors. Most receptors activate different types of guanosine 5'-triphosphate-binding proteins (G proteins) that are coupled to different ion channels and enzymes, and regulate their activities. Among these enzymes are phospholipase C, which metabolises phosphatidylinositol 4,5-bisphosphate and produces inositol 1,4,5-trisphosphate (IP₃) and 1,2-diacylglycerol (DAG), and adenylate cyclase, which metabolises ATP to produce cAMP (Abdel-Latif 1986, Nishizuka 1995). IP₃ releases Ca²⁺ from intracellular store whereas DAG activates protein kinase C (PKC), which phosphorylates a number of proteins (Nahorski et al. 1994). In addition to the activation of the phosphatidylinositol metabolism, vasoconstrictors, such as NA and 5-hydroxytryptamine, have been shown to depolarize the arterial smooth muscle and consequently activate voltage-operated Ca²⁺ channels in the plasma membrane of the smooth muscle, leading to an increased influx of Ca²⁺. The activation of the described mechanisms increases [Ca²⁺]_i, which is the primary signal for smooth muscle contraction (Allen and Walsh 1994).

Following the elevation of $[Ca^{2+}]_i$, Ca^{2+} binds to calmodulin and forms a Ca^{2+} calmodulin complex, which removes the autoinhibition of myosin light chain kinase (Allen and Walsh 1994). The activated myosin light chain kinase phosphorylates reversibly the light chain of myosin and activates the myosin ATPase (Walsh 1994, Winder et al. 1998). The phosphorylated myosin cyclically binds to actin filaments producing force or the shortening of the smooth muscle (Walsh 1994). Should the $[Ca^{2+}]_i$ fall, the myosin light chain kinase is inactivated, which allows dephosphorylation of myosin by myosin light chain phosphatase and causes the smooth muscle to relax (Stull et al. 1991, Cirillo et al. 1992, Rembold 1992, Allen and Walsh 1994). However, the contractile force does not depend directly on $[Ca^{2+}]_{i}$, since the force may be enhanced by augmenting the responsiveness of the contractile machinery or the sensitivity of the myofilaments to $[Ca^{2+}]_i$ (Andrea and Walsh 1992, Ruegg 1999). Changes in free calmodulin concentrations, myosin light chain phosphorylation elicited by small G proteins (e.g. Rho A) and the enzymes associated with them (Rhoassociated kinase), regulation of myosin phosphatase activity and thin filament-associated proteins, such as calponin and caldesmon, are the possible mechanisms for regulation of Ca²⁺ sensitivity (Hori and Karaki 1998, Winder et al. 1998). These mechanisms that change the Ca^{2+} sensitivity, together with the major regulatory mechanisms for cellular Ca^{2+} metabolism, play an important role in the regulation of vascular smooth muscle tone (Walsh 1994, Takuwa 1996, Somlyo et al. 1999).

A multitude of studies in experimental hypertension have demonstrated increased receptor-mediated arterial smooth muscle contractility in these models (Perry and Webb 1991, Brodde and Michel 1992, Orlov et al. 1993). Nevertheless, some investigations have detected only slight differences in the contractile responses between hypertensive and normotensive

animals (Bockman et al. 1992, Tolvanen et al. 1996). Possible mediators for the increased vascular contractility in experimental hypertension include enhanced responsiveness of G proteins (Kanagy and Webb 1994), increased turnover and accumulation of inositol phosphates (Vila et al. 1993) and augmented release of Ca^{2+} from sarcoplasmic reticulum (SR) by IP₃ (Kawaguchi et al. 1993). Furthermore, the generation of DAG stimulated by vasoconstrictor agents and subsequent activation of protein kinase C have been reported to contribute to the enhanced vascular responsiveness (Okamura et al. 1992, Nguyen et al. 1993, Nahorski et al. 1994). Thus, an overactive receptor-mediated contraction pathway could contribute to the pathogenesis of hypertension.

1.3.2 Cellular calcium regulation

Numerous cellular functions are highly influenced by Ca^{2+} metabolism. These include VSMC growth and proliferation as well as contraction of vascular smooth muscle, which is initiated by increased $[Ca^{2+}]_i$ (Karaki and Weiss 1988). There are both extracellular and intracellular Ca^{2+} stores in the vascular smooth muscle (Cirillo et al. 1992), and therefore the $[Ca^{2+}]_i$ is adjusted by a complex interaction between Ca^{2+} entry and extrusion across the plasmalemma, and Ca^{2+} release from and uptake to SR (Marks 1992). Both the plasmalemma and SR maintain a barrier to an approximately 10 000-fold Ca^{2+} concentration gradient. The plasmalemmal Ca^{2+} permeability is under the control of membrane potential and various agonists, whereas Ca^{2+} permeability of SR is controlled by second messangers (Van Breemen and Saida 1989).

Under physiological conditions the Ca^{2+} influx across the plasmalemma takes place either via ion channels or exchangers. The Ca^{2+} channels in plasmalemma are either voltagegated or receptor-operated (Horowitz et al. 1996). The voltage-gated Ca²⁺ channels have been sorted by electrophysiological and pharmacological techniques into two different subgroups: one type is activated by small depolarizations and is rapidly inactivated (T-type), whereas the other requires stronger depolarizations and is more slowly inactivated (L-type) (Spedding and Paoletti 1992). Evidence suggests that sustained depolarization of smooth muscle underlies the increase in arterial tone during hypertension by increasing the open probability of voltagedependent L-type Ca^{2+} channels (Wellman et al. 2001), which increases $[Ca^{2+}]_i$ and contributes to vasoconstriction (Knot and Nelson 1998, Amberg et al. 2003). The L-type channels can be selectively blocked by dihydropyridine Ca²⁺ channel antagonists like nifedipine, and the T-type channels by mibefradil (Nelson et al. 1990, Spedding and Paoletti 1992, Mishra and Hermsmeyer 1994). Some Ca²⁺ ions enter the VSMCs also due to the passive permeability of the plasma membrane to Ca^{2+} (Cirillo 1992). Furthermore, the plasma membrane binds Ca^{2+} and buffers increases in $[Ca^{2+}]_i$, and it may become less permeable to Ca^{2+} following an increase in the extra- and intracellular $[Ca^{2+}]_i$ (Dominiczak and Bohr 1990, Cirillo et al. 1992). In vascular smooth muscle, Ca^{2+} can be extruded from the cell by the plasmalemmal Ca^{2+} pump or the Na⁺/Ca²⁺ exchanger (Allen and Walsh 1994, Horowitz et al. 1996). The Ca^{2+} pump uses energy from ATP hydrolysis and accounts for most of the Ca^{2+} efflux at normal $[Ca^{2+}]_i$. The Na⁺/Ca²⁺ exchange is an antiporter which under basal conditions permits the efflux of one Ca²⁺ ion coupled with the influx of three Na⁺ ions (Cirillo 1992).

The SR plays a major role in Ca^{2+} storage in VSMCs, although special Ca^{2+} binding molecules are also present (Karaki and Weiss 1988, Horowitz et al. 1996). Ca^{2+} is actively sequestered and released by SR following plasmalemmal receptor activation (Minneman 1988, Martonosi et al. 1990, DeLong and Blasie 1993). Activation of cell surface receptors forms IP₃, which releases Ca^{2+} by binding to IP₃-receptors in SR (Marks 1992, Allen and Walsh 1994, Somlyo et al. 1999). Intracellular Ca^{2+} stores can also be mobilised by Ca^{2+} induced Ca^{2+} release, where the influx of a small amount of Ca^{2+} releases more Ca^{2+} from SR via ryanodine receptors (Marks 1992, Allen and Walsh 1994, Horowitz et al. 1996). The physiological significance of this mechanism may be the amplification of IP₃-induced Ca^{2+} release, since Ca^{2+} is a coagonist of IP₃-induced Ca^{2+} release (Finch et al. 1991, Nahorski et al. 1994). The membrane of SR contains also a Ca^{2+} pump, which transports Ca^{2+} ions from the cytosol into SR (Van Breemen and Saida 1989, Allen and Walsh 1994, Horowitz et al. 1996).

Recently, Ca^{2+} oscillations and gradients in vascular smooth muscle have been studied more closely due to the physiological phenomenom of Ca^{2+} ion selectively triggering varying responses in the same cell, for instance Ca^{2+} -mediated responses being different in smooth muscle cells located at different sites (Lee et al. 2002). This is suggested to result from temporal fluctuations and spatial variations of cytoplasmic $[Ca^{2+}]_i$, which depend on the interaction of ion transport proteins of plasma membrane and membranes of SR, nuclear envelope and mitochondria (Lee et al. 2002). All smooth muscle $[Ca^{2+}]_i$ oscillations depend on plasma membrane-SR interactions, but there are two fundamentally different types of $[Ca^{2+}]_i$ oscillations, depending on their immediate source of Ca^{2+} . When $[Ca^{2+}]_i$ rises more or less evenly across the entire cell, no apparent Ca^{2+} waves are observed (Peng et al. 2001), but when the endoplasmic reticulum /SR is the immediate Ca^{2+} source for each Ca^{2+} spike, $[Ca^{2+}]_i$ initially rises in a specific cellular locus, and this regional elevation in $[Ca^{2+}]_i$, propagates in a wagelike fashion throughout the length of the cell (Lee et al. 2001). In VSMCs, both nonwavelike and wavelike $[Ca^{2+}]_i$ oscillations are observed.

The vascular smooth muscle regulates blood flow through selective vasoconstriction and vasomotion, of which the latter is associated with $[Ca^{2+}]_i$ oscillations and the tonic contraction has been thought to be initiated by SR Ca²⁺ release and then maintained by elevated Ca²⁺ influx (Lee et al. 2002). However, confocal microscopy has shown that in many blood vessels agonist-induced contractions are maintained by asynchronous wavelike $[Ca^{2+}]_i$ oscillations in single smooth muscle cells, which summate to give a steady-state elevation in $[Ca^{2+}]_i$ for the whole tissue (Ruehlmann et al. 2000). Moreover, the asynchronous wavelike $[Ca^{2+}]_i$ oscillations appear to be instrumental in the initiation of vasomotion in the rat mesenteric artery (Peng et al. 2001). Ca²⁺ waves have also been associated with the induction of dilatation of cerebral resistance arteries, where the wavelike Ca²⁺ release is thought to stimulate K_{Ca} on the plasma membrane and the relaxing effect of the resulting hyperpolarization-induced closing of voltage-gated Ca²⁺ channels outweighs the local stimulation of contraction (Jaggar 2001). This dual function of Ca^{2+} waves presents an intriguing yet a complex example of vascular heterogeneity. It seems that the manner in which localized Ca^{2+} signals are coupled to either contraction or relaxation is to a large extent determined by the specific ion pumps and channels contained within the plasma membrane – SR junctional complexes (Lee et al. 2002).

Abnormally high $[Ca^{2+}]_i$ has been found in blood cells, cultured aortic and mesenteric arterial smooth muscle cells, and in intact aortas and renal arteries of hypertensive animals (Spieker et al. 1986, Jelicks and Gupta 1990, Sada et al. 1990, Sugiyama et al. 1990, Oshima et al. 1991, Papageorgiou and Morgan 1991, Bendhack et al. 1992, Arvola et al. 1993b, Ishida-Kainouchi et al. 1993), but not all reports confirm this abnormality (Liu et al. 1994, Neusser et al. 1994). Importantly, studies on VSMCs from resistance arteries have exhibited comparable basal $[Ca^{2+}]_i$ between SHR and WKY rats (Storm et al. 1992, Bukoski et al. 1994, Bian and Bukoski 1995), suggesting that the elevations in $[Ca^{2+}]_i$ found in aortic smooth muscle cells and in other cell types of hypertensive animals are unlikely to contribute to the heightened peripheral vascular resistance in SHR (Dominiczak and Bohr 1990, Bian and Bukoski 1995).

Contractile responsiveness of VSMCs from conduit arteries of SHR are enhanced to depolarization and Bay K 8644, an agonist of dihydropyridine-sensitive Ca^{2+} channels, when compared with WKY (Aoki and Asano 1986, Aoki and Asano 1987, Bruner and Webb 1990). Furthermore, augmented vascular sensitivity to the effects of nifedipine has been found in prehypertensive and adult SHR (Aoki and Asano 1986, Aoki and Asano 1987, Asano et al. 1995). In resistance arteries, an increase in the Ca^{2+} influx by the voltage-dependent Ca^{2+} channels has been found in the early hypertensive stage, but not in prehypertensive SHR (Arii et al. 1999). The increased amplitude of the whole-cell Ca^{2+} current in the arterial smooth muscle cells from SHR compared with WKY rats may be attributed to enhanced sensitivity of dihydropyridine receptors in the Ca^{2+} channels in SHR, while the opening properties of a single Ca^{2+} channel have been suggested to be unaltered (Kubo et al. 1998). These results indicate that Ca^{2+} entry through voltage-operated Ca^{2+} channel is enhanced in SHR when compared with WKY rats, which could partially account for altered Ca^{2+} homeostasis and increased vascular reactivity, and thus contribute to increased peripheral resistance and the genesis of hypertension (Arii et al. 1999).

In patients with essential hypertension and in SHR, less Ca^{2+} seems to be bound to the plasma membrane, and the Ca^{2+} permeability of the membrane seems to be increased (Lamb et al. 1988, Dominiczak and Bohr 1990). The extrusion of Ca^{2+} through the plasmalemma by the Na⁺/Ca²⁺ exchange has been reported to be enhanced in aortic VSMCs of SHR (Ashida et al. 1989), whereas depressed activities have been found in tail arteries (Thompson et al. 1990). Studies on the Ca²⁺ pump-mediated Ca²⁺ efflux in VSMCs of SHR have also yielded contradictory results (Kwan and Daniel 1982, Ashida et al. 1989, Monteith et al. 1996, Monteith et al. 1997). Furthermore, the ability of SR to sequester Ca²⁺ has been proposed to be attenuated in SHR (Dohi et al. 1990, Kojima et al. 1991). In addition, SR of SHR appears to have a larger capacity to store Ca²⁺, but the filling of SR is slower when compared with

WKY rats (Kanagy et al. 1994). These findings could result from reduced activity of SR Ca^{2+} pump. Nevertheless, the activity and density of SR Ca^{2+} pump have been reported to be increased in VSMCs of SHR (Levitsky et al. 1993), and also the levels of SR Ca^{2+} pump mRNA were shown to be higher in VSMCs from SHR than in those from WKY rats (Monteith et al. 1997).

Collectively, if $[Ca^{2+}]_i$ is to be elevated, then the Ca^{2+} entry must be increased, or the storage of Ca^{2+} into SR must be decreased, of the extrusion of Ca^{2+} must be decreased. There is no clear evidence whether any of these abnormalities are present in hypertensive VSMCs (Gonzales and Suki 1995). Finally, it has been suggested that one more link between the metabolism of Ca^{2+} and the control of arterial tone could be the extracellular Ca^{2+} receptor in the perivascular sensory nerves, the activation of which can cause vasorelaxation via the release of a hyperpolarizing mediator (Bukoski 1998, Ishioka and Bukoski 1999).

1.3.3 Na⁺-K⁺-ATPase

The Na⁺-K⁺-ATPase carries out the coupled extrusion and uptake of Na⁺ and K⁺ across the plasma membrane of VSMCs (Kaplan 2002). The Na⁺-K⁺-ATPase creates a significant part of the membrane potential in VSMC by extruding three Na⁺ ions for every two K⁺ ions pumped inwards, consequently generating a hyperpolarizing current (Kaplan 2002). Cardiac glycosides such as digoxin and ouabain inhibit vascular Na⁺-K⁺-ATPase (Blaustein 1993, O'Donnell and Owen 1994). Na⁺-K⁺-ATPase activity is suggested to influence $[Ca^{2+}]_i$ in VSMC via voltage-operated Ca²⁺ channels and Na⁺/Ca²⁺ exchange. Therefore, decreased Na⁺-K⁺-ATPase activity leads to membrane depolarization and increased Ca²⁺ influx through voltage-operated Ca²⁺ channels. In addition, decreased activity promotes Na⁺ retention in VSMC, which reduces the driving force of Na⁺/Ca²⁺ exchange leading to attenuated Ca²⁺ extrusion (Bova et al. 1990, Rayson and Gilbert 1992). However, it has been suggested that the latter mechanism does not play an important role in the modification of $[Ca^{2+}]_i$ in VSMCs of SHR (Zhu et al. 1994).

Disturbed function of Na^+-K^+ -ATPase has been suggested to contribute to the pathogenesis of both essential and experimental hypertension (Hermsmeyer 1987), but reports on this matter remain controversial. Na^+-K^+ -ATPase activity has been reported to be either decreased, normal or increased in VSMCs of SHR when compared with those of WKY rats (Tamura et al. 1986, Manjeet and Sim 1987, Rinaldi and Bohr 1989, Kuriyama et al. 1992, Orlov et al. 1992, Redondo et al. 1995). The reason for decreased activity of Na^+-K^+ -ATPase in hypertension could be an inherent defect of a circulating digitalis- or ouabain-like inhibitor of the sodium pump (Blauestein, Hamlyn et al. 1996). The plasmas from patients with essential hypertension have been proposed to contain ouabain-like factors (Masugi et al. 1987), and the concentrations of these Na^+-K^+ -ATPase inhibitors may be elevated, especially in chronic renal insufficiency (Kelly et al. 1986). Circulating sodium pump inhibitor concentrations have even been suggested to be sufficient to affect the Na^+-K^+ -ATPase in human mesenteric arteries, thus the interaction of digitalis-like Na^+,K^+ -pump inhibitors with

the Na⁺-K⁺-ATPase could be of importance (Bagrov and Fedorova 1998), but this remains unclear (Pidgeon et al. 1996). In both rats and humans, reports on the effects of ouabain on blood pressure have been inconsistent and therefore it is not known whether the inhibition of Na⁺-K⁺-ATPase actually elevates blood pressure (Pidgeon et al. 1996).

The function of Na^+,K^+ -pump affects nucleid acid synthesis and smooth muscle cell proliferation, which could contribute to vascular remodelling (Orlov et al. 2001). Moreover, vascular superoxide seems to impair smooth muscle cell Na^+-K^+ -ATPase activity (Gupta et al. 2002), which could be of importance in both hypertension and renal failure. Previously, increased activity of Na^+-K^+ -ATPase has been linked to the increased vascular tone and smooth muscle growth found in SHR (Berk et al. 1989, Davies et al. 1991, Kemp et al. 1992). The activity of Na^+-K^+ -ATPase may be enhanced in hypertension due to an increase in passive membrane permeability of Na^+ , and since the pump does not appear to fully compensate for this, the vascular tone may be elevated (O'Donnell and Owen 1994). These contradictory results on vascular Na^+-K^+ -ATPase in hypertension possibly reflect the fact that the pump's functional, enzymatic and biochemical properties may not be uniformly altered in hypertension (Young et al. 1988), and that the type and duration of hypertension affect these results. Thus, it remains to be clarified whether the altered Na^+-K^+ -ATPase activity in VSMCs is a key factor in hypertension (O'Donnell and Owen 1994).

1.3.4 K⁺ channels

The intracellular concentration of K^+ in the vascular smooth muscle is approximately 25-fold higher when compared with the extracellular concentration of K^+ , and therefore the opening of K^+ channels is accompanied by an efflux of K^+ from the cytosol, resulting in a loss of positive charge and hyperpolarization of the cell membrane (Quast et al. 1994, Jackson 2000). Potassium channels are the dominant ion conductive pathways in VSMCs (Nelson and Quayle 1995, Jackson 1998), therefore significantly contributing to the determination and regulation of smooth muscle membrane potential, which in turn controls $[Ca^{2+}]_i$ and thus contraction of vascular smooth muscle (Standen and Quayle 1998). Therefore, the factors that regulate the activity of K⁺ channels in arterial smooth muscle significantly influence arterial tone, arterial diameter, vascular resistance and blood pressure (Standen and Quayle 1998).

VSMCs express at least four different types of K^+ channels in their plasma membranes (Nelson and Quayle 1995, Jackson 1998). The identified subtypes of K^+ channels in vascular smooth muscle are ATP-sensitive K^+ channels (K_{ATP}), K_{Ca} , voltage-dependent K^+ channels, and K_{IR} channels (Jackson et al. 1997, Jackson 1998, Jackson and Blair 1998, Jackson 2000). The K_{ATP} channels were first found in cardiac myocytes (Noma 1983) and they close as intracellular ATP concentration increases. The open state probability of these channels increases via elevation in cytosolic adenosine 5'-diphosphate or intracellular acidification (Nelson 1993, Ishizaka 1999). However, K_{ATP} are also regulated by several signal transduction pathways independent from changes in ATP. Glibenclamide, a selective K_{ATP} channel blocker, causes arterial constriction in a number of species, including humans, whereas K_{ATP} channel agonists such as cromakalim, levcromakalim, diazoxide and pinacidil dilate arteries (Quast et al. 1994, Jackson 2000). Functional experiments have suggested that there are open K_{ATP} channels in the VSMCs, and this has been confirmed by electrophysiological measurements (Jackson et al. 1997). The basal opening state of coronary vascular K_{ATP} appears to be activated to a greater extent in SHR than WKY rats (Numaguchi et al. 1996). An impaired action of levcromakalim on K_{ATP} was found in SHR, which was restored by the normalisation of blood pressure. This suggests that the impairment can be attributed to high blood pressure (Ohya et al. 1996).

 K_{Ca} channels are found in most cells and are activated by increases in $[Ca^{2+}]_i$ and membrane depolarization (Nelson and Quayle 1995, Carl et al. 1996). K_{Ca} are divided into large, intermediate and small according to their conductance. Large-conductance K_{Ca} (BK_{Ca}) seem to be the most important K^+ channels in the regulation of arterial tone (Amberg et al. 2003). In arteries that display myogenic tone, activity of BK_{Ca} channels has been reported to contribute to resting membrane potential: blockade of the channels with iberiotoxin (selective inhibitor of BK_{Ca}), charybdotoxin (non-selective inhibito of BK_{Ca}), iberiotoxin or tetraethylammonium (non-selective K⁺ channel inhibitor) ions leads to membrane depolarization and vasoconstriction. The small-conductance K_{Ca} can be blocked by apamin (Nelson et al. 1990, Brayden and Nelson 1992, Kitazono et al. 1995, Nelson and Quayle 1995, Nelson et al. 1995). BK_{Ca} channels seem to play a central role in the regulation of vascular tone due to focal increases in subsarcolemmal Ca^{2+} (i.e. Ca^{2+} sparks) by Ca^{2+} released through ryanodine receptors in the SR (Nelson et al. 1995, Jaggar et al. 1998, Knot et al. 1998, Amberg et al. 2003). Furthermore, BK_{Ca} channels play a negative feedback role to limit active vasoconstriction and prevent vasospasm (Jackson 2000, Amberg et al. 2003). BK_{Ca} channels may be activated by vasodilators acting through the cGMP and cAMP cascades (Nelson and Quayle 1995, Paterno et al. 1996), epoxides of AA (Campbell et al. 1996), and carbon monoxide (Wang and Wu 1997). Furthermore, vasodilators and vasoconstrictors may influence the frequency and amplitude of Ca²⁺ sparks and thus effect BK_{Ca} channel activity (Jaggar et al. 1998, Porter et al. 1998, Amberg et al. 2003). Expression of BK_{Ca} channels in membranes of VSMCs was found to be increased in hypertension (Liu et al. 1997b,) which has been suggested to occur as a negative feedback response to the increased vascular reactivity observed in hypertension (Rusch and Liu 1997). However, it was recently reported that the expression of the B1 subunit of the BK_{Ca} channels is downregulated in hypertension (Amberg et al. 2003) warranting future studies on the expression of BK_{Ca} channels in the VSMCs.

Voltage-activated K^+ channels, inhibited by 4-aminopyridine, are another ubiquitous class of K^+ channels expressed by VSMCs (Nelson and Quayle 1995). These channels are activated by membrane depolarization with threshold potentials for substantial activation of - 30mV, and they may participate in the regulation of resting membrane potential and vascular tone (Nelson and Quayle 1995, Jackson et al. 1997, Jackson 1998). Their role in vivo has not been explored, largely because of the lack of availability of inhibitors selective for the channels expressed in vascular muscle cells. However, electrophysiological studies indicate a

decreased functional expression of voltage-dependent K^+ channels in vascular muscle cells from hypertensive animals, which may contribute to depolarization and predispose to increased vascular tone in hypertension (Martens and Gelband 1996). K_{IR} pass inward K^+ current much more readily than outward current with physiological ion gradients and also show a parallel rightward shift in the potential at which rectification appears (activation potential) and a large increase in conductance with increases in the extracellular K^+ concentration (Jackson 2000). The role of K_{IR} in the regulation of resting membrane potential and smooth muscle tone remains somewhat unclear. Some reports suggest that K_{IR} channels mediate K^+ -induced vasodilation in cerebral and coronary resistance arteries (Nelson and Quayle 1995, Knot et al. 1996, Quayle et al. 1997), but the role if K_{IR} in K^+ -induced vasodilation of skeletal muscle arteries is yet to be clarified.

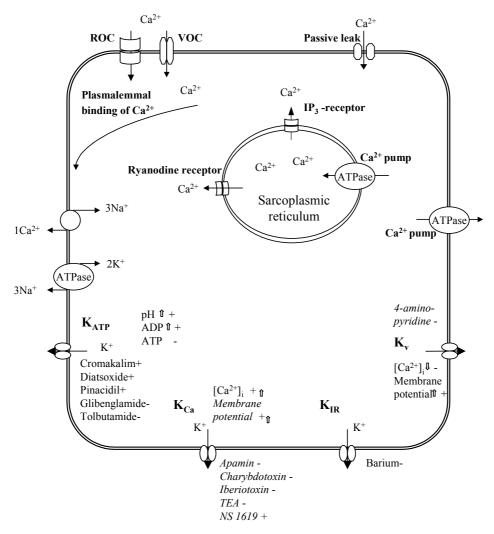


Figure 3. Schematic diagram shows the major mechanisms involved in the cellular Ca²⁺ regulation and some physiological and pharmacological properties of smooth muscle K⁺ channels. Abbreviations: ADP, adenosine 5'-diphosphate; ATP, adenosine 5'-triphosphate; Ca²⁺ pump, Ca²⁺-Mg²⁺ ATPase; IP₃, inositol 1,4,5-trisphosphate; ROC, receptor-operated calcium channel; VOC, voltage-operated calcium channel; [Ca²⁺]_i, intracellular free calcium concentration, $E_{\rm K}$, potassium equilibrium potential; K_{ATP}, ATP-sensitive K⁺ channel; K_{Ca}, Ca²⁺-activated K⁺ channel; K_{IR}, inward rectifier K⁺ channel; K_V, voltage-dependent K⁺ channel; TEA, tetraethylammonium; V_m, membrane potential. -, inhibition; + stimulation;¹, increase;¹, decrease.

2 Arterial tone and structure in experimental hypertension and renal failure

2.1 Hypertension induced by NO-deficiency

Variants of the endothelial NOS gene may be associated with elevated blood pressure (Wang et al. 1997), although some reports do not support this view (Kato et al. 1999, Takami et al. 1999). However, sufficient production of NO in the vascular endothelium seems to be essential for the maintenance of normal blood pressure (Huang et al. 1995), and defects either in the production or action of NO are likely to be associated with essential hypertension (Moncada and Higgs 1993). Therefore, chronic inhibition of NOS by L-NAME is theoretically an interesting model of experimental hypertension (Baylis et al. 1992). Previously, chronic administration of L-NAME has been shown to result in sustained and dose- and time-dependent hypertension in normotensive rats (Baylis et al. 1992, Deng et al. 1993, Arnal et al. 1992, Ribeiro et al 1992, Zatz and Baylis 1998). Furthermore, inhibiting NOS by L-NAME consistently leads to impaired endothelium-dependent arterial relaxations (Küng et al. 1995, Dowell et al. 1996, Henrion et al. 1996, Henrion et al. 1997, Kalliovalkama et al. 1998) as well as arterial remodelling (Deng et al. 1993, Li and Schiffrin 1994, Takemoto et al. 1997, Kalliovalkama et al. 1998, Sakamoto et al. 2002). Data regarding the endothelium-independent arterial relaxations and vascular contractility during NO-deficiency appear somewhat inconsistent (Küng et al. 1995, Dowell et al. 1996, Henrion et al. 1996, Kalliovalkama et al 1998). L-NAME hypertension has also elicited changes in cardiac chamber geometry, myocardial diastolic mechanical properties and increased the amount of myocardial fibrosis (Akuzawa et al. 1998, Matsubara et al. 1998, Bernatova et al 2002). These alterations typically lead to left ventricular hypertrophy, which may further be augmented by metabolic disturbances and ultrastructural alterations such as increased permeability of the cardiac capillaries followed by extracellular and mitochondrial edema (Tribulova et al. 2000, Bernatova et al. 2002). Moreover, in rats exposure to chronic administration of L-NAME seems to produce nephrosclerosis and hyaline arteriopathy as well as impair renal function, which resemble the changes and complications caused by advanced essential hypertension (Hropot et al. 1994, Akuzawa et al. 1998, De Gracia et al. 2000).

The pathogenesis of cardiovascular remodeling in this model has been suggested to result from increased oxidative stress in the endothelium (Usui et al. 1999). Excessive production of superoxide radicals may result from local vascular RAS activation and, in particular, from an increase in local ACE expression in rats with long-term NOS inhibition (Takemoto et al. 1997, Usui et al. 1999). Blockade of the synthesis of NO seems also to upregulate the expression of vascular endothelial growth factor in the coronary arterial smooth muscle cells, which potentially could contribute to the increased wall-to-lumen ratios (Sakamoto et al. 2002). Furthermore, hypertension and cardiovascular remodelling induced by NOS blockade have been suggested to be in part due to an elevation of plasma aldosterone

concentration secondary to increased Ang II type 1 receptor expression in the adrenal gland (Usui et al. 1998). There is also evidence that the autonomic nervous system plays a major role in the L-NAME–induced hypertension via both sympathetic overactivity and vagal suppression (Cunha et al. 1993, Souza et al. 2001). It is of note, that substantial levels of constitutive NOS activity have been shown to remain in the aorta of WKY rats during chronic L-NAME administration (Zhao et al. 1999). Therefore, other mechanisms than simple inhibition of endothelium-derived NO synthesis may be involved in the long-term cardiovascular effects of L-NAME in the rat arteries (Zhao et al. 1999). Recently, the involvement of Rho-kinase in vascular remodeling in L-NAME hypertension was reported, which is of interest because the Rho-kinase pathway has been demostrated to play a role in various models of vascular disease (Ikegaki et al. 2001). Nevertheless, since the exact role of diminished NO production in human hypertension is not absolutely solved, it remains to be established whether NOS inhibition is a relevant experimental model of human forms of hypertension.

2.2 Hypertension induced by NaCl

The causal relation between salt intake and high blood pressure has been widely recognized, but the exact pathophysiological mechanisms and the reason for this relation are less clear (Law et al. 1991, MacGregor and Sever 1996, Cutler et al. 1997, Kurokawa 2001). Blood pressure is affected by dietary salt in many, but not in all subjects (Dustan et al. 1986, Weinberger et al. 1986, Haddy 1991). Nevertheless, a large international study showed that the amount of salt consumed related to blood pressure levels, both within populations and between populations, and was responsible for much of the increase in blood pressure that occurs with increasing age in Western populations (Elliot et al. 1996). Moreover, a Finnish prospective study suggested that high sodium intake predicts mortality and risk of coronary heart disease, independent of other cardiovascular risk factors, including blood pressure (Tuomilehto et al. 2001). Intervention studies have also demonstrated the importance of salt intake in regulating human blood pressure (Forte et al. 1989, Geleijnse et al. 1997). An experiment in chimpanzees, a group of which were fed their natural diet, i.e. similar to that which humans and chimpanzees ate during evolution (10 mmol sodium per day), was compared to a group where salt intake was increased to the same amount that we now consume, 150-200 mmol per day. Over the 2 years of the study, there was a progressive increase in blood pressure (systolic blood pressure increasing by >30 mmHg) in those chimpanzees on the higher salt intake diet (Denton et al. 1995). Human and animal studies have shown a genetic predisposition to develop hypertension in response to an increase in dietary NaCl intake (Sanders 1996). Dissection of the etiologic factors causing salt sensitivity has proved difficult in large, genetically heterogenous populations of rats and humans. For this reason, inbred models of hypertension (such as the Dahl rats or transgenic mice) have been used to examine the mechanism of salt sensitivity (Sanders 1996). In humans, the African-American population has a disproportionately high age-adjusted prevalence of hypertension and many are particularly sensitive to dietary salt (Burt et al. 1995).

In a physiological sense, almost all NaCl is present in the extracellular fluid (ECF) and NaCl is the main determinant of ECF tonicity. Therefore, extracellular fluid volume (ECFV) is determined by the NaCl content of the body, and thus varies with salt intake (Kurokawa 2001). In rats, the expansion of plasma and blood volume as well as of total body water seems to be greater in the young animals subjected to salt-loading than of those in adulthood (Zicha and Kunes 1999). The renin-angiotensin-aldosterone-system (RAAS) participates in the regulation of ECFV by reacting to changes in ECFV and leading to reabsorption of Na²⁺ to maintain ECFV (Kurokawa 2001). The RAAS is activated in states of low salt intake and suppressed by high salt intake (Aguilera and Catt 1981, Schmid C et al. 1997). Renal renin activity is suppressed to a greater extent by high salt intake in younger rats, and the inactivation of renal RAS is accompanied by decreased resistance in the afferent arterioles which leads to increased glomerular capillary pressure predisposing to glomerular damage (Dworkin et al. 1984, Jelínek et al. 1990). Moreover, as the ECFV increases during salt loading, more shear stress subsequently acts in parallel to the blood vessel surface activating the endothelium and altering the expression of various genes in the arterial wall (Ying and Sanders 2002). There is also evidence suggesting that a neurogenic mechanism is involved in salt hypertension, via an early interaction between vasopressinergic and adrenergic neurons in the central nervous system. This leads to a persistent hyperadrenergic state, which together with a suggested baroreceptor reflex dysfunction serve as neurogenic alterations that increase peripheral resistance and further amplify the salt-induced hypertension (Gavras and Gavras 1989, Tanoue et al. 2002).

Both structural changes of the vascular bed, including vascular rigidity and hypertrophy of large arteries, and abnormal control of vascular tone contribute to the hemodynamic effects exerted by increased salt intake (Zicha and Kunes 1999, Et-taouil et al. 2001). High salt intake results in changes in the microvascular structure and function even in the absence of increased arterial blood pressure (Boegehold 2002). Moreover, endothelium-dependent vasodilation and vascular smooth muscle hyperpolarization are impaired by high salt diet (Sylvester et al. 2002, Lombard et al. 2003) and salt loading is reported to downregulate various NOS isotypes, including endothelial, which could contribute to the development and maintenance of elevated peripheral resistance (Ni and Vaziri 2001). Due to the many age-related abnormalities of BP regulation in experimental NaCl hypertension, it should be noted that the maturation of the structure and biochemical composition of rat arteries is achieved relatively late in ontogeny. Aortic collagen biosynthesis reaches the adult level at the age of 7-9 wk, whereas the morphological and mechanical properties of conduit arteries become stabilized in rats aged 10-12 wk and small arteries at 9-11 wk (Zicha and Kunes 1999).

2.3 Renal failure

Cardiovascular complications are the leading cause of mortality in patients with ESRD and the excess cardiovascular risk and mortality is already demonstrable in early renal disease and CRF (London 2003). Even after stratification by age, gender, race, and the presence or absence of diabetes, cardiovascular mortality in dialysis patients is 10 to 20 times higher than in the general population (Foley et al. 1998). Cardiovascular disease in uraemia includes disorders of the heart (left ventricular hypertrophy, cardiomyopathy) and disorders of the vascular system (atherosclerosis, arteriosclerosis). The vascular characteristics of patients with renal failure seem to exhibit abnormal elastic properties of large arteries, reflected as decreased distensibility and compliance (Barenbrock et al. 1994, London et al. 1996). These alterations are independent of the level of blood pressure and tensile stress but appear to be related to the uremic state per se (Luik et al. 1997, Mourad et al. 1997). Furthermore, in patients with ESRD, both aortic and carotid arterial stiffness and presence of vascular calcification are reported to be strong independent predictors of mortality (Blacher et al. 1998, Blacher et al. 1999, Blacher et al. 2001, London 2003). Increased intima-media thickness of carotid arteries is already present in patients with renal failure before they start hemodialysis, supporting the concept that the arterial alterations are not due to the hemodialysis treatment, but due to renal failure per se or the associated secondary metabolic abnormalities (Shoji et al. 2002). Coronary-artery calcification is common even among young adults with ESRD (Goodman et al. 2000). In experimental renal failure, the increased arterial wall thickness is suggested to result mainly from an increase in extracellular matrix, although hyperplasia of the VSMCs could also be involved (Amann et al. 1997). Plasma ET levels are elevated in patients with kidney failure (Warrens et al. 1990), which could partially underlie the associated vascular remodelling (Demuth et al. 1998). Nevertheless, in addition to the structural changes, alterations in arterial function contribute to increased vascular stiffness as well (Mourad et al. 1997).

Accumulating evidence suggests that CRF is associated with impaired endothelial function (Annuk et al. 2003). The functional changes have been attributed to the impaired NO-mediated endothelium-dependent vasodilatation as observed in brachial arteries of hemodialysis patients (Joannides et al. 1997, van Guldener et al. 1997). Even in patients with mild renal insufficiency, endothelial function is abnormal in the absence of atherosclerotic vascular disease, which suggests that uraemia may directly promote the development of atherosclerosis early in the progression of CRF (Thambyrajah et al. 2000). Potential candidates for an atherogenic "uremic factor" include homocysteine, increased oxidative stress, endogenous inhibitors of NO synthase such as ADMA and N^G-monomethyl-L-arginine (L-NMMA), chronic inflammation and accumulation of oxLDL (Steinvinkel et al. 1999, Morris et al. 2001, Annuk et al. 2003). In the arteries of reduced renal mass hypertensive rats,

not only the endothelium-dependent relaxations mediated by NO, but also those evoked by EDHF, have been attenuated (Kimura and Nishio 1999). Nevertheless, unaltered reactivity to endothelium-dependent vasodilators has also been observed (Verbeke et al. 1994, Liu et al. 1997a). The increased oxygen-derived free radical activity and reduced enzymatic antioxidant defence mechanisms have been suggested to result, in part, from the altered arterial function and hypertension in renal failure (Durak et al. 1994, Vaziri et al. 1998). In fact, there is increasing evidence that elevated vascular superoxide production plays a central role in the development of vascular endothelial dysfunction in renal failure (Annuk et al. 2001, Hasdan et al. 2002, Annuk et al. 2003) Nevertheless, increased plasma levels of superoxide dismutase (SOD) and catalase have also been observed in renal failure (Martin-Mateo et al. 1998). It is noteworthy that endothelial dysfunction may also contribute to the development and progression of renal failure, and because endothelial dysfunction is detected at an early phase in the process of renal injury, it appears to be an attractive target for therapy (Rabelink and Koomans 1997).

In recent years several reports have discussed the role of NO synthesis and its inhibition in the development and progression of renal failure. The NO synthesis can be inhibited by analogues of arginine, including endogenous ADMA (Vallance et al. 1992). ADMA is present in normal human plasma, but it accumulates in renal failure (Vallance et al. 1992, Kielstein et al. 1999), suggesting changes in either biosynthesis or excretion (Marescau et al. 1997). In CRF, circulating concentrations of ADMA are thought to rise sufficiently to inhibit NO synthesis, the inhibition of which might contribute to the changes in arterial function and to the hypertension associated with CRF (Vallance et al. 1992, MacAllister et al. 1996, Kielstein et al. 1999). The clinical role of ADMA remains questionable (Anderstam et al. 1997) and a lack of relationship was recently reported between the plasma ADMA and creatinine levels in CRF patiens, that exhibited low total NO production levels (Schmidt and Baylis 2000). In isolated human uremic resistance arteries, impaired endothelial function has been reported, but the sole responsibility of endogenous inhibitors of NO synthase, such as ADMA, for the deficient vasorelaxations remains uncertain (Morris et al. 2001). Moreover, dietary L-arginine supplementation has been suggested to increase NO generation and enhance vasodilatation (Peters and Noble 1996), and to prevent the progression of glomerular sclerosis by ameliorating glomerular capillary hypertension in experimental models of kidney disease (Katoh et al. 1994). Nevertheless, elevated or unaltered plasma levels of L-arginine have been observed in uremic patients even without dietary supplementation (Noris et al. 1993, Kielstein et al. 1999), and both increased and decreased basal NO production have been reported in the vasculature of rats with reduced renal mass (Aiello et al. 1997, Vaziri et al. 1998). The enhanced endothelial NOS expression and the larger amount of NO formed in arteries of reduced renal mass rats can serve as a defence mechanism to limit systemic blood pressure elevation in experimental renal failure (Aiello et al. 1997).

More than 20 years ago Ayachi showed that feeding calcium to the spontaneously hypertensive rat lowered the animal's blood pressure (Ayachi 1979). This finding, along with other similar observations (McCarron et al. 1981, McCarron et al. 1982, Resnick et al. 1983), sparked a burst of activity exploring the relationship between dietary calcium intake and blood pressure regulation in human and animal models of hypertension. While the question of whether elevated calcium intake lowers blood pressure in humans remains controversial (Appel 1997, Taubes 1998), data showing an inverse correlation between calcium intake and blood pressure in several models of hypertension in the rat and dog are highly consistent (McCarron et al. 1985, Peuler et al. 1987, Kageyama et al. 1987, Kotchen et al. 1989, Bukoski et al. 1989, Mäkynen et al. 1996).

3.1 Dietary calcium in experimental hypertension

In animal models of hypertension the antihypertensive effect of a high calcium diet has been consistent (Hatton et al. 1994, Mäkynen et al. 1996), and calcium supplementation has been found to be especially effective in sodium volume-dependent hypertension (Arvola et al. 1993a, Mäkynen et al. 1996). Some experimental studies have suggested that the blood pressure lowering effect of dietary calcium could be due to increased natriuresis following calcium supplementation (Pörsti et al. 1991, Butler et al. 1995). Calcium has been shown to alter sodium reabsorption in the proximal tubule, loop of Henle and cortical collecting tubules (Butler et al. 1995). Calcium has also been shown to blunt the pressure effects of dietary sodium in human hypertension (McCarron 1997). A multitude of mechanisms have been postulated to explain how increased calcium intake lowers blood pressure. Studies have focused on neural, humoral, and renal effects, whereas others have attempted to relate the antihypertensive action of calcium to improved vascular function (Hatton and McCarron 1994, Hatton et al. 1995).

Plausible antihypertensive mechanisms of a high calcium diet include suppression of sympathetic vasoconstrictor tone via modulation of central (Peuler et al. 1987) or peripheral (α_1 -adrenoceptor responsiveness) nervous system activity (Hatton et al. 1993, Hatton and McCarron 1994), improved function of cell membrane Na⁺-K⁺ATPase, reduced voltage-dependent Ca²⁺ entry in arterial smooth muscle (Arvola et al. 1993a), augmented arterial sensitivity to NO, enhanced hyperpolarization of vascular smooth muscle (Mäkynen et al. 1996), and suppression of serum levels of calciotropic hormones, including 1,25(OH)₂ vitamin D₃ and PTH (Bukoski et al. 1995). An interesting link between the intake and metabolism of calcium and the control of arterial tone may be the extracellular Ca²⁺ receptor

in the perivascular sensory nerves, the activation of which causes vasorelaxation via the release of a hyperpolarizing mediator (Bukoski 1998, Ishioka and Bukoski 1999). The Ca²⁺ receptor seems to be present in the perivascular nerve network of a variety of rat tissues, including the mesenteric vasculature, intrarenal arteries and cerebral arteries (Wang and Bukoski 1998).

3.2 Dietary calcium in human hypertension

The concept that dietary calcium could be of significance in blood pressure regulation emerged in the early 1980s (McCarron et al. 1982, McCarron et al. 1984). The reports suggested that low intake of calcium-containing foods was associated with hypertension and that dietary calcium consumption by adults was inversely related to the probability of being hypertensive. Subsequently, there is a wealth of evidence supporting the view that an adequate intake of calcium protects against high blood pressure in humans (Van Leer et al. 1995, McCarron 1997). After 20 years of investigation, a consensus is at hand: a large body of recent data consistently prove the antihypertensive effect of increased intake of calcium (McCarron and Reusser 1999, Vaskonen 2003). The Recommended Dietary Allowance (RDA) for calcium has long been 800 mg/day, but the recognition of the many health benefits of calcium has led to increases in dietary calcium recommendations up to 1500 mg/day, depending on sex and age group (NIH Consensus Panel 1994, Bryant et al. 1999). Dietary calcium intake up to 2000 mg/day is generally regarded as safe (NIH Consensu Panel 1994).

Calcium supplementation has been suggested to lower blood pressure in patients with essential hypertension (Bucher et al. 1996) and dietary calcium seems to even reduce the effect of a high sodium chloride intake on blood pressure (Haddy 1991, McCarron 1997). However, findings from these and other meta-analyses demonstrate considerable heterogeneity in the blood pressure response to increased calcium. This may be explained by several factors, including a threshold effect, consistent with the suggested 600-700 mg/day calcium threshold (McCarron et al. 1984, Zemel 2001). A key factor contributing to the heterogeneity of response is the baseline blood pressure status of the study group. The systolic blood pressure response to calcium supplementation was –3.9 mmHg in the hypertensive patients versus –0.15 mmHg in the normotensive individuals in the six studies that provided separate analyses based on blood pressure status (Bucher et al. 1996, Zemel 2001). Thus, the inclusion of normotensives may have diluted the effect of the dietary intervention.

The heterogeneity in blood pressure response to calcium may also be explained by the intake of other nutrients, interactions among nutrients, and the source of dietary calcium. Indeed, studies that utilized dietary sources of calcium demonstrated approximately twofold greater, and more consistent, effects on blood pressure compared to those that utilized supplements (McCarron and Reusser 1999). The Dietary Approaches to Stop Hypertension (DASH) study compared dietary food patterns and recognized the significance of adding dairy products in the diet of the hypertensive subgroup (Appel et al. 1997). The results from the

DASH study suggested that significant population-wide reductions in coronary heart disease and stroke could be achieved by switching from a typical U.S. diet to the DASH combination diet (fruit and vegetable/dairy products) (Svetky et al. 1999). This would probably be due to achieved reduction in blood pressure but the decreased circulating homocysteine levels might also be of importance (Appel et al. 1997, Appel et al. 2000, Zemel 2001).

4 Calcium metabolism in renal failure

CRF is associated with disturbances of calcium and phosphate metabolism (Drücke 2001). Patients with CRF tend to develop secondary hyperparathyroidism (SH) (Llach 1995), which is characterized by hyperplasia of the parathyroid glands and enhanced synthesis of PTH (Mihai and Farndon 2000, Slatopolsky 2001, Silver et al. 2002). SH develops because phosphate excretion is decreased in renal failure, and elevated plasma phosphate together with reduced synthesis of 1,25-dihydroxyvitamin D₃ (1,25D) contribute to the development of SH (Slatopolsky et al. 2001). Furthermore, the elevated plasma calcium and phosphate levels in SH could play important roles in the uremic cardiovascular disease (Rostand and Drücke 1999).

In early renal failure, deficient 1,25D synthesis is an important factor leading to slightly decreased plasma calcium levels, but reduced expression of vitamin D receptors and the Ca²⁺-sensing receptor may also be present in the parathyroid cells and contribute to hypocalcemia (Korkor 1987, Mihai and Farndon 2000). Later, in advanced renal failure, hyperphosphataemia becomes an important pathogenic factor augmenting the development of SH (Llach and Forero 2001). Increased serum levels of PTH occur even in patients with mild to moderate renal impairment and the three main up-regulators of PTH in man are low serum 1,25D, low ionised calcium and high phosphorus (Slatopolsky et al. 1999). Plasma phosphorus level per se, independent of the levels of Ca²⁺ and 1,25D, is an important stimulator of PTH secretion (Lopez-Hilker et al. 1990), but the extracellular Ca²⁺ significantly contributes to the regulation of plasma PTH levels as well (Drücke 2001, Silver et al. 2002).

The control of PTH gene transcription by 1,25D is mediated by the vitamin D receptor, a protein with high affinity and specificity for the vitamin D hormone (Slatopolsky et al. 1999). The vitamin D receptor expression in the parathyroid glands of CRF patients seems to be markedly reduced (Korkor 1987) and similar results have been reported in experimental uremia as well (Merke et al. 1987). As renal failure progresses, there is a progressive decrease in the number of vitamin D receptors in the parathyroid glands, which makes the parathyroid glands more resistant to 1,25D. Therefore, 1,25D is suggested to be an important regulator of parathyroid cell growth, and in renal failure low levels of 1,25D may allow parathyroid cells to proliferate (Slatopolsky et al. 1999). In experimental animal models of renal failure 1,25D administration has suppressed parathyroid hyperplasia, perhaps through changes in serum calcium (Szabo et al. 1989) and the direct effect on the parathyroid gland mentioned above.

There is evidence for an intrinsic abnormality of the parathyroid glands in uremia that leads to a disordered calcium-regulated PTH secretion and insensitivity to the suppressive effect of calcium on PTH secretion (Brown et al. 1982). The parathyroid glands express a calcium-sensing mechanism via a specific calcium receptor (Brown et al. 1993, Silver et al. 2002), that enables the PTH secretion react to, for instance, hypocalcemia within 1-3 minutes (Slatopolsky et al. 1999). Calcium can also regulate PTH gene transcription (Okazaki et al. 1991) and cell proliferation (Silver et al. 2002). In addition to the impaired control of parathyroid function by calcium in CRF, the frequently observed decrease in dietary calcium intake and the impairment of intestinal calcium absorption due to low 1,25D also contribute to the development of hyperparathyroidism via a tendency towards hypocalcaemia (Drüeke 2001). Oral calcium supplements are used early in CRF to avoid calcium deficiency and control the development of SH (Fournier et al. 1996, Drüeke 2001). Moreover, oral calcium administration reduces hyperphosphataemia by binding phosphate in the intestine, which further helps to manage SH (Drüeke 2001).

Some reports have suggested that high-dose calcium supplements result in an uncontrolled intestinal absorption of unbound calcium and its potential deposition in soft tissues via an increase in Ca x P product (Drücke 2001). An association has been published between the prescribed dose of oral calcium carbonate and arterial wall stiffness (Guérin et al. 2000) and another study reported an increase in coronary artery calcification in young dialysis patients that were given twice as much calcium-containing phosphate binders compared to the controls (Goodman et al. 2000). This risk of inducing extra-skeletal calcifications seems to be further enhanced by vitamin D (Drücke 2001). Moreover, high PTH and phosphate levels predispose to ectopic calcifications (Slatopolsky et al. 2001). Excess of PTH is also associated with elevated blood pressure, and it may directly influence the function of arterial smooth muscle (Rostand and Drücke 1999). Recently, raised PTH and phosphate levels emerged as cardiovascular mortality markers in a 6-year prospective study on Caucasian hemodialysis patients (Marco et al. 2003). Therefore, disturbed calcium-phosphate balance seems to contribute to the cardiovascular pathology in RF (Slatopolsky et al. 2001), and treatment of hyperphosphataemia and decrease of the Ca x P product are cornerstones in the management of advanced stages of SH (Locatelli et al. 2002).

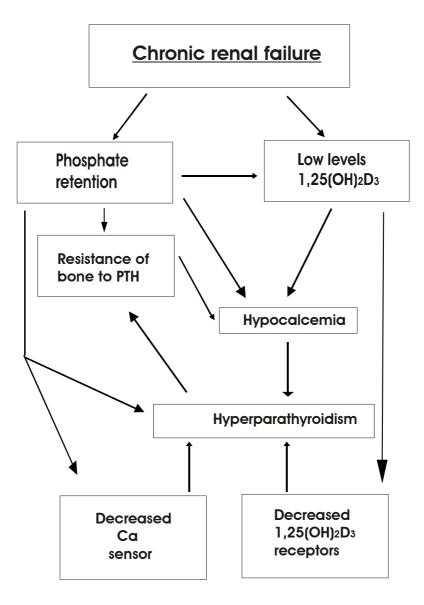


Figure 4. Schematic representation of the factors involved in the pathogenesis of SH.

AIMS OF THE PRESENT STUDY

The objective of the present series of investigations was to examine the control of arterial tone in NO deficiency, NaCl-hypertension and renal failure. The effects of high calcium intake in NO-deficient hypertension and the influences of calcium supplementation and vitamin Dinduced hypercalcemia in NaCl-hypertension on rat conduit artery function were studied. Furthermore, the changes in the tone of resistance arteries were evaluated in renal failure following the treatment of SH by high calcium diet.

The detailed aims were:

1. To examine the influence of high calcium diet on the control of arterial tone in NO deficient Wistar rats.

2. To investigate the effects of increased calcium intake and hypercalcemia induced by oral 1- α -OH vitamin D₃ in NaCl-hypertensive WKY rats.

3. To study the effects of renal failure on the function of the vascular endothelium and arterial smooth muscle in WKY rats.

4. To study the effects of renal failure on the function of the endothelium and smooth muscle of resistance arteries in WKY rats.

5. To examine the influence of the treatment of SH by increased calcium intake on the tone of resistance arteries in Sprague-Dawley rats subjected to 5/6 nephrectomy.

MATERIALS AND METHODS

1 Experimental animals

Normotensive male Wistar and Sprague-Dawley rats were obtained from the colony of the Medical School at the University of Tampere (I, V), whereas WKY rats were obtained from Møllegaard's Breeding Centre, Ejby, Denmark (II) and WKY rats also from M&B A/S, Ry, Denmark (III, IV). The rats were housed two (III, IV, V) or four (I, II) to a cage in a standard animal laboratory room (temperature +22°C, a controlled environmental 12 h light-dark cycle). The studies were approved by the Animal Experimentation Committee of the University of Tampere, and by the Provincial Government of Western Finland, Department of Social Affairs and Health (III, IV, V).

2 Diets and drug treatments

All animals in studies III and IV, and those on control diet in study I received standard laboratory food pellets containing 1.1% calcium, 0.7% sodium chloride and 0.2% magnesium (Ewos, Södertälje, Sweden). The calcium supplementation in study I was accomplished by adding CaCO₃ after which the chow contained 3.0% calcium. In study II the control diet contained 6% sodium chloride and 1% calcium, the calcium supplemented chow contained 6% sodium chloride and 3% calcium, and the vitamin D chow contained 6% sodium chloride, 1% calcium and 10H-D₃ (Etalpha[®]; Lövens, Ballerup, Denmark; vitamin D precursor which is hydroxylated to active 1,25(OH)₂D₃ in the liver) 21-27 µg per kg of chow, i.e. the daily average dose was 1.2 µg per kg of rat. In study V the control chow contained 0.3% calcium, whereas the high calcium diet contained 3.0% calcium. The extra calcium was supplied as the carbonate salt, and otherwise the chows were practically identical (AnalyCen, Lindköping, Sweden).

All the rats were freely provided with tap water excluding the L-NAME-treated animals in study I that received L-NAME (20 mg/kg/day) in bottled drinking fluid. The daily prepared solutions were kept in light-proof bottles. In order to obtain the desired daily L-NAME dose, the concentration in drinking water was adjusted according to 24 h fluid consumption measurements.

3 Blood pressure measurements

The systolic blood pressures of conscious rats restrained in plastic holders were measured indirectly by the tail cuff method at +28°C. All measurements were performed with an IITC Inc. Model 129 Blood Pressure Meter (Woodland Hills, California, USA) equipped with a

photoelectric pulse detector. The blood pressure or each rat was obtained by averaging three reliable recordings.

4 Urine collection and measurement of fluid intake and food consumption

Urine was collected for 24 h individually in metabolic cages where animals had free access to food and water (III, IV, V). Urine volumes were measured and samples stored at -20°C. The consumption of drinking fluid was measured by weighing the bottles after a 24 h period. Food consumption was monitored during periods in special metabolic cages.

5 Blood and heart samples

The rats were anaesthetised by the intraperitoneal administration of urethane (1.3g/kg) and the carotid arteries were cannulated. Blood samples were drawn into chilled tubes on ice containing 2.7 mM ethylenediaminetetraacetic acid (I, II), and into tubes and glass capillaries containing heparin (III, IV, V), after which the samples were centrifuged, and the plasma stored at -70°C until analysis. After exsanguination, the thoracic and abdominal cavities of the animals were opened, the hearts (I, III, IV, V) and the kidneys (V) removed and weighed. The tissue samples were frozen in liquid nitrogen and stored at -70°C until analyses.

6 Biochemical determinations

6.1 Nitrite and nitrate

To measure nitrite and nitrate (Nox) concentrations in plasma and urine (III), vanadium chloride in HCl was used to convert NOx to NO, which was quantified by the ozone-chemiluminescence method (Braman and Hendrix 1989). The samples were first treated with ethanol at -20°C for two hours to precipitate proteins. Then a 20 μ l sample was injected into a cylinder containing saturated VCl₃ solution (0.8 g VCl₃ per 100 ml of 1 M HCl) at 95°C, and NO formed under these reducing conditions was measured by the NOA 280 analyser (Sievers Instruments Inc. Boulder, Colorado, USA) using sodium nitrate as the standard.

6.2. Sodium, potassium, calcium, magnesium, urea nitrogen, phosphate, creatinine, haemoglobin, PTH, 1,25(OH)₂D₃ and proteins

Plasma sodium, potassium (II, III, IV, V) and magnesium (II) concentrations were measured by potentiometric direct dry chemistry, urea nitrogen (III, IV, V) by colorimetric enzymatic dry chemistry, and phosphate (III, IV, V) by colorimetric end-point dry chemistry (Vitros 950 analyzer, Johnson & Johnson Clinical Diagnostics, Rochester, New York, USA). Creatinine (III, IV, V) was determined by the kinetic colorimetric assay according to Jaffe, and plasma proteins (V) were measured by colorimetric measurement according to Biuret (Cobas Integra analyzer, F. Hoffman-La Roche Ltd, Diagnostics Division, Basel, Switzerland). PTH (V) levels were measured by an immunoradiometric assay specific for intact rat PTH (Catalog #50-2000, Immunotopics, San Clemente, California, USA), and vitamin D (V) by radioassay designed for the quantitative determination of 1,25(OH)₂D₃ (competitive protein-binding assay, Catalog #40-6041, Nichols Institute Diagnostics, San Juan Capistrano, California, USA). Ionised calcium (II, III, IV, V) was measured by an ion selective electrode (Ciba Corning 634 Ca²⁺/pH Analyzer, Ciba Corning Diagnostics, Sudbury, UK). Haemoglobin (III, IV, V) was determined by photometric analysis using Technicon cyanide free haemoglobin reagent (Technicon H*2TM, Technicon Instruments Corporation, Tarrytown, New York, USA).

7 Mesenteric arterial responses in vitro

7.1. Arterial preparations and organ bath solutions

The superior mesenteric arteries (I, II, III) were carefully cleaned of adherent connective tissue, excised, and placed on a Petri dish containing physiological salt solution (pH 7.4) of the following composition (mM): NaCl 119.0, NaHCO₃ 25.0, glucose 11.1, KCl 4.7, CaCl₂ 1.6, KH₂PO₄ 1.2 and MgSO₄ 1.2, and aerated with 95 % O₂ and 5 % CO₂. Standard sections of the mesenteric artery (3 mm in length) were cut, beginning 5 mm distally from the mesenteric artery-aorta junction. The endothelium we either left intact or removed by gently rubbing it with a jagged injection needle (Arvola et al. 1992). The rings were placed between stainless steel hooks (diameter 0.3 mm) and mounted in an organ bath chamber (volume 20 ml) in physiological salt solution described above. The small second (IV) or third order (V) branches from the mesenteric arterial bed were carefully excised under a dissecting microscope (Nikon, Japan) and mounted over two 40 µm wires in a small organ bath chamber (volume 5 ml) containing physiological salt solution. The endothelium were left intact or removed by perfusing air through the vascular lumen. The preparations were aerated with 95% O₂ and 5 % CO₂ at +37°C, and rinsed with fresh solutions at least every 20 min, during which time the pH in the baths remained stable. In solutions containing high concentrations of K^+ (20-125mM), NaCl was replaced with KCl on an equimolar basis. In Ca²⁺-free solutions, CaCl₂ was omitted without substitution.

7.2. Arterial contractile and relaxation responses

In studies I, II and III the arterial rings were initially equilibrated for 1 h at +37°C with a resting preload of 1.5 g. The force of contraction was measured with an isometric forcedisplacement transducer and registered on a polygraph (FT 03 transducer and Model 7 E Polygraph; Grass Instrument Co., Quincy, MA, USA). The presence of the functional endothelium in vascular preparations was confirmed by a clear relaxation response to 1 μ M ACh in NA-precontracted arterial rings, and the absence of endothelium by the lack of this response. If any relaxation was observed in the endothelium-denuded rings, the endothelium was further rubbed. In studies IV and V a Mulvany multimyograph Model 610A (J.P. Trading, Aarhus, Denmark) was employed for studies with vascular preparations. In this system the isometric micromyographs consist of two jaws, one of which is connected to a length displacement device and the other to a force transducer linked to a computer with Myodag software (J.P. Trading). The small arterial rings were placed over two thin wires, each of which was attached to one of the myograph jaws. Normalisation of the vascular preparations was then performed so that the internal diameter of the vessel was set at 90 % of that obtained when exposed to an intraluminal pressure of 100 mmHg in the relaxed state (Mulvany and Halpern 1977). The presence of intact endothelium in the vascular preparations was confirmed by a clear relaxation to 1 µM ACh in NA-precontracted rings, and the absence of endothelium by the complete lack of this response.

Agonist-induced contractions. The contractions of the endothelium-intact preparations to NA were studied in the absence (I, II, III, IV, V) and presence of L-NAME (0.1 mM) (I, III), and in the presence of diclofenac (3 μ M) (II) or diclofenac plus L-NAME (I, II, III). In study III, the contractions to NA were also elicited in the presence of L-Arginine (1 mM). The contractions elicited by ET-1 were investigated in the endothelium-denuted preparations in study V.

Depolarization-induced contractions. The concentration-response curves of the endothelium-denuded rings to KCl were determined in the absence (II, III, IV, V) and presence of L-NAME (0.1 mM) (I).

 Ca^{2+} contractions. The contractile responses of the endothelium-denuded rings to cumulative addition of Ca²⁺ to the organ bath chamber after precontraction with KCl (125 mM) in Ca²⁺-free buffer in the presence of L-NAME (0.1 mM) (I) or phentolamine (1 μ M) and atenolol (10 μ M) (III) were studied. Thereafter, the effect of nifedipine (0.5 nM) on these responses was examined (III).

Endothelium-dependent relaxations to ACh and adenosine 5'-diphosphate (ADP). Mesenteric arterial relaxations were studied in response to ADP (II) and ACh (II, III, IV, V) in rings precontracted with NA (1 μ M in II, III; 3 μ M in IV and 5 μ M in V). The ACh-induced relaxations after NA-precontraction were also elicited in the presence of diclofenac

(II), L-NAME (I, III, IV, V), diclofenac and L-NAME (I, II, III, IV, V), diclofenac, L-NAME and tetraethylammonium (1 mM) (II), diclofenac, L-NAME, and apamin (50 nM) plus charybdotoxin (0.1 μ M) (III, IV, V). The responses to ACh were further studied in the presence of SOD (50 U/ml) (III); L-NAME and SOD (I); SOD plus catalase (100 U/ml) (III) and SOD, L-NAME and catalase (I). The ACh-induced relaxations were also examined in the presence of L- arginine (1 mM) (III). Furthermore, the relaxations to ACh and ADP were investigated in rings precontracted with KCl (50 mM) in the absence and presence of L-NAME and diclofenac (I).

Endothelium-independent relaxations to sodium nitroprusside (SNP), isoprenaline, cromakalim, levcromakalim and 11,12-epoxyeicosatrienoic acid (EET). The relaxation responses of NA-precontracted (I, II, III, IV, V) and KCl-precontracted (I, II) endotheliumdenuded rings to SNP were examined. The vasorelaxations elicited by isoprenaline and cromakalim (I, II, III, IV) or levcromakalim (V) were studied in endothelium-denuded rings precontracted with NA (I, II, III, IV, V). Moreover, responses to isoprenaline were also studied in rings precontracted with KCl (II). In study I, the endothelium-independent responses to SNP, isoprenaline and cromakalim were studied in the presence of L-NAME when testing vessels from L-NAME-treated animals. In study V, the relaxation responses to EET were examined after precontraction with NA.

8 Morphological studies

In study II the rat aortas were fixed in 4 % formaldehyde, embedded in paraffin, and a 5 μ m transverse section was cut and stained with hematoxylin and eosin. Fibrosis, inflammation, calcification and wall thickening were scored. The apoptosis of aortic smooth muscle cells was measured by the in situ end-labelling technique (ApopTaq-kit, Oncor Inc., Gaithersburg, Maryland, USA). Prostates from castrated and non-castrated rats were used as controls in apoptosis staining, and the results were scored in a blinded fashion using an Olympus BX50 microscope (Olympus, Tokyo, Japan). One hundred cells were counted from 10 fields on each slide (at 200x magnification) and results expressed as percentage of apoptotic cells.

In study IV the small vascular rings from the second or third order branches of the rat mesenteric artery were mounted on the Mulvany myograph Model 610. The myograph together with the Myodaq software determine and record the lumen diameter of each preparation during the standard normalisation process which sets internal diameter of the vessel at 90 % of that obtained when the intraluminal pressure is set at 100 mmHg. In study V the morphology of small arteries was examined with a pressure myograph (Living Systems Instrumentation Inc., Burlington, Vermont, USA), and the development of myogenic tone was inhibited by Ca^{2+} -free solution containing 30 mmol/l EDTA (Suo et al. 2002).

9 Compounds

The following drugs and chemicals were used: ACh chloride, apamin, catalase, charybdotoxin, cromakalim, 11,12-epoxyeicosatrienoic acid, isoprenaline hydrochloride, NA bitartrate, L-NAME hydrochloride, SOD, tetraethylammonium chloride (Sigma Chemical Co., St. Louis, Missouri, USA), levcromakalim (SmithKline Beecham AB, West Sussex, U.K.), atenolol (Leiras Pharmaceutical, Turku, Finland), ketamine (Parke-Davis Scandinavia AB, Solna, Sweden), cefuroxim, diazepam, nifedipine (Orion Pharma Ltd., Espoo, Finland), metronidazole (B. Braun AG, Melsungen, Germany), buprenorphine (Reckitt & Colman, Hull, U.K.), SNP, NA hydrogentartrate (Fluka Chemie AG, Buchs SG, Switzerland), diclofenac, phentolamine (Voltaren[®] injection solution, Ciba-Geigy, Basle, Switzerland), sodium salt of adenosine 5'-diphosphate (Boehringer Mannheim GmbH, Germany), and 10H-D₃ (Etalpha[®], Lövens, Ballerup, Denmark). The stock solutions of the compounds used in the *in vitro* studies were made by dissolving the compounds in distilled water, with the exception of cromakalim, levcromakalim and nifedipine (in 50 % ethanol), and EET (in 99% ethanol). Drinking fluids containing L-NAME hydrochloride were made by dissolving the compound in tap water. All solutions were freshly prepared before use and protected from light. The chemicals used in the preparation of physiological salt solution were of highest grade available and obtained from E. Merck AG (Darmstadt, Germany).

10 Analysis of results

The statistical analysis was performed using one-way or two-way analysis of variance (ANOVA) supported by Bonferroni test or by two-tailed t-test when carrying out pairwise comparisons between the study groups. ANOVA for repeated measurements was applied for data consisting of repeated observations at successive time points. Spearman's correlation coefficient was used in the correlation analyses. All results were expressed as mean \pm SEM. The data were analysed with BMDP Statistical Software version PC90 (Los Angeles, California, USA).

Study	Treatment Rats Vessel	E+ relaxations (precontraction)	E- relaxations (precontraction)	Contractions
Experimen	tal hypertension			
I	L-NAME hypertensive Wistar rats Mesenteric artery	ACh (NA) + L-NAME and diclofenac ACh(KCl) ACh (NA) + SOD + catalase	Isoprenaline (NA) Cromakalim (NA) Nitroprusside (NA and KCl)	NA + L-NAME and diclofenac KCl Calcium
П	NaClACh (NA)hypertensive+ diclofenacWKY rats+ L-NAMEMesenteric artery+ TEAACh(KCl)+ L-NAMEADP (NA)ADP (KCl)+ L-NAME		Isoprenaline (NA and KCl) Cromakalim (NA) Nitroprusside (NA and KCl)	NA + diclofenac + L-NAME KCl
Renal failu	re			
Ш	5/6 nephrectomized WKY rats Mesenteric artery	WKY rats + L-NAME		NA + L-NAME + diclofenac KCl Calcium + nifedipine
IV	5/6 nephrectomized WKY rats Small mesenteric artery	ACh (NA) + L-NAME + diclofenac + apamin and charybdotoxin	Isoprenaline (NA) Cromakalim (NA) Nitroprusside (NA)	NA KCl
V	5/6 nephrectomized Sprague-Dawley rats Small mesenteric artery	ACh (NA) + L-NAME + diclofenac + apamin and charybdotoxin	Isoprenaline (NA) Cromakalim (NA) Nitroprusside (NA) EET (NA)	NA KCl ET-1

Table 1. Summary of the experimental design of the studies on arterial reactivity.

ACh, acetylcholine; ADP, adenosine 5'diphosphate; E+, endothelium-dependent; E-, endothelium-independent; EET, 11,12-epoxyeicosatrienoic acid; ET-1, endothelin-1; L-NAME, N_G-nitro-L-arginine methyl ester; NA, noradrenaline; SOD, superoxide dismutase; TEA, tetraethylammonium.

RESULTS

1 Blood pressure, arterial morphology and apoptosis, heart weight, total renal mass, drinking fluid and urine volumes

Blood pressure. The systolic blood pressures of untreated Wistar (I) and WKY (II) rats were 139 mmHg and 154 mmHg, respectively, when measured at the end of the follow up periods. The long-term administration of L-NAME resulted in the elevation of blood pressure up to 198 mmHg in Wistar rats (I) and the NaCl-diet elevated the blood pressure to 184 mmHg (II), whereas calcium supplementation clearly attenuated (I) or completely prevented (II) the increase in blood pressure, while 10H-D₃-treatment also attenuated the development of NaCl-hypertension (II). Concomitant calcium supplementation during the 10H-D₃-treatment did not affect the blood pressure in NaCl-hypertension (II). The CRF in WKY (III, IV) rats was not associated with elevation of blood pressure, nor did the high calcium diet affect the blood pressure in CRF (V). When analysed by two-way ANOVA, a small but significant increase in blood pressure was uncovered in Sprague-Dawley (V) rats with renal failure when compared with sham-operated controls (i.e. both NTX groups compared with both sham-operated groups).

Arterial morphology and apoptosis. No differences were observed in the smooth muscle apoptosis in the aortic wall in study II. Signs of mild fibrosis with some perivascular lymphocytes were observed in 20% and 27% of the aortas in the Control and Calcium groups of study II, respectively, while calcifications in the adventitia or media with inflammatory cells were observed in 50% and 57% of the aortic sections from the 1OH-D₃ and 1OH-D₃+Calcium groups in study II, respectively. The CRF was not associated with alterations in mesenteric arterial morphology (unpublished observation in study III). The lumen diameters of the small mesenteric arteries in studies IV and V were corresponding between the groups, and no difference in the wall thickness or in the cross-sectional area was detected in study V. However, the wall to lumen ratio of isolated perfused third order mesenteric artery brances in study V was increased in the NTX and NTX+calcium groups when compared with the Sham group.

Heart weight and total renal mass. The heart-body weight ratios were comparable in L-NAME hypertensive and normotensive Wistar rats while calcium supplementation was without significant effect on the relative heart weights (I) In CRF the heart-body weight ratios remained comparable in WKY (III, IV) and Sprague-Dawley (V) rats and high calcium intake did not influence the relative heart weights in CRF (V). The total renal mass following the subtotal nephrectomy and the development of CRF was approximately 70% in the nephrectomized animals compared with sham-operated controls in study V.

Drinking fluid and urine volumes. At the end of the study, the intake of drinking fluid

and the output of urine were higher in the rats with CRF when compared with the control animals (III, IV, V).

2 Plasma sodium, potassium, calcium, 1,25(OH)₂D₃, magnesium, urea nitrogen, creatinine, PTH, phosphate, proteins, haemoglobin and NOx

1OH-D₃-administration increased plasma total Ca^{2+} concentrations, while plasma Na⁺, K⁺ and Mg²⁺ were similar in all study groups in study II. In rats with CRF, the plasma creatinine and urea nitrogen values were increased, while plasma sodium, haemoglobin and calcium concentrations were decreased when compared with control WKY rats (III, IV). Renal failure did not influence plasma potassium, phosphate, pH (III, IV) or NOx (III). In study V, CRF elevated the plasma urea nitrogen, creatinine, PTH and phosphate concentrations and decreased plasma ionized calcium, 1,25(OH)₂D₃, proteins and haemoglobin. The high calcium intake significantly lowered the plasma PTH and phosphate concentrations and elevated the ionized calcium (V).

3 Control of arterial tone in vitro

3.1. Arterial tone in L-NAME and NaCl hypertension and the influence of calcium supplementation and vitamin D-induced hypercalceamia

3.1.1 Arterial contractile responses

Both in the presence of L-NAME and in the presence of L-NAME and diclofenac, the vascular rings of the study groups showed comparable sensitivity to NA (I). In the presence of L-NAME, the rings of the control and L-NAME groups showed similar contractile sensitivity to KCl, whereas the sensitivity was somewhat higher in the calcium supplemented groups (I). Arterial sensitivity to the addition of Ca^{2+} during depolarization with 125 mM KCl was similar in the calcium and control groups but higher in the calcium group when compared with the L-NAME and calcium+L-NAME groups (I).

In study II, maximal wall tension and sensitivity to NA, in the absence and presence of diclofenac and L-NAME, was corresponding in the control and low-NaCl groups, and the high Ca²⁺ diet was without significant effects on the NA-elicited contractions. In contrast, the 1OH-D₃ treatment markedly decreased maximal wall tension in response NA without changing the sensitivity (II). In the vascular rings of both 1OH-D₃-treated groups lower maximal wall tension response to NA was also observed after pretreatment with L-NAME and diclofenac, while sensitivity remained comparable in all groups. Maximal wall tension in response to KCl was also reduced in both of the 1OH-D₃-treated groups when compared with

the other groups (II), yet the sensitivity to KCl was comparable in all the study groups (II).

3.1.2 Arterial relaxation responses

Endothelium-independent relaxations. The relaxations of endothelium-denuded NAprecontracted rings to SNP, isoprenaline and cromakalim, were impaired in L-NAME hypertensive rats when compared with normotensive controls (I). In addition, when hyperpolarization of smooth muscle was prevented by precontractions with KCl, the relaxations to SNP were still impaired in L-NAME rats. Furthermore, calcium supplementation clearly improved the vasodilatations to SNP, isoprenaline and cromakalim in L-NAME-treated animals (I). Even the control rats exhibited improved vasorelaxation to SNP after precontraction with KCl and to isoprenaline after NA-precontraction following calcium supplementation (I).

The high NaCl intake impaired the vasodilatations to SNP, isoprenaline and cromakalim in NA-precontracted arterial rings when compared with the low-NaCl group (II). High calcium diet enhanced these responses, and the relaxations in the calcium group were not different from those in the low-NaCl group (II). The chronic 1OH-D₃-induced hypercalcemia, without or with calcium intake, had no effects on the endothelium- independent relaxations, except for the response to 0.33 μ M cromakalim was reduced in both 1OH-D₃-treated groups when compared with the control rats (II).

Endothelium-dependent relaxations. The relaxations induced by ACh in NAprecontracted arterial rings in the presence of L-NAME were markedly impaired in the L-NAME treated rats when compared with the control rats, while these responses were significantly improved by calcium supplementation in both of these groups (I). The addition of diclofenac to the organ bath chamber improved the relaxations to ACh in the L-NAME and control groups but not in the calcium supplemented groups (I). Diclofenac also abolished the difference in the ACh response between the control and the calcium supplemented groups, whereas the relaxations still remained impaired in the L-NAME group when compared with the others (I). The responses to ACh were almost completely abolished in all groups when induced in KCl-precontracted rings in the presence of L-NAME and diclofenac (I). When L-NAME and SOD were added to the organ bath, the relaxations to ACh were enhanced in the L-NAME group, but the responses remained impaired when compared with the other groups (I). In addition, SOD augmented the relaxation to ACh in the control group, whereby the difference in response to ACh between the control group and the calcium supplemented groups was abrogated (I). The further addition of catalase had no effect on the relaxation to ACh in any of the study groups.

In study II, the relaxations induced by ACh and ADP in NA-precontracted arterial rings were impaired in the control group when compared with the low-NaCl group, while these responses were improved by calcium supplementation so that they did not differ from those of the low-NaCl group. The 1OH-D₃-induced chronic hypercalcemia, without or with concomitant calcium supplementation, had no influence on the endothelium-dependent vasorelaxation in NaCl-hypertensive rats (II). The addition of diclofenac had no significant effect on the relaxations to ACh, but the addition of L-NAME reduced the responses to ACh, yet clear relaxations were still observed in the calcium and low-NaCl groups (II). The addition of tetraethylammonium reduced the L-NAME and diclofenac-resistant relaxation to higher concentrations of ACh by 30-40 % in the low-NaCl and calcium groups, and almost completely abolished the response to ACh in the control, 10H-D₃ and 10H-D₃+calcium groups (II). When the relaxations to ACh and ADP were elicited after precontractions with KCl, the responses were lesser than after precontractions with NA, and the relaxations were more pronounced in the calcium and low-NaCl groups than in the other groups. The responses to ACh and ADP in KCl-precontracted rings were abolished in the presence of L-NAME.

3.2 Effects of renal failure on the control of macro- and microvessel tone

3.2.1 Arterial contractile responses

Vasoconstrictor responses. CRF did not alter the arterial contractile sensitivity to NA or KCl in large mesenteric arterial rings (III) nor in the resistance arterial rings (IV, V). The maximal contractions to NA in the absence (III, IV, V) and presence (III) of L-NAME were also comparable between renal failure and control rats both in conduit vessels (III) and in resistance arteries (IV, V). However, in the presence of L-NAME and diclofenac the maximal contractile force generation induced by NA was higher in the conduit arterial rings of the renal failure rats, and the maximal contractions to KCl were also more pronounced in the renal failure rats when compared with the control rats (III).

3.2.2 Arterial relaxation responses

Endothelium-independent relaxations. CRF did not modulate arterial relaxations to SNP in large mesenteric arteries (III) or small arteries (IV). However, the relaxations to isoprenaline and cromakalim were impaired in large arteries of rats with CRF (III). The vasorelaxation to isoprenaline was also attenuated in the resistance vessels of CRF rats but the relaxation responses to cromakalim remained similar between the study groups (IV).

Endothelium-dependent relaxations. The relaxations induced by higher concentrations of ACh (1-10 μ M) in NA-precontracted conduit arterial rings were impaired in the renal failure rats when compared with the control rats (III). L-NAME diminished the relaxations in both study groups, but the attenuation was more pronounced in the renal failure group than in

the control group (III). Diclofenac was without significant effects on ACh-induced relaxations in both groups (III). The addition of apamin and charybdotoxin further reduced the relaxations to ACh in the control rats but not in the renal failure rats, thereby abolishing the difference in the remaining relaxation to ACh (III).

The relaxations induced by ACh in endothelium-intact NA-precontracted mesenteric small arterial rings were similar in the renal failure rats when compared with the control rats (IV). In the presence of L-NAME the relaxations to ACh were reduced, but the responses remained comparable between the study groups (IV). Diclofenac was without effect on the ACh-induced relaxations (IV). In contrast, the further addition of apamin and charybdotoxin clearly reduced the relaxations in both study groups (IV), but still no significant difference was observed in the small arterial reactivity between the renal failure and control groups (IV).

3.3 Influence of phosphate binding by high calcium diet on the control of microvessel tone in renal failure

3.3.1 Arterial contractile responses

Vasoconstrictor responses. CRF did not alter the arterial contractile sensitivity to NA or KCl in the resistance arterial rings. The maximal contractions to NA were also comparable between renal failure and control rats. The resistance arteries of the renal failure rats on high calcium diet exhibited somewhat higher sensitivity to KCl than the arteries from the control group. The sensitivity and maximal wall tension induced by ET-1 remained comparable between all study groups in the resistance arterial rings.

3.3.2 Arterial relaxation responses

Endothelium-independent relaxations. CRF did not modulate arterial relaxations to SNP or isoprenaline in the small arteries. The resistance vessels of CRF rats were less sensitive to levcromakalim and EET, while these impairments in endothelium-independent vasorelaxation were normalized by high calcium intake. *Endothelium-dependent relaxations.* The relaxations induced by ACh in NA-precontracted small artery preparations were impaired in the NTX group, whereas the groups on the high calcium diet did not differ from the sham-operated control animals. In the presence of L-NAME, the relaxations to ACh were diminished in all groups, but the responses remained less marked in the NTX group when compared with the others. Further addition of diclofenac was without effects on the responses to ACh. The addition of apamin and charybdotoxin clearly reduced the relaxations in all study groups, and the difference in the remaining response to ACh was abolished. The reduction in relaxation by apamin and charybdotoxin was smaller in the NTX group when compared to other groups.

Variable	NO deficiency	+Ca ²⁺	NaCl Hypertension	+Ca ²⁺	+10H-D ₃	+Ca ²⁺ +1OH-D ₃
E+ relaxations (precontraction)					- 5	
ADP (NA and KCl)			\downarrow	↑	\leftrightarrow	\leftrightarrow
Acetylcholine (NA)			\downarrow	↑	\leftrightarrow	\leftrightarrow
+ L-NAME	\downarrow	\uparrow				
+ diclofenac			\downarrow	\uparrow	\leftrightarrow	\leftrightarrow
+ L-NAME and diclofenac	\downarrow	↑	\downarrow	1	\leftrightarrow	\leftrightarrow
+ L-NAME, diclofenac, TEA			\downarrow	Ť	\leftrightarrow	\leftrightarrow
+ L-NAME and SOD	\downarrow	1				
Acetylcholine (KCl)			\downarrow	↑	\leftrightarrow	\leftrightarrow
+ diclofenac and L-NAME	\leftrightarrow	\leftrightarrow				
E- relaxations (precontraction)						
Nitroprusside (NA)	\downarrow	\uparrow	\downarrow	↑	\leftrightarrow	\leftrightarrow
Nitroprusside (KCl)	\downarrow	\uparrow	\downarrow	↑	\leftrightarrow	\leftrightarrow
Isoprenaline (NA)	\downarrow	\uparrow	\downarrow	\uparrow	\leftrightarrow	\leftrightarrow
Isoprenaline (KCl)	\leftrightarrow	\leftrightarrow	\leftrightarrow	\leftrightarrow	\leftrightarrow	\leftrightarrow
Cromakalim (NA)	\downarrow	\uparrow	\downarrow	\uparrow	\downarrow	\downarrow

Table 2. Summary of the alterations in arterial relaxations in hypertensive rats after high calcium intake and vitamin D -induced hypercalcemia compared with untreated hypertensive controls.

ADP, adenosine 5'diphosphate; E+, endothelium-dependent; E-,

endothelium-independent; L-NAME, N_G-nitro-L-arginine methyl ester; NA, noradrenaline; SOD, superoxide dismutase; TEA, tetraethylammonium. \uparrow , \downarrow and \leftrightarrow indicate an increase, reduction and no change when compared with the corresponding control group, respectively.

Variable	Renal failure 6 weeks		Renal failure 12 weeks	
	large artery	small artery	small artery	small artery +Ca ²⁺
E+ relaxations (precontraction)				
Acetylcholine (NA)	\downarrow	\leftrightarrow	\downarrow	\uparrow
+ L-NAME	\downarrow	\leftrightarrow	\downarrow	\uparrow
+ L-NAME and diclofenac	\downarrow	\leftrightarrow	\downarrow	\uparrow
+ L-NAME, diclofenac, AP and CHBD	\leftrightarrow	\leftrightarrow	\leftrightarrow	\leftrightarrow
+ SOD	\downarrow			
E- relaxations (precontraction)				
EET (NA)			\downarrow	\uparrow
Nitroprusside (NA)	\leftrightarrow	\leftrightarrow	\leftrightarrow	\leftrightarrow
Isoprenaline (NA)	\downarrow	\downarrow	\leftrightarrow	\leftrightarrow
Cromakalim (NA)	\downarrow	\leftrightarrow	$\downarrow *$	^∗

Table 3. Summary of the alterations in arterial relaxations in renal failure rats when compared with sham-operated controls and the effect of high calcium intake.

ADP, adenosine 5'diphosphate; AP, apamin; CHBD, charybdotoxin; E+, endothelium-independent; EET, 11,12-epoxyeicosatrienoic acid; \uparrow , \downarrow and \leftrightarrow indicate an increase, reduction and no change when compared with the corresponding control group, respectively. * levcromakalim.

DISCUSSION

The present investigation examined the effects of dietary calcium on conduit artery responses in L-NAME and NaCl hypertension as well as the effect of vitamin D induced hypercalcaemia on conduit artery responses in NaCl hypertension. The influence of CRF on the tone of large and small mesenteric arteries was evaluated. Furthermore, the effect of treating SH by dietary calcium on the resistance vessel tone was studied.

1 Experimental models of the study

The two models of experimental hypertension employed were chronic inhibition of NOS (Baylis et al. 1992) and NaCl-induced hypertension. NO deficiency presents an interesting model of hypertension, since the endothelial production of NO is essential for the maintanence of normal blood pressure (Huang et al. 1995), and several disease states including essential hypertension have been associated with defects in the production or action of NO (Moncada and Higgs 1993). In this study, in agreement with previous experiments, oral administration of L-NAME resulted in a marked hypertension (Ribeiro et al. 1992), which reached its maximum within four weeks, whereas calcium supplementation reduced the elevation of blood pressure significantly. Moreover, dietary NaCl administration also resulted in elevation of blood pressure, which was completely prevented by calcium supplementation. The 1OH-D₃-treatment also attenuated the development of NaCl-hypertension, and although this could be in part due to the reduced arterial constrictor responses the impaired growth of the 1OH-D₃-treated rats may have caused the decrease in blood pressure. The significant impairment in growth of 1OH-D₃-treated animals may have even been a toxemic effect but this remains unclear since the vitamin D levels in plasma were not measured. There are previous reports of chronic vitamin D-induced hypercalcaemia impairing growth in rats (Bukoski et al 1993). The third experimental model in this study was the subtotal (5/6) nephrectomy in rats, which did not result in elevation of blood pressure in WKY but a small increase was observed in Sprague-Dawley rats.

2 Cardiovascular remodelling and morphology in experimental hypertension and renal failure

Cardiac hypertrophy is the primary chronic compensatory mechanism to increased haemodynamic overload in hypertension (Mosterd et al. 1999). Therefore, L-NAME administration and the consequent NOS-inhibition -induced hypertension would be expected to cause cardiac hypertrophy. However, the heart weight-body weight ratios did not differ

between L-NAME hypertensive and normotensive control rats. This finding agrees with reports, which suggest that this is explained by the negative metabolic effects of L-NAME on protein synthesis (Arnal et al. 1993, Bartunek et al. 2000) and the subsequent inhibition of cardiovascular growth processes (Li et al. 1996, Banting et al. 1997). Calcium supplementation had no effect on the relative heart weights in L-NAME hypertension when compared with the control rats. Since $1,25(OH)_2D_3$ and intracellular Ca^{2+} are known regulators of apoptosis (Van den Bemd et al. 2000), we examined whether there would be alterations in NaCl hypertension following the treatments with $1,25(OH)_2D_3$ and Ca^{2+} . However, no differences were found in the apoptosis of aortic smooth muscle cells, but approximately half of the aortic cross sections showed signs of calcifications after the $1,25(OH)_2D_3$ treatment. This may have played a role in the reduced vasoconstrictor responses of mesenteric arteries after $1,25(OH)_2D_3$ -induced hypercalcemia in NaCl hypertension.

Patients with CRF have been characterized by abnormal elastic properties of large arteries, reflected as decreased distensibility and compliance (Barenbrock et al. 1994, London et al. 1996). The increased stiffness of the conduit arteries has even been seen in the absence of structural changes (Mourad et al. 1997). Consistent with this, experimental renal failure in our study was not associated with morphological changes in large mesenteric arteries. Moreover, no changes in cardiac weight were observed in renal failure rats compared with control animals. However, we found that the small arteries of rats that were investigated 12 weeks following the subtotal nephrectomy featured increased wall to lumen ratio, which was not affected by increased calcium intake. Since the cross-sectional area of arterial wall was not increased, the observed change in vascular morphology in CRF rats is compatible with eutrophic inward remodelling (Mulvany 1999). The vascular wall to lumen ratio exhibits the ability of the vessel to contract against intravascular pressure, while the cross-sectional area indicates the amount of material within the vascular wall, and provides information of vascular growth (Mulvany 1999). Therefore, the results concerning small arterial relaxation following increased calcium intake in renal failure indicated that high calcium diet improved vasorelaxation, although the structure of the resistance vessels was not corrected.

3 Influence of dietary calcium and vitamin D-induced hypercalcemia on arterial contractions in experimental hypertension and renal failure

There is a multitude of approaches that can be applied to study the vascular constrictor responses. The contractile force can be related to segment length or weight, media cross sectional area or lumen diameter, and thus the results depend on the experimental method (Mulvany et al. 1991, Arvola et al. 1993b, Bennet et al. 1996). Various approaches to report arterial contractions were also applied in different original communications of this study. In study I the contractile forces were expressed as the actual forces that were recorded (g),

whereas in studies II, III, IV and V the contractile forces were related to the arterial segment length and expressed as wall tensions (mN/mm). Thus, the numerical values obtained from the studies are incomparable as such. Moreover, the present literature does not reach a consensus regarding the method of the expression of arterial contractile forces.

In the present study, the maximal large artery contractile tension generated by NA or KCl was markedly reduced in the 1OH-D₃-treated groups in the absence and presence of NO synthase and COX inhibition, and also in endothelium-denuded preparations. Thus, alterations in the synthesis or release of NO and prostanoids did not explain these changes, yet the underlying abnormality may have located in the arterial smooth muscle following the $1,25(OH)_2D_3$ -treatment in NaCl-induced hypertension. Previously, contradictory results on the effects of vitamin D on arterial contractile properties in rats have been published, but some reports have described an enhanced contractile reponse to NA in isolated rat mesenteric arteries following the treatment with $1,25(OH)_2D_3$ for short periods of 3-7 days (Hatton et al. 1994, Bian et al. 1996).

Chronic L-NAME hypertension did not affect the contractile responses of the isolated conduit arteries of Wistar rats, whereas calcium supplementation slightly increased the sensitivity of the arterial rings to KCl. However, the deviation was small. In CRF, the maximal contraction force of the large mesenteric arteries to KCl in endothelium-denuded rings and to NA in endothelium-intact rings were increased. The maximal wall tensions in the resistance arteries to NA, KCl and ET-1 were unaffected by renal failure. Morever, the high calcium intake in renal failure was without significant effect on these responses. Thus, the present results suggest that calcium supplementation does not have a significant effect on the arterial contractile responses, whereas 1,25(OH)₂D₃-treatment markedly reduces the maximal tension of the rat large mesenteric artery in NaCl-hypertension. This reduction in vascular contractility could be due to the observed vascular calcification following 1,25(OH)₂D₃-treatment.

In the experimental hypertension models of this study some differences in vasoconstrictor sensitivity following the present treatments were observed. Calcium supplementation slightly increased the vascular sensitivity to KCl in both the normotensive and L-NAME-hypertensive groups, and to Ca^{2+} in the normotensive group. In CRF, the large artery sensitivity to NA, KCl and Ca^{2+} was not altered, nor was the resistance vessel sensitivity to NA, KCl and ET-1. Taken together, no marked changes in the arterial sensitivity to constrictor agonists were recorded in the experimental models of the present study.

4 Influence of dietary calcium and vitamin D-induced hypercalcemia on arterial relaxation in experimental hypertension

ACh induces relaxation in arterial smooth muscle by releasing dilatory factors from the

vascular endothelium, the most prominent of these being NO, PGI_2 and EDHF (Busse and Fleming 1993). The relaxations of arterial rings to ACh were impaired in the L-NAME hypertensive and in NaCl hypertensive rats, and effectively improved by calcium supplementation. In contrast, the 1OH-D₃ treatment, without or with high calcium intake, did not have any influences on the endothelium-dependent relaxations in NaCl hypertension. The finding concerning the impaired ACh-induced relaxation in L-NAME hypertension is not new (Küng et al. 1995), but the influence of high calcium intake on arterial relaxations in L-NAME hypertension has not been previously reported. Since calcium supplementation also significantly reduced the elevated blood pressure of the L-NAME rats and completely prevented the NaCl-induced hypertension, the beneficial effect on the vasculature was at least partly mediated via the decrease in blood pressure. Although not directly determined in the present studies, high calcium intake may have also affected the natriuresis of the hypertensive animals. Previously, such an effect has been reported in numerous studies (Pörsti et al. 1991, Hatton and McCarron 1994, Butler et al. 1995). Moreover, the 1OH-D₃-treatment also attenuated the development of NaCl-hypertension.

The chemical antagonism between superoxide anions and NO is an important modulator of vascular tone. In addition, superoxide can inhibit the vascular synthesis of PGI₂ without affecting that of the vasoconstrictor thromboxane A₂ (Katusic 1996). Therefore, increased cardiovascular production of superoxide could contribute to the development of hypertension. In the present L-NAME study, the relaxations to ACh were examined after the addition of the superoxide anion scavenger SOD to the organ bath. The reduction of blood pressure by calcium supplementation may have reduced the production of superoxide in the arteries of L-NAME-treated rats, because the addition of SOD enhanced the relaxations to ACh in the L-NAME group but not in the calcium+L-NAME group. Moreover, SOD also enhanced the relaxations to ACh in the control group but was without effect in the calcium group, suggesting that calcium supplementation reduced the vascular production of superoxide also in the normotensive rats. The present indirectly detected result of reduced superoxide production following calcium supplementation has not been previously reported.

The attenuated vasorelaxation in hypertension could partly be explained by enhanced release of EDCFs (Lüscher and Vanhoutte 1986, Küng et al. 1995). Previously, increased production of vasoconstrictor prostanoids has been shown to potentially contribute to the impaired vasodilatation in L-NAME hypertensive rats (Küng et al. 1995). In the present study, the COX inhibitor diclofenac enhanced the relaxations to ACh in L-NAME hypertensive but not in NaCl hypertensive rats, suggesting that there is an imbalance in the production of vasoconstrictor and vasodilator prostanoids in the vessels of L-NAME hypertensive rats, which favours vasoconstriction. Calcium supplementation appeared to reduce the production of these factors in L-NAME rats, because in the responses to ACh after diclofenac no significant changes were detected in the supplemented groups. Furthermore,

diclofenac enchanced the relaxations to ACh in the control group but was without effect in the calcium group, suggesting that calcium supplementation decreased the release of vasoconstrictor prostanoids also in the control rats. In addition, decreased arterial superoxide production may also have contributed to the enhanced endothelium-mediated vasodilatation after diclofenac administration, because COX is a significant source of superoxide (Katusic 1996). In the present model of NaCl hypertension, the administration of diclofenac to the organ bath was without significant effect on the responses to ACh, whereby the role of COX-derived contractile substances seemed negligible in the endothelium-mediated relaxations.

In the L-NAME hypertension study, we had to recognize the phenomenom that the inhibitory effect of orally administered L-NAME on ACh-induced relaxations is known to decline during successive responses in isolated arterial preparations from L-NAME treated rats (Deng et al. 1993). We found that 100 μ M L-NAME was needed in the organ bath to prevent this, and therefore the ACh-induced relaxations were performed in the presence of 100 μ M L-NAME.

In the presence of diclofenac, the inhibition of NO synthase by L-NAME clearly reduced the responses to ACh in all groups of the NaCl-hypertension study, indicating important contribution of NO to the relaxations. However, the calcium and low-NaCl groups showed distinct L-NAME-resistant relaxations, which indicates that these remaining responses to ACh were mediated by endothelial products other than NO or COX.

The endothelium-dependent relaxations, which remain resistant to NOS and COX inhibition, are mediated by another vasoactive substance, EDHF (Cohen and Vanhoutte 1995). The chemical characteristics of EDHF remain unknown, but functionally this factor is a K^+ channel opener, the action of which can be inhibited by K^+ channel blockers or by depolarization of the cell membrane with high concentrations of K⁺ (Adeagbo and Triggle 1993). In the L-NAME hypertension study, although all of the present groups showed distinct NOS- and COX inhibitor-resistant relaxations to ACh, the remaining responses in the L-NAME group were attenuated when compared with other groups, whereas the responses in the calcium+L-NAME group did not differ from control. Thus, calcium supplementation prevented the impairment of endothelium-dependent hyperpolarization in L-NAME treated Wistar rats. The precontraction of arterial rings with KCl almost abolished the remaining NOS- and COX inhibitor-resistant relaxatios to ACh, suggesting that these responses were indeed mediated by EDHF. Furthermore, the findings in the NaCl-hypertension study supported the view that calcium supplementation normalized the impaired endotheliummediated relaxations by enhancing smooth muscle hyperpolarization. The 1OH-D₃-induced chronic hypercalcemia was without effects on the endothelium-dependent vasorelaxations in NaCl-hypertension. Decreased endothelium-dependent hyperpolarization has previously been observed in various forms of experimental hypertension (Fujii et al. 1992, Van de Voorde et al. 1992, Mäkynen et al. 1996), and the present results suggested that the same holds true in L-NAME-induced hypertension as well.

Impaired endothelium-dependent hyperpolarization could result from decreased endothelial release of EDHF or from reduced sensitivity of smooth muscle to EDHF. The present results, whereby the relaxations induced by the K_{ATP} opener cromakalim were attenuated in L-NAME hypertension, suggest that the sensitivity of smooth muscle to hyperpolarizing factors was decreased. Furthermore, isoprenaline has been also been reported to hyperpolarize arterial smooth muscle via K_{ATP} and K_{Ca} (Randall and McCulloch 1995, Song and Simard 1995). Therefore, the finding, whereby relaxation to isoprenaline was impaired in L-NAME hypertensive rats, is in agreement with the view of reduced hyperpolarization of arterial smooth muscle in these animals. Moreover, enhanced hyperpolarization could also explain the improved vasorelaxation to isoprenaline and to cromakalim following calcium supplementation in NaCl hypertension as well. Because similar endothelium-independent changes in vasorelaxation were detected in both L-NAME and NaCl hypertension with or without increased calcium intake, they are likely to result from the long-term elevation of blood pressure and the antihypertensive effect of high calcium diet. In our study, the 1OH-D₃ treatment also attenuated the development of NaCl-induced hypertension, but this effect may have been explained by reduced arterial constrictor responses and the impaired growth of the rats. It is noteworthy, that the 1OH-D₃-induced hypercalcemia did not have any favourable influences on vasorelaxation in NaCl hypertension, but it prevented the changes induced by high calcium diet in the control of arterial tone. Therefore, the vascular effects of calcium supplementation may partially be mediated via the suppression of Ca^{2+} regulating hormones.

The arterial relaxations induced by the NO donor SNP have been found to be enhanced or to remain unaffected in L-NAME hypertensive rats (Bryant et al. 1995, Dowell et al. 1996). In our studies, the L-NAME- and the NaCl-hypertensive rats showed attenuated relaxations to SNP in both NA- and KCl-precontracted endothelium-denuded arterial rings, suggesting that the sensitivity of arterial smooth muscle to NO was decreased. Calcium supplementation normalized this abnormality in the calcium+L-NAME group. Previously, high calcium diet has been suggested to enhance sensitivity to exogenous NO in deoxycorticosterone-NaCl hypertension (Mäkynen et al. 1996) and in SHR as well (Tolvanen et al. 1998).

5 Arterial relaxation in renal failure

In rats with CRF induced by the subtotal renal artery ligation, the large mesenteric artery relaxation to ACh was attenuated, and although L-NAME diminished the relaxations, this effect was more pronounced in the renal failure rats than in controls. Therefore, endothelium-mediated relaxations in the renal failure WKY rats were predominantly mediated by NO,

whereas the controls showed distinct L-NAME resistant relaxations to ACh. In contrast to L-NAME, diclofenac had no effect on ACh-induced relaxations, suggesting that the products of the COX pathway were not playing a significant role in the responses to ACh in the conduit arteries of these rats.

The distinct NOS and COX inhibitor-resistant relaxations to ACh were more pronounced in the sham-operated control rats. The combination of apamin and charybdotoxin was without effect on the L-NAME and diclofenac-resistant relaxation to ACh in the renal failure rats, but it significantly inhibited the response in the control rats, so that the difference in the remaining relaxation to ACh between the groups was abolished. This suggests that decreased endothelium-dependent dilatation of large mesenteric arteries in the renal failure rats was associated with reduced relaxation via mechanism including the activation of K⁺ channels and the subsequent hyperpolarization of arterial smooth muscle.

As previously reported, the sensitivity of arterial smooth muscle to NO was not altered in renal failure (Verbeke et al. 1994). However, the endothelium-independent relaxations in the large mesenteric arteries induced by isoprenaline and cromakalim were impaired in WKY rats with renal failure. In addition to the elevation of intracellular cAMP, isoprenaline has been reported to open K_{ATP} in the smooth muscle of rat mesenteric artery as discussed above (Randall and McCullough 1995). Therefore, the impaired function of K⁺ channels in smooth muscle could explain the reduced relaxations to the endothelium-independent agonists and the impaired endothelium-mediated hyperpolarization in experimental renal failure.

In spite of the crucial role of small arteries in the regulation of peripheral vascular resistance, the present knowledge about resistance vessel function in CRF is scarce. Isolated subcutaneous resistance arteries from uremic patients with systolic hypertension have shown impaired endothelium-mediated relaxation (Morris et al. 2001), but no such impairment has been found in the skin microcirculation of normotensive patients with renal failure (Cupisti et al. 2000). Experimental renal failure has been reported to impair the vasorelaxation induced by hypoxia in rat resistance arteries (Liu et al. 1997a) whereas the endothelium-mediated relaxation to Ach in the presence of COX inhibition in vitro did not differ from control (Thuraisingham and Raine 1999). However, in these experimental studies, blood pressure was also elevated in the uraemic rats (Liu et al. 1997, Thuraisingham and Raine 1999).

The resistance arteries of the renal failure WKY rats showed similar responses to ACh in the absence and presence of L-NAME when compared with the control animals. Moreover, diclofenac was also without significant effect on the responses, but the further adding of apamin and charybdotoxin in the organ bath chamber markedly diminished the NOS and COX inhibitor-resistant relaxations that still remained comparable between the study groups. This suggests that the endothelium-dependent vasodilatation via the mechanisms that include the activation of K^+ channels and the subsequent hyperpolarization of arterial smooth muscle was preserved in the resistance arteries of the uraemic WKY rats.

The sensitivity of resistance artery smooth muscle to NO was not altered in renal failure, since the vasorelaxations of endothelium-denuded rings to nitroprusside were similar in both study groups. Moreover, the endothelium-independent relaxations induced by cromakalim were also similar, indicating corresponding endothelium-independent hyperpolarization via the opening of KATP. However, the endothelium-independent small artery relaxations induced by the β -adrenoceptor agonist isoprenaline were impaired in the renal failure group. In the rat mesenteric resistance arteries, isoprenaline is proposed to induce relaxation by hyperpolarizing vascular smooth muscle via the opening of K_{ATP} (Fujii et al. 1999). The signalling cascade leading to vasorelaxation is suggested to be β -adrenoceptor/Gs protein/adenylate cyclase/K_{ATP} (Fujii et al. 1999). In our study, the vasorelaxation via K_{ATP} was not altered since the relaxations induced by the KATP opener cromakalim did not differ from controls. Therefore, the impaired resistance artery relaxation by isoprenaline in the uraemic rats could not be attributed to changes in the hyperpolarization mechanisms of arterial smooth muscle, but was probably inflicted by a defect at the level β-adrenoceptor/Gs protein/adenylate cyclase coupling step.

6 Influence of high calcium diet on resistance artery relaxation in more advanced renal failure

The present results showed a clear impairment in endothelium-dependent relaxations 12 weeks after surgical subtotal nephrectomy in Sprague-Dawley rats, while high calcium diet for 8 weeks normalized the response to ACh in NTX rats. Since the impaired response to ACh in the renal failure group was also observed in the presence of NOS and COX inhibition, but not in the presence of K_{Ca} blockade, these results imply that endothelium-dependent vasodilatation in experimental renal failure was reduced via a mechanism, which involved activation of K⁺ channels in arterial smooth muscle. Reduced endothelium-dependent relaxation via K⁺ channels could result from decreased endothelial release of EDHF, or reduced sensitivity of smooth muscle to EDHF. We found that the endothelium-independent relaxations to KATP opener levcromakalim and the KCa opener EET were reduced in the renal failure rats, while the high calcium intake also normalized these impairments. Therefore, the deficient vasodilatation at the level of smooth muscle K⁺ channels was corrected by increased calcium intake in renal failure. The high calcium diet was without significant effect on blood pressure. The improved vasorelaxation did not correlate with plasma levels of creatinine and urea, but was associated with reduced plasma levels of PTH and phosphate, and elevated plasma levels of ionized calcium. These results support the view that alterations in calciumphosphate balance contribute to the impairment of vasodilatation during deficient kidney function (Rostand and Drücke 1999).

Changes in the plasma concentration of phosphate, calcium and PTH could affect blood

vessels in many ways. Elevated phosphate may influence metabolism of vascular smooth muscle, and induce phenotypic changes that predispose the vessel wall to calcification (Jono et al. 2000). A link between extracellular calcium and arterial tone is the calcium receptor located in pervascular nerves, the activation of which causes vasorelaxation via the release of a hyperpolarizing mediator (Mupanomunda et al. 1999). The link between PTH and vascular tone is complex. PTH excess may increase cytoplasmic Ca²⁺ or alter the production of endothelium-derived vasoactive factors (Rostand and Drücke 1999). Acutely PTH causes vasodilatation, but subacute infusion of physiological doses of PTH raises blood pressure, since vascular desensitization to PTH takes place rapidly (Rostand and Drücke 1999). Moreover, at the cellular level PTH is linked to increased production of 20-hydroxyeicosatetraenoic acid, an endogenous vasoconstrictor that acts in part by inhibiting the opening of K_{Ca} in smooth muscle (Roman et al. 2000).

In clinical renal failure, the importance of effective treatment of hyperphosphatemia in the prevention of cardiovascular complications is well recognized (Rostand and Drücke 1999, Slatopolsky et al. 2001). The treatment of SH remains slightly controversial, since a 2-3-fold elevation in plasma PTH has been considered beneficial to bones of patients with impaired kidney function. However, the cardiovascular actions of moderate chronic elevations in PTH are not known in renal failure. Recently, it has been proposed that aggressive management of calcium and phosphorus disturbances at their earliest stage could prevent or attenuate the subsequent emergence of hyperparathyroidism and mitigate the associated risk of cardiovascular events (Drücke and McCarron 2003). Therefore, these results warrant such clinical studies, where the vascular actions of long-term treatments of SH are elucidated in patients with moderate kidney failure.

SUMMARY AND CONCLUSIONS

The present study was designed to examine the effects of high calcium intake on blood pressure and arterial function in L-NAME- and NaCl-induced forms of experimental hypertension. Furthermore, the influence of vitamin D-induced hypercalcemia in NaCl hypertension was studied. The influence of CRF on the control of large and small artery tone was investigated and resistance artery relaxation was further evaluated following the treatment of SH by high calcium diet in CRF.

The major findings and conclusions are:

1. The attenuated mesenteric arterial relaxations in different forms of experimental hypertension and renal failure seemed to be especially related to reduced vasodilatation via potassium channels:

1.1. Chronic L-NAME hypertension was associated with the impairment of endothelium-dependent and –independent vasorelaxation. Concomitant long-term calcium supplementation clearly improved arterial dilatation in this form of NO deficiency and reduced the elevated blood pressure. The mechanisms underlying the augmented vasodilatation following calcium supplementation in this model of experimental hypertension may have included enhanced arterial hyperpolarization, increased sensitivity to NO in smooth muscle, decreased vascular production of superoxide and vasoconstrictor prostanoids.

1.2. Long-term calcium supplementation and 1OH-D_3 -induced hypercalcemia differently influenced the control of arterial tone in NaCl hypertension. Increased calcium intake lowered blood pressure, and improved endothelium-dependent and – independent vasorelaxation, while vascular contractile properties remained unaffected. This is in good agreement with the findings in chronic NO deficiency, but the effect of high calcium intake on blood pressure was more pronounced in NaCl hypertension. The reduced blood pressure and improved arterial relaxation after calcium supplementation may have been explained by enhanced arterial hyperpolarization and increased sensitivity to NO in smooth muscle. Enhanced natriuresis following calcium supplementation may have played a role as well in the reduction of blood pressure, although this was not directly determined in either of the present experimental hypertension studies. The chronic 1OH-D_3 -induced hypercalcemia also moderately lowered blood pressure in NaCl hypertension, which was probably explained by

reduced arterial contractile properties and impaired growth of the rats, which may have been an adverse effect of the 1OH-D₃. The 1OH-D₃ treatment also prevented all beneficial changes induced by high calcium diet in the control of arterial tone.

1.3. Rats with 6-week renal failure showed impaired endothelium-mediated relaxation of the large arteries in the absence and presence of NOS inhibition but not under conditions when hyperpolarization was blocked. In addition, endothelium-independent relaxations via the activation of β -adrenoceptors and the opening of K⁺ channels were reduced. Therefore, impaired large artery relaxation also in experimental renal failure could be attributed to reduced vasodilation via potassium channels. The resistance arteries of renal failure rats featured impaired relaxations via the activation of β -adrenoceptors, but the endothelium-mediated small artery relaxations did not differ from control 6 weeks after the subtotal nephrectomy.

1.4. Experimental 12-week renal failure in subtotally nephrectomized rats resulted in modest elevation of blood pressure. CRF rats also showed impaired resistance artery relaxation via K^+ channels, which was normalized by increased calcium intake that also prevented the development of SH. The improved small artery relaxation was associated with reduced plasma levels of PTH and phosphate, and elevated plasma levels of ionized calcium. However, high calcium intake did not correct the eutrophic inward remodelling of the resistance arteries in rats with CRF.

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Pasi Jolma

REFERENCES

Abdel-Latif AA (1986): Calcium-mobilizing receptors, polyphosphoinositides and the generation of second messangers. Pharmacol Rev 38:227-272.

Adeagbo ASO and Triggle CR (1993): Varying extracellular $[K^+]$: a functional approach to separating EDHFand EDNO-related mechanisms in perfused rat mesenteric arterial bed. J Cardiovasc Pharmacol 21:423-429.

Agapitov AV and Haynes WG (2002): Role of endothelin in cardiovascular disease. J Renin Angiotensin Aldosterone Syst 3:1-15.

Aguilera G and Catt K (1981): Regulation of vascular angiotensin II receptors during altered sodium intake. Circ Res 49:751-758.

Aiello S, Noris M, Todeschini M, Zappella S, Foglieni C, Benigni A, Corna D, Zoja C, Cavallotti D and Remuzzi G (1997): Renal and systemic nitric oxide synthesis in rats with renal mass reduction. Kidney Int 52:171-181.

Akita S, Sacks FM, Svetkey LP, Conlin PR, Kimura G and DASH-Sodium Trial Collaborative Research Group (2003): Effects of the Dietary Approaches to Stop Hypertension (DASH) diet on the pressure-natriuresis relationship. Hypertension 42:8-13.

Akuzawa N, Nakamura T, Kurashina T, Saito Y, Hoshino J, Sakamoto H, Sumino H, Ono Z and Nagai R (1998): Antihypertensive agents prevent nephrosclerosis and left ventricular hypertrophy induced in rats by prolonged inhibition of nitric oxide synthesis. Am J Hypertens 11:697-707.

Allen BG and Walsh MP (1994): The biochemical basis of the regulation of smooth-muscle contraction. Trends Biochem Sci 19:362-368.

Amann K, Wolf B, Nichols C, Tornig J, Schwarz U, Zeier M, Mall G and Ritz E (1997): Aortic changes in experimental renal failure: hyperplasia or hypertrophy of smooth muscle cells? Hypertension 29:770-775.

Amberg GC, Bonev AD, Rossow CF, Nelson MT and Santana LF (2003): Modulation of the molecular composition of large conductance, Ca^{2+} activated K⁺ channels in vascular smooth muscle during hypertension. J Clin Invest 112:717-724.

Anderstam B, Katzarski K and Bergström J (1997): Serum levels of N^G , N^G -dimethyl-L-arginine, a potential endogenous nitric oxide inhibitor in dialysis patients. J Am Soc Nephol 8:1437-1442.

Andrea JE and Walsh MP (1992): Protein kinase C of smooth muscle. Hypertension 20:585-595.

Annuk M, Zilmer M, Lind L, Linde T and Fellstrom B (2001): Oxidative stress and endothelial function in chronic renal failure. J Am Soc Nephrol 12:2747-2752.

Annuk M, Zilmer M and Fellstrom B (2003): Endothelium-dependent vasodilation and oxidative stress in chronic renal failure: Impact on cardiovascular disease. Kidney Int 63:S50-S53.

Aoki K and Asano M (1986): Effects of Bay K 8644 and nifedipine on femoral arteries of spontaneously hypertensive rats. Br J Pharmacol 88:221-230.

Aoki K and Asano M (1987): Increased responsiveness to calcium agonist BAY K 8644 and calcium antagonist nifedipine in femoral arteries of spontaneously hypertensive rats. J Cardiovasc Pharmacol 10:S62-S64.

Appel LJ, Moore TJ, Obarzanek E, Vollmer WM, Svetkey LP, Sacks FM, Bray GA, Vogt TM, Cutler JA, Windhauser MM, Lin PH and Karanja N (1997): A clinical trial of the effects of dietary patterns on blood pressure. DASH collaborative research group. N Engl J Med 336:1117-1124.

Appel LJ, Miller ER, Jee SH, Stolzenberg-Solomon R, Lin P-H, Erlinger T, Nadeau MR and Selhub J (2000): Effect of dietary patterns on serum homocysteine. Results of a randomised, controlled feeding study. Circulation 102:852-857.

Archer SL, Gragasin FS, Wu X, Wang S, McMurtry S, Kim DH, Platonov M, Koshal A, Hashimoto K, Campbell WB, Falck JR and Michelakis ED (2003): Endothelium-derived hyperpolarizing factor in human internal mammary artery is 11,12-epoxyeicosatrienoic acid and causes relaxation by activating smooth muscle BK(Ca) channels. Circulation 11:769-776.

Arii T, Ohyanagi M, Shibuya J and Iwasaki T (1999): Increased function of the voltage-dependent calcium channels, without increase of Ca^{2+} release from the sarcoplasmic reticulum in the arterioles of spontaneous hypertensive rats. Am J Hypertens 12:1236-1242.

Arnal JF, Warin L and Michel JB (1992): Determinants of aortic cyclic guanosine monophosphate in hypertension induced by chronic inhibition of nitric oxide synthase. J Clin Invest 90:647-652.

Arnal JF, El Amrani AI, Chatellier G, Ménard J and Michel JB (1993): Cardiac weight in hypertension induced by nitric oxide synthase blockade. Hypertension 22:380-387.

Arvola P, Pörsti I, Vuorinen P, Pekki A and Vapaatalo H (1992): Contractions induced by potassium-free solution and potassium relaxation in vascular smooth muscle of hypertensive and normotensive rats. Br J Pharmacol 106:157-165.

Arvola P, Ruskoaho H and Pörsti I (1993a): Effect of high calcium diet on arterial smooth muscle function and electrolyte balance in mineralocorticoid-salt hypertensive rats. Br J Pharmacol 108:948-958.

Arvola P, Ruskoaho H, Wuorela H, Pekki A, Vapaatalo H and Pörsti I (1993b): Quinapril treatment and arterial smooth muscle responses in spontaneously hypertensive rats. Br J Pharmacol 108:980-990.

Asano M, Nomura Y, Ito K, Uyama Y, Imaizumi Y and Watanabe M (1995): Increased function of voltagedependent Ca^{2+} channels and Ca^{2+} -activated K⁺ channels in resting state of femoral arteries from spontaneously hypertensive rats at prehypertensive stage. J Pharmacol Exp Ther 275:775-783.

Ashida T, Kuramochi M and Omae T (1989): Increased sodium-calcium exchange in arterial smooth muscle of spontaneously hypertensive rats. Hypertension 13:890-895.

Auch-Schwelk W and Vanhoutte PM (1991): Endothelium-derived contracting factor released by serotonin in aorta of the spontaneously hypertensive rat. Am J Hypertens 4:769-772.

Ayachi S (1979): Increased dietary calcium lowers blood pressure in the spontaneously hypertensive rat. Metabolism 28:1234-1238.

Bagrov AY and Fedorova OV (1998): Effects of two putative endogenous digitalis-like factors, marinobufagenin and ouabain, on the Na+,K+-pump in human mesenteric arteries. J Hypertens 16:1953-1958.

Ballermann BJ, Zeidel ML, Gunning ME and Brenner BM (1991): Vasoactive peptides and the kidney. The Kidney. WB Saunders Co. 510-583.

Banting JD, Thompson KE, Friberg P and Adams MA (1997): Blunted cardiovascular growth induction during prolonged nitric oxide synthase blockade. Hypertension 30:416-421.

Barenbrock M, Spieker C, Laske V, Heidenreich S, Hohage H, Bachmann J, Hoeks APG and Rahn KH (1994): Studies of the vessel wall properties in hemodialysis patients. Kidney Int 44:1397-1400.

Baron A, Frieden M and Beny JL (1997): Epoxyeicosatrienoic acids activate a high conductance Ca(2+)-dependent K+ channel on pig coronary artery endothelial cells. J Physiol 504:537-543.

Bartunek J, Weinberg EO, Tajima M, Rohrbach S, Katz SE, Douglas PS and Lorell BH (2000): Chronic N^Gnitro-L-arginine methyl ester-induced hypertension: novel molecular adaptation to systolic load in absence of hypertrophy. Circulation 101:423-429.

Bauersachs J, Popp R, Hecker M, Sauer E, Fleming I and Busse R (1996): Nitric oxide attenuates the release of endothelium-derived hyperpolarizing factor. Circulation 94:3341-3347.

Baylis C, Mitruka B and Deng A (1992): Chronic blockade of nitric oxide synthesis in the rat produces systemic hypertension and glomerular damage. J Clin Invest 90:278-281.

Behrendt D and Ganz P (2002): Endothelial function: from vascular biology to clinical applications. Am J Cardiol 90:40L-48L.

Bellinghieri G, Santoro D, Mazzaglia G and Savica V (1999): Hypertension in dialysis patients. Miner Electrolyte Metab 25:84-89.

Bendhack LM, Sharma RV and Bhalla RC (1992): Altered signal transduction in vascular smooth muscle cells of spontaneously hypertensive rats. Hypertension 19:II142-II148.

Bennett MA, Hillier C and Thurston H (1996): Endothelium-dependent relaxation in resistance arteries from spontaneously hypertensive rats: effect of long-term treatment with perindopril, quinapril, hydralazine or amlodipine. J Hypertens 14:389-397.

Berk BC, Vallega G, Muslin AJ, Gordon HM, Canessa M and Alexander RW (1989): Spontaneously hypertensive rat vascular smooth muscle cells in culture exhibit increased growth and Na^+/K^+ exchange. J Clin Invest 83:822-829.

Bernatova I, Pechanova O, Bapal P, Kysela S, Stvrtina S and Andriantsitohaina R (2002): Wine polyphenols improve cardiovascular remodelling and vascular function in NO-deficient hypertension. Am J Physiol Heart Circ Physiol 282:H942-H948.

Bian K and Bukoski RD (1995): Myofilament calcium sensitivity of normotensive and hypertensive resistance arteries. Hypertension 25:110-116.

Bian K, Ishibashi K and Bukoski RD (1996): $1,25(OH)_2D_3$ modulates intracellular Ca²⁺ and force generation in resistance arteries. Am J Physiol 270:H230-H237.

Blacher J, Guerin AP, Pannier B, Marchais SJ and London GM (2001): Arterial calcifications, arterial stiffness, and cardiovascular risk in end-stage renal disease. Hypertension 38:938-942.

Blacher J, Guerin AP, Pannier B, Marchais SJ, Safar ME and London GM (1999): Impact of aortic stiffness on survival in end-stage renal disease. Circulation 99:2434-2439.

Blacher J, Pannier B, Guerin AP, Marchais SJ, Safar ME and London GM (1998): Carotid artery stiffness as a predictor of cardiovascular and all-cause mortality in end-stage renal disease. Hypertension 32:570-574.

Black CE, Huang N, Neligan PC, Forrest CR, Lipa JE and Pang CY (2003): Vasoconstrictor effect and mechanism of action of endothelin-1 in human radial artery and vein: implication of skin flap vasospasm. J Cardiovasc Pharmacol 41:460-467.

Blaustein MP (1993): Physiological effects of endogenous ouabain: control of intracellular Ca²⁺ stores and cell responsiveness. Am J Physiol 264:C1367-C1387.

Bockman CS, Jeffries WB, Pettinger WA and Abel PW (1992): Enhanced release of endothelium-derived relaxing factor in mineralocorticoid hypertension. Hypertension 20:304-313.

Boegehold MA (2002): Microvascular structure and function in salt-sensitive hypertension. Microcirculation 9:225-241.

Bohlen HG (1986): Localization of vascular resistance changes during hypertension. Hypertension 8:181-183.

Bolton TB, Lang RJ and Takewaki T (1984): Mechanism of action of noradrenaline and carbachol on smooth muscle of guinea-pig anterior mesenteric artery. J Physiol 351:549-572.

Bolz SS, Fisslthaler B, Pieperhoff S, De Wit C, Fleming I, Busse R and Pohl U (2000): Antisense oligonucleotides against cytochrome P450 2C8 attenuate EDHF-mediated Ca(2+) changes and dilation in isolated resistance arteries. FASEB J 14:255-260.

Boulanger CM (1999): Secondary endothelial dysfunction: hypertension and heart failure. J Mol Cell Cardiol 31:39-49.

Bova S, Goldman WF, Yuan X-J and Blaustein MP (1990): Influence of Na⁺ gradient on Ca²⁺ transients and contraction in vascular smooth muscle. Am J Physiol 259:H409-H423.

Brodde O-E and Michel MC (1992): Adrenergic receptors and their signal transduction mechanism in hypertension. J Hypertens 10:133-145.

Braman RS and Hendrix SA (1989): Nanogram nitrite and nitrate determination in environmental and biological materials by vanadium (III) reduction with chemiluminescence detection. Anal Chem 61:2715-2718.

Brayden JE and Nelson MT (1992): Regulation of arterial tone by activation of calcium-dependent potassium channels. Science 256:532-535.

Brown EM, Gamba G, Riccardi D, Lombardi M, Butters R, Kifor O, Sun A, Hediger MA, Lytton J and Hebert SC (1993): Cloning and characterization of an extracellular Ca^{2+} sensing receptor from bovine parathyroid. Nature 366:575-580.

Brown EM, Wilkson RE, Eastman RC, Pallotta J and Marynick SP (1982): Abnormal regulation of parathyroid hormone release by calcium in secondary hyperparathyroidism due to chronic renal failure. J Clin Endocrin Metab 54:172-179, 1982.

Bruner CA and Webb RC (1990): Increased vascular reactivity to Bay K 8644 in genetic hypertension. Pharmacology 41:24-35.

Brunner HR, Laragh JR, Baer L, Newton MA, Goodwin FT, Krakoff LR, Bard RH and Buhler FR (1972): Essential hypertension: renin and aldosterone, heart attact and stroke. N Engl J Med 286:441-449.

Bryant CE, Allcock GH and Warner TD (1995): Comparison of effects of chronic and acute administration of N^{G} -nitro-L-arginine methyl ester to rat on inhibition of nitric oxide-mediated responses. Br J Pharmacol 114:1673-1679.

Bucher HC, Cook RJ, Guyatt GH, Lang JD, Cook DJ, Hatala R and Hunt DL (1996): Effects of dietary calcium supplementation and blood pressure. A meta-analysis of randomized controlled trials. JAMA 275:1016-1022.

Bülbring E and Tomita T (1987): Catecholamine action on smooth muscle. Pharmacol Rev 39:49-96.

Bukoski RD (1998): The perivascular sensory nerve Ca^{2+} receptor and blood pressure regulation: a hypothesis. Am J Hypertens 11:1117-1123.

Bukoski RD, Xue H and McCarron DA (1989): Mesenteric artery contractile properties during dietary calcium manipulation in spontaneously hypertensive and Wistar Kyoto normotensive rats. Am J Hypertension 2:440-448.

Bukoski RD, Li J and Bo J (1993): Effect of long-term administration of $1,25-(OH)_2$ vitamin D₃ on blood pressure and resistance artery contractility in the spontaneously hypertensive rat. Am J Hypertens 6:944-950,

Bukoski RD, Lastelic BA, Xue H, Li J and Bian K (1994): Intracellular Ca^{2+} and force generation determined in resistance arteries of normotensive and hypertensive rats. J Hypertens 12:15-21.

Bukoski RD, Ishibashi K and Bian K (1995): Vascular actions of calcium regulating hormones. Sem Nephrol 15:536-549.

Burnham MP, Bychkov R, Félétou M, Richards GR, Vanhoutte PM, Weston AH and Edwards G (2002): Characterization of an apamin-sensitive small conductance Ca(2+)-activated K(+) channel in porcine coronary artery endothelium: relevance to EDHF. Br J Pharmacol 135:1133-1143.

Burnier M (2001): Angiotensin II type 1 receptor blockers. Circulation 103:904-912.

Burt VL, Whelton P, Roccella EJ, Brown C, Cutler JA, Higgins M, Horan MJ and Labarthe D (1995): Prevalence of hypertension in the US adult population. Results from the Third National Health and Nutrition Examination Survey, 1988-1991. Hypertension 25:305-313.

Busse R and Fleming I (1993): The endothelial organ. Curr Opin Cardiol 8:719-727.

Busse R, Hecker M and Fleming I (1994): Control of nitric oxide and prostacyclin synthesis in endothelial cells. Arznein-Forsch Drug Res 44:392-396.

Butler TV, Cameron J and Kirchner KA (1995): Dietary calcium supplementation restores pressure natriuresis responses in Dahl-S rats. Am J Hypertens 8:615-621.

Cai H, Griendling KK and Harrison DG (2003): The vascular NAD(P)H oxidases as therapeutic targets in cardiovascular diseases. Trends Pharmacol Sci 24:471-478.

Cai S, Garneau L and Sauve R (1998): Single channel characterization of the pharmacological properties of the K(Ca2+) channel of intermediate conductance in bovine aortic endothelial cells. J Membr Biol 163:147-158.

Campbell WB, Gebremehdin D, Pratt PF and Harder DR (1996): Identification of epoxyeicosatrienoic acids as endothelium-derived hyperpolarizing factors. Circ Res 78:415-423.

Campbell WB and Harder DR (1999): Endothelium-derived hyperpolarizing factors and vascular cytochrome P450 metabolites of arachidonic acid in the regulation of tone. Circ Res 84:484-488.

Campia U, Choucair WK, Bryant MB, Quyymi AA, Cardillo C and Panza JA (2002): Role of cyclooxygenase products in the regulation of vascular tone and in the endothelial vasodilator function in normal, hypertensive, and hypercholesterolemic humans. Am J Cardiol 89:286-290.

Cao Z, Dean R, Wu L, Casley D and Cooper ME (1999): Role of angiotensin receptor subtypes in mesenteric vascular proliferation and hypertrophy. Hypertension 34:408-414

Chen G and Suzuki H (1989): Some electrical properties of the endothelium-dependent hyperpolarisation recorded from rat arterial smooth muscle cells. J Physiol 410:91-106.

Carl A, Lee HK and Sanders KM (1996): Regulation of ion channels in smooth muscle by calcium. Am J Physiol 271:C9-C34.

Celermajer DS (1997): Endothelial dysfunction: does it matter? Is it reversible? J Am Coll Cardiol 30:325-333.

Chalmers J, Arnolda L, Kapoor V, Llewellyn-Smith I, Minson J and Pilowsky P (1992): Amino acid neurotransmitters in the central control of blood pressure and in experimental hypertension. J Hypertens Suppl 10:S27-S37.

Chen G and Suzuki H (1990): Calcium dependency of the endothelium-dependent hyperpolarization in smooth muscle cells of the rabbit carotid artery. J Physiol 421:521-534.

Chen G, Suzuki H and Weston AH (1988): Acetylcholine releases endothelium-derived hyperpolarizing factor and EDRF from rat blood vessels. Br J Pharmacol 95:1165-1174.

Chobanian AV, Bakris GL, Black HR, Cushman WC, Green LA, Izzo JL Jr, Jones DW, Materson BJ, Oparil S, Wright JT Jr and Roccella EJ; National Heart, Lung, and Blood Institute Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure; National High Blood Pressure Education Program Coordinating Committee (2003): The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. JAMA 21;289:2560-2572.

Cirillo M (1992): Intracellular calcium and blood pressure. Child Nephrol Urol 12:78-84.

Cirillo M, Capasso G and De Santo NG (1992): Altered cellular calcium metabolism in hypertension: A reassessment and a hypothesis. Contrib Nephrol 100:35-47.

Coats P, Johnston F, MacDonald J, McMurray JJV and Hillier C (2001): Endothelium-derived hyperpolarizing factor: identification and mechanisms of action in human subcutaneous resistance arteries. Circulation 103:1702-1708.

Cockroft JR, Chowienczyk PJ, Benjamin N and Ritter JM (1994): Preserved endothelium-dependent vasodilatation and vasoconstriction in patients with essential hypertension. N Engl J Med 330:1036-1040.

Cockroft JR, O'Kane KPJ and Webb DJ (1995): Tissue angiotensin generation and regulation of vascular tone. Pharmac Ther 65:193-213.

Cohen RA (1995): The role of nitric oxide and other endothelium-derived substances in vascular disease. Prog Cardiovasc Dis 38:105-128.

Cohen RA and Vanhoutte PM (1995): Endothelium-dependent hyperpolarization. Beyond nitric oxide and cyclic GMP. Circulation 92:3337-3349.

Coleman HA, Tare M and Parkington HC (2001): K+ currents underlying the action of endothelium-derived hyperpolarizing factor in guinea-pig, rat and human blood vessels. J Physiol 531:359-373.

Colombari E, Sato MA, Cravo SL, Bergamaschi CT, Campos RR Jr and Lopes OU (2001): Role of the medulla oblongata in hypertension. Hypertension 38:549-554.

Corriu C, Félétou M, Canet E and Vanhoutte PM (1996): Endothelium-derived factors and hyperpolarisations of the isolated carotid artery of the guinea-pig. Br J Pharmacol 119:959-964.

Cowan CL and Cohen RA (1991): Two mechanisms mediate relaxation by bradykinin of pig coronary artery: NO-dependent and independent responses. Am J Physiol 261:H830-H835.

Cowley AW Jr (1997): Role of the renal medulla in volume and arterial pressure regulation. Am J Physiol 273:R1-R15.

Cowley AW Jr and Roman RJ (1996): The role of the kidney in hypertension. JAMA 275:1581-1589.

Culman J and Unger T (1992): Central mechanisms regulating blood pressure: circuits and transmitters. Eur Heart J 13 Suppl: 13-17.

Cunha RS, Cabral AM and Vasquez EC (1993): Evidence that the autonomic nervous system plays a major role in the L-NAME-induced hypertension in conscious rats. Am J Hypertens 6:806-809.

Cupisti A, Rossi M, Placidi S, Fabbri S, Morelli E, Vagheggini G, Meola M and Barsotti G (2000): Responses of the skin microcirculation to acetylcholine in patients with essential hypertension and in normotensive patients with chronic renal failure. Nephron 85:114-119.

Cutler JA, Follmann D and Allender PS (1997): Randomized trials of sodium reduction: an overview. Am J Clin Nutr 65(2 Suppl):6438-651S.

Danser AH (1996): Local renin-angiotensin systems. Mol Cell Biochem 157:211-216.

Davies JE, Ng LL, Ameen M, Syme PD and Aronson JK (1991): Evidence for altered Na^+/K^+ antiport activity in cultured skeletal muscle cells and vascular smooth muscle cell from the spontaneously hypertensive rat. Clin Sci 80:509-516.

Deferrari G, Ravera M, Deferrari L, Vettoretti S, Ratto E and Parodi D (2002): Renal and cardiovascular protection in type 2 diabetes mellitus: angiotensin II receptor blockers. J Am Soc Nephrol 13:S224-S229.

De Gracia MC, Osuna A, O'Valle F, del Moral RG, Wangensteen R, del Rio CG and Vargas F (2000): Deoxycorticosterone suppresses the effects of losartan in nitric oxide-deficient hypertensive rats. J Am Soc Nephrol 11:1995-2000.

De Wardener HE (2001): The hypothalamus and hypertension. Physiol Rev 81:1599-1658.

DeLong LJ and Blasie JK (1993): Effect of Ca^{2+} binding on the profile structure of the sarcoplasmic reticulum membrane using time-resolved x-ray diffraction. Biophys J 64:1750-1759.

Demuth K, Blacher J, Guerin AP, Benoit MO, Moatti N, Safar ME and London GM (1998): Endothelin and cardiovascular remodelling in end-stage renal disease. Nephrol Dial Transplant 13:375-383.

Deng LY, Thibault G and Schiffrin EL (1993): Effect of hypertension induced by nitric oxide synthase inhibition on structure and function of resistance arteries in the rat. Clin Exp Hypertens 15:527-537.

Denton D, Weisinger R, Munday NI, Wickings EJ, Dixson A, Moisson P, Pingard AM, Shade R, Carey D and Ardaillou R (1995): The effect of increased salt intake on blood pressure of chimpanzees. Nature Med 1:1009-1016.

Dibona GF and Kopp UC (1997): Neural control of renal function. Physiol Rev 77:75-197.

Doggrell SA and Brown L (1998): Rat models of hypertension, cardiac hypertrophy and failure. Cardiovasc Res 39:89-105.

Dohi Y, Aoki K, Fujimoto S, Kojima M and Matsuda T (1990): Alteration in sarcoplasmic reticulum-dependent contraction of tail arteries in response to caffeine and noradrenaline in spontaneously hypertensive rats. J Hypertens 8:261-267.

Dominiczak AF and Bohr DF (1990): Cell membrane abnormalities and the regulation of intracellular calcium concentration in hypertension. Clin Sci 79:415-421.

Dora KA and Garland CJ (2001): Properties of smooth muscle hyperpolarization and relaxation to K^+ in the rat isolated mesenteric artery. Am J Physiol Heart Circ Physiol 280:H2424-H2429.

Doughty JM, Plane F and Langton PD (1999): Charybdotoxin and apamin block EDHF in rat mesenteric artery if selectively applied to the endothelium. Am J Physiol 276:H1107-H1112.

Doughty JM, Boyle JP and Langton PD (2000): Potassium does not mimic EDHF in rat mesenteric arteries. Br J Pharmacol 130:1174-1182.

Dowell FJ, Henrion D, Duriez M and Michel JB (1996): Vascular reactivity in mesenteric resistance arteries following chronic nitric oxide synthase inhibition in Wistar rats. Br J Pharmacol 117:341-346.

Drücke TB (2001): The place of calcium and calcimimetics in the treatment of secondary hyperparathyroidism. Nephrol Dial Transplant 16:S15-S17.

Drücke TB and McCarron DA (2003): Paricalcitol as compared with calcitriol in patients undergoing hemodialysis. N Engl J Med 349:496-499.

Durak I, Akyol Ö, Basesme E, Canbolat O and Kavatcu M (1994): Reduced erythrocyte defense mechanisms against free radical toxicity in patients with chronic renal failure. Nephron 66:76-80.

Dustan HP, Valdes G, Bravo EL and Tarazi RC (1986): Excessive sodium retention as a characteristic of salt-sensitive hypertension. Am J Med Sci 292:67-74.

Dworkin LD, Hostetter TH, Rennke HG and Brenner BM (1984): Hemodynamic basis for glomerular injury in rats with desoxycorticosterone-salt hypertension. J Clin Invest 73:1448-1461.

Dzau VJ, Sasamura H and Hein L (1993): Heterogeneity of angiotensin synthetic pathways and receptor subtypes: physiological and pharmacological implications. J Hypertens 11:S13-S22.

Edwards G, Dora KA, Gardener MJ, Garland CJ and Weston AH (1998): K+ is an endothelium-derived hyperpolarizing factor in rat arteries. Nature 396:269-272.

Edwards G and Weston AH (1998): Endothelium-derived hyperpolarizing factor -- a critical appraisal. Prog Drug Res 50:107-133.

Edwards G, Félétou M, Gardener MJ, Thollon C, Vanhoutte PM and Weston AH (1999a): Role of gap junctions in the responses to EDHF in rat and guinea-pig small arteries. Br J Pharmacol 128:1788-1794.

Edwards G, Gardener MJ, Félétou M, Brady G, Vanhoutte PM and Weston AH (1999b): Further investigation of endothelium-derived hyperpolarizing factor (EDHF) in rat hepatic artery: studies using 1-EBIO and ouabain. Br J Pharmacol 128:1064-1070.

Edwards G, Thollon C, Gardener MJ, Félétou M, Vilaine J, Vanhoutte PM and Weston AH (2000): Role of gap junctions and EETs in endothelium-dependent hyperpolarization of porcine coronary artery. Br J Pharmacol 129:1145-1154.

Elliot P, Stamler J, Nichols R, Dyer AR, Stamler R, Kesteloot H, Marmot M, for the Intersalt Cooperative Research Group (1996): Intersalt revisited: further analyses of 24-hour sodium excretion and blood pressure within and across populations. Br Med J 312:1249-1253.

Esler M (1995): Sympathetic nervous system: contribution to human hypertension and related cardiovascular diseases. J Cardiovasc Pharmacol 26:S24-S28.

Esler M (2000): The sympathetic system and hypertension. Am J Hypertens 13:998-105S.

Et-Taouil K, Schiavi P, Levy BI and Plante GE (2001): Sodium intake, large artery stiffness, and proteoglycans in the spontaneously hypertensive rat. Hypertension 38:1172-1176.

Félétou M and Vanhoutte PM (1988): Endothelium-dependent hyperpolarization of canine coronary smooth muscle. Br J Pharmacol 93:515-524.

Félétou M and Vanhoutte PM (1999): Endothelium-derived hyperpolarizing factor. Drug News Perspect 12:217-222.

Fichtlscherer S, Rosenberger G, Walter DH, Breuer S, Dimmeler S and Zeiher AM (2000): Elevated C-reactive protein levels and impaired endothelial vasoreactivity in patients with coronary artery disease. Circulation 102:1000-1006.

Finch EA, Turner TJ and Goldin SM (1991): Calcium as a coagonist of inositol 1,4,5-triphosphate-induced calcium release. Science 252:443-446.

FissIthaler B, Popp R, Kiss L, Potente M, Harder DR, Fleming I and Busse R (1999): Cytochrome P450 2C is an EDHF synthase in coronary arteries. Nature 30:493-497.

Floras JS and Hara K (1993): Sympathoneural and haemodynamic characteristics of young subjects with mild essential hypertension. J Hypertens 11:647-655.

Foley RN, Parfrey PS and Sarnak MJ (1998): Epidemiology of cardiovascular disease in chronic renal disease. J Am Soc Nephrol 9(12 Suppl):S16-S23.

Forte JG, Pereira Miguel JM, Pereira Miguel MJ, de Padua F and Rose G (1989): Salt and blood pressure: a community trial. J Human Hypertens 3:179-184.

Fournier A, Oprisiu R, Morinière P and El Esper N (1996): Low doses of calcitriol or calcium carbonate for the prevention of hyperparathyroidism in predialysis patients? Nephrol Dial Transplant 11:1493-1495.

Frohlich ED (1997): Arthur C. Corcoran Memorial Lecture. Influence of nitric oxide and angiotensin II on renal involvement in hypertension. Hypertension 29:188-193.

Fujii K, Onaka U, Kenichi G, Abe I and Fujishima M (1999): Impaired isoproterenol-induced hyperpolarization in isolated mesenteric arteries of aged rats. Hypertension 34:222-228.

Fujii K, Tominaga M, Ohmori S, Kobayashi K, Koga T, Takata Y and Fujishima M (1992): Decreased endothelium-dependent hyperpolarization to acetylcholine in smooth muscle of the mesenteric artery of spontaneously hypertensive rats. Circ Res 70:660-669.

Fujino K (1984): Brain catecholamines in spontaneously hypertensive and DOCA-salt hypertensive rats. Acta Med Okayama 38:325-340.

Fukao M, Hattori Y, Kanno M, Sakuma I and Kitabatake A (1995): Thapsigargin- and cyclopiazonic acidinduced endothelium-dependent hyperpolarization in the rat mesenteric artery. Br J Pharmacol 115:987-992.

Furchgott RF and Zawadski JV (1980): The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. Nature 288:373-376.

Furchgott RF (1996): The 1996 Albert Lasker Medical Research Awards. The discovery of endothelium-derived relaxing factor and its importance in the identification of nitric oxide. JAMA 276:1186-1188.

Galle J, Quaschning T, Seibold S and Wanner C (2003): Endothelial dysfunction and inflammation: What is the link? Kidney Int 63(Suppl 84):S45-S49.

Garland CJ and McPherson GA (1992): Evidence that nitric oxide does not mediate the hyperpolarisation and relaxation to acetylcholine in the rat small mesentery artery. Br J Pharmacol 105:429:435.

Garland CJ, Plane F, Kemp BK and Cocks TM (1995): Endothelium-dependent hyperpolarization: a role in the control of vascular tone. Trends Pharmacol Sci 16:23-30.

Garland CJ and Plane F (1996): Relative importance of endothelium-derived hyperpolarizing factor for the relaxation of vascular smooth muscle in different arterial beds. In Endothelium-Derived Hyperpolarizing Factor (Vol. 1), (Vanhoutte PM ed.) Harwood Academic Publishers 173-179.

Gavras H, Brown JJ, Lever AF, Macadam RF and Robertson JI (1971): Acute renal failure, tubular necrosis and myocardial infarction induced in the rabbit by intravenous angiotensin II. Lancet 2:19-22.

Gavras H and Gavras I (1989): Salt-induced hypertension: the interactive role of vasopressin and of the sympathetic nervous system. J Hypertens 7:601-606.

Geleijnse JM, Hofman A, Witteman JCM, Hazebroek AAJM, Valkenburg HA and Grobbee DE (1997): Long term effects of neonatal sodium restriction on blood pressure. Hypertension 29:913-917.

Geisterfer AA, Peach MJ and Owens GK (1988): Angiotensin II induces hypertrophy, not hyperplasia, of cultured rat aortic smooth muscle cells. Circ Res 62:749-756.

Goddard J and Webb DJ (2000): Plasma endothelin concentration in hypertension. J Cardiovasc Pharmacol 35:S25-S31.

Goldstein DS (1983): Arterial baroreflex sensitivity, plasma catecholamines, and pressor responsiveness in essential hypertension. Circulation 68:234-240.

Gonzalez JM and Suki WN (1995): Cell calcium and arterial blood pressure. Semin Nephrol 15:564-568.

Goodman WG, Goldin J, Kuizon BD, Yoon C, Gales B, Sider D, Wang Y, Chung J, Emerick A, Greaser L, Elashoff RM and Salusky IB (2000): Coronary-artery calcification in young adults with end-stage renal disease who are undergoing dialysis. N Engl J Med 342:1478-1483.

Grassi G, Cattaneo BM, Seravalle G, Lanfranchi A and Mancia G (1998): Baroreflex control of sympathetic nerve activity in essential and secondary hypertension. Hypertension 31:68-72.

Gryglewski RJ (1995): Interactions between endothelial mediators. Pharmacol Toxicol 77:1-9.

Guérin AP, London GM, Marchais SJ and Métivier F (2000): Arterial stiffening and vascular calcifications in end-stage renal disease. Nephrol Dial Transplant 15:1014-1021.

Gupta S, Chough E, Daley J, Oates P, Tornheim K, Ruderman NB and Keaney JF (2002): Hyperglycemia increases endothelial superoxide that impairs smooth muscle cell Na^+-K^+ -ATPase activity. Am J Physiol Cell Physiol 282:C560-C566.

Guyton AC, Coleman TG, Cowley AV Jr, Scheel KW, Manning RD Jr and Norman RA Jr (1972): Arterial pressure regulation. Overriding dominance of the kidneys in long-term regulation and in hypertension. Am J Med 52:584-594.

Guzzetti S, Piccalugga E, Casati R, Cerutti S, Lombardi F, Pagani M and Malliani A (1988): Sympathetic predominance in essential hypertension: a study employing spectral analysis of heart rate variability. J Hypertens 6:711-717.

Haddy FJ (1991): Roles of sodium, potassium, calcium and natriuretic factors in hypertension. Hypertension 18:179-183.

Haim M, Tanne D, Boyko V, Reshef T, Goldbourt U, Leor J, Mekori YA and Behar S (2002): Soluble intercellular adhesion molecule-1 and long-term risk of acute coronary events in patients with chronic coronary heart disease: data from the Bezafibrate Infarction Prevention (BIP) study. J Am Coll Cardiol 39:1133-1138.

Hamlyn JM, Hamilton BP and Manunta P (1996): Endogenous ouabain, sodium balance and blood pressure: a review and a hypothesis. J Hypertens 14:151-167.

Hamsten A, de Faire U, Walldius G, Dahlen G, Szamosi A, Landou C, Blomback M and Wiman B (1987): Plasminogen activator inhibitor in plasma: risk factor for recurrent myocardial infarction. Lancet 2:3-9.

Hasdan G, Benchetrit S, Rashid G, Green J, Bernheim J and Rathaus M (2002): Endothelial dysfunction and hypertension in 5/6 nephrectomized rats are mediated by vascular superoxide. Kidney Int 61:586-590.

Hatton DC and McCarron DA (1994): Dietary calcium and blood pressure in experimental models of hypertension. A review. Hypertension 23:513-530.

Hatton DC, Scrogin KE, Levine D, Feller D and McCarron DA (1993): Dietary calcium modulates blood pressure through alpha 1-adrenergic receptors. Am J Physiol 264:F234-F238.

Hatton DC, Xue H, DeMerrit JA and McCarron DA (1994): $1,25(OH)_2$ vitamin D_3 -induced alterations in vascular reactivity in the SHR. Am J Med Sci 307:S154-S158.

Hatton DC, Yue Q and McCarron DA (1995): Mechanisms of calcium's effects on blood pressure. Semin Nephrol 15:593-602.

Haynes WB and Webb DJ (1998): Endothelin as a regulator of cardiovascular function in health and disease. J Hypertens 16:1081-1098.

Henrion D, Dowell FJ, Levy BI and Michel JB (1996): In vitro alterations of aortic vascular reactivity in hypertension induced by chronic N^G-nitro-L-arginine methyl ester. Hypertension 28:361-366.

Henrion D, Dechaux E, Dowell FJ, Maclour J, Samuel JL, Levy BI and Michel JB (1997): Alteration of flowinduced dilatation in mesenteric resistance arteries of L-NAME treated rats and its partial association with induction of cyclo-oxygenase-2. Br J Pharmacol 121:83-90.

Hermsmeyer RK (1987): Vascular muscle membrane cation mechanisms and total peripheral resistance. Hypertension 10:20-22.

Herrera-Acosta J (1994): The role of systemic and glomerular hypertension in progressive glomerular injury. Kidney Int 45:S6-S10.

Hingorani AD, Cross J, Kharbanda RK, Mullen MJ, Bhagat K, Taylor M, Donald AE, Palacios M, Griffin GE, Deanfield JE, MacAllister RJ and Vallance P (2000): Acute systemic inflammation impairs endothelium-dependent dilatation in humans. Circulation 102:994-999.

Hoebel BG, Kostner GM and Graier WF (1997): Activation of microsomal P450 mono-oxygenase by Ca2+ store depletion and its contribution to Ca2+ entry in porcine aortic endothelial cells. Br J Pharmacol 121:1579-1588.

Hori M and Karaki H (1998): Regulatory mechanisms of calcium sensitization of contractile elements in smooth muscle. Life Sci 62:1629-1633.

Horiuchi M, Akishita M and Dzau VJ (1999): Recent progress in angiotensin II type 2 receptor research in the cardiovascular system. Hypertension 33:613-621.

Horowitz A, Menice CB, Laporte R and Morgan KG (1996): Mechanisms of smooth muscle contraction. Physiol Rev 76:967-1003.

Hropot M, Grotsch H, Klaus E, Langer KH, Linz W, Wiemer G and Scholkens BA (1994): Ramipril prevents the detrimental sequels of chronic NO synthase inhibition in rats: hypertension, cardiac hypertrophy and renal insufficiency. Naunyn Schmiedebergs Arch Pharmacol 350:646-652.

Huang AH, Busse R and Bassenge E (1988): Endothelium-dependent hyperpolarization of smooth muscle cells in rabbit femoral arteries is not mediated by EDRF (nitric oxide). Naunyn-Schmiedebergs Arch Pharmacol 338:438-442.

Huang PL, Huang Z, Mashimo H, Bloch KD, Moskowitz MA, Bevan JA and Fishman MC (1995): Hypertension in mice lacking the gene for endothelial nitric oxide synthase. Nature 377:239-242.

Hutri-Kähonen N, Kähönen M, Wu X, Sand J, Nordback I, Taurio J and Pörsti I (1999): Control of vascular tone in isolated mesenteric arterial segments from hypertensive patients. Br J Pharmacol 127:1735-1743.

Ignarro LJ, Buga GM, Wood KS, Byrns RE and Chaudhuri G (1987): Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. Proc Natl Acad Sci U S A 84:9265-9269.

Ikegaki I, Hattori Y, Yamaguchi T, Sasaki Y, Satoh SI, Asano T and Shimokawa H (2001): Involvement of Rhokinase in vascular remodeling caused by long-term inhibition of nitric oxide synthesis in rats. Eur J Pharmacol 427:69-75.

Illiano S, Nagao T and Vanhoutte PM (1992): Calmidazolium, a calmodulin inhibitor, inhibits endothelium-dependent relaxations resistant to nitro-L-arginine the canine coronary artery.

Ishida-Kainouchi M, Matsuura H, Ishida T, Kajiyama G and Oshima T (1993): Platelet calcium handling in spontaneously hypertensive rats and in three strains of normotensive rats. J Hypertens 11:509-514.

Ishioka N and Bukoski RD (1999): A role for *N*-arachidonylethanolamine (anandamide) as the mediator of sensory nerve-dependent Ca^{2+} -induced relaxation. J Pharmacol Exp Ther 289:245-250.

Ishizaka H, Gudi SR, Frangos JA and Kuo L (1999): Coronary arteriolar dilatation to acidosis: role of ATP-sensitive potassium channels and pertussis toxin-sensitive G proteins. Circulation 99:558-563.

Ito S and Carratero OA (1992): Impaired response to acetylcholine despite intact endothelium-derived relaxing factor/nitric oxide in isolated microperfused afferent arterioles of the spontaneously hypertensive rats. J Cardiovasc Pharmacol 20:187-189.

Jackson WF (1998): Potassium channels and regulation of the microcirculation. Mircocirculation 5:85-90.

Jackson WF (2000): Ion channels and vascular tone. Hypertension 35:173-178.

Jackson WF and Blair KL (1998): Characterization and function of Ca2+ activated K+ channels in hamster cremasteric arteriolar muscle cells. Am J Physiol Heart and Circ Physiol 274:H27-H34.

Jackson WF, Huebner JM and Rusch NJ (1997): Enzymatic isolation and characterization of single vascular smooth muscle cells from cremasteric arterioles. Microcirculation 4:35-50.

Jaggar JH (2001): Intravascular pressure regulates local and global Ca^{2+} signalling in cerebral artery smooth muscle cells. Am J Physiol Cell Physiol 281:C439-C448.

Jaggar JH, Wellman GC, Heppner TJ, Porter VA, Perez GJ, Gollasch M, Kleppisch T, Rubart M, Stevenson AS, Lederer WJ, Knot HJ, Bonev AD and Nelson MT (1998): Ca^{2+} channels, ryanodine receptors and $Ca^{(2+)}$ -activated K⁺ channels: a functional unit for regulating arterial tone. Acta Physiol Scand 164:577-587.

Jameson M, Dai F, Lüscher T, Skopec J, Diederich A and Diederich D (1993): Endothelium-derived contracting factors in resistance arteries of young spontaneously hypertensive rats before development of overt hypertension. Hypertension 21:280-288.

Janiak P, Pillon A, Prost JF and Vilaine JP (1992): Role of angiotensin subtype 2 receptor in neointima formation after vascular injury. Hypertension 20:737-745.

Jelicks LA and Gupta RK (1990): NMR measurement of cytosolic free calcium, free magnesium, and intracellular sodium in the aorta of young spontaneously hypertensive rats before development of overt hypertension. Hypertension 21:280-288.

Jelinek J, Hackenthal E and Hackenthal R (1990): Role of the renin-angiotensin system in the adaptation to high salt intake in immature rats. J Dev Physiol 14:89-94.

Joannides R, Bakkali EH, Le Roy F, Rivault O, Godin M, Moore N, Fillastre JP and Thuillez C (1997): Altered flow-dependent vasodilation of conduit arteries in maintenance haemodialysis. Nephrol Dial Transplant 12:2623-2628.

Johns A, Freay AD, Adams DJ, Lategan TW, Ryan US and van Breemen C (1988): Role of calcium in the activation of endothelial cells. J Cardiovasc Pharmacol 12:S119-S123.

Jono S, McKee MD, Murry CE, Shioi A, Nishizawa Y, Mori K, Morii H and Giachelli C (2000): Phosphate regulation of vascular smooth muscle cell calcification. Circ Res 87:E10-E17.

Julius S (1996): The evidence for a pathophysiologic significance of the sympathetic overactivity in hypertension. Clin Exp Hypertens 18:305-321.

Kageyama Y and Bravo EL (1987): Neurohumoral and hemodynamic responses to dietary calcium supplementation in deoxycorticosterone-salt hypertensive dogs. Hypertension 9:III166-III170.

Kalliovalkama J, Jolma P, Tolvanen JP, Kähönen M, Hutri-Kähönen N, Wu X, Holm P and Pörsti I (1999): Arterial function in nitric oxide-deficient hypertension: influence of long-term angiotensin II receptor antagonism. Cardiovasc Res 42:773-782.

Kanagy NL and Webb RC (1994): Enhanced vascular reactivity to mastoparan, a G protein activator, in genetically hypertensive rats. Hypertension 23:946-950.

Kanagy NL, Ansari MN, Ghosh S and Webb RC (1994): Recycling and buffering of intracellular calcium in vascular smooth muscle from genetically hypertensive rats. J Hypertens 12:1365-1372.

Kaplan NM (1998): Clinical hypertension. Williams and Wilkins, Baltimore.

Kaplan JH (2002): Biochemistry of Na,K-ATPase. Annu Rev Biochem 71:511-535.

Karaki H and Weiss GB (1988): Calcium release in smooth muscle. Life Sci 42:111-122.

Kato N, Sugiyama T, Morita H, Nabika T, Kurihara H, Yamori Y and Yazaki Y (1999): Lack of evidence for association between the endothelial nitric oxide synthase gene and hypertension. Hypertension 33:933-936.

Katoh T, Takahashi K, Klahr S, Reyes AA and Badr KF (1994): Dietary supplementation with L-arginine ameliorates glomerular hypertension in rats with subtotal nephrectomy. J Am Soc Nephrol 4:1690-1694.

Katusic ZS (1996): Superoxide anion and endothelial regulation of arterial tone. Free Radic Biol Med 20:443-448.

Kawaguchi H, Sano H, Okada H, Iizuka K, Okamoto H, Kudo T, Murakami T and Kitabatake A (1993): Increased calcium release from sarcoplasmic reticulum stimulated by inositol triphosphate in spontaneously hypertensive rat heart cells. Mol Cell Biochem 119:51-57.

Kelly RA, O'Hara DS, Mitch WE, Steinman TI, Goldszer RC, Solomon HS and Smith TW (1986): Endogenous digitalis-like factors in hypertension and chronic renal insufficiency. Kidney Int 30:723-729.

Kielstein JT, Boger RH, Bode-Boger SM, Schaffer J, Barbey M, Koch KM and Frolich JC (1999): Asymmetric dimethylarginine plasma concentrations differ in patients with end-stage renal disease: relationship to treatment method and atherosclerotic disease. J Am Soc Nephrol 10:594-600.

Kim L, Lee T, Fu J and Ritchie ME (1999): Characterization of MAP kinase and PKC isoform and effect of ACE inhibition in hypertrophy in vivo. Am J Physiol 277:H18808-H18816.

Kimura K and Nishio I (1999): Impaired endothelium-dependent relaxation in mesenteric arteries of reduced renal mass hypertensive rats. Scand J Clin Lab Invest 59:199-204.

Kitazono T, Faraci FM, Taguchi H and Heistad DD (1995): Role of potassium channels in cerebral blood vessels. Stroke 26:1713-1723.

Knot HJ and Nelson MT (1998): Regulation of arterial diameter and wall $[Ca^{2+}]$ in cerebral arteries of rat by membrane potential and intravascular pressure. J Physiol 508:199-209.

Knot HJ, Zimmermann PA and Nelson MT (1996): Extracellular $K^{(+)}$ -induced hyperpolarizations and dilatations of rat coronary and cerebral arteries involve inward rectifier $K^{(+)}$ channels. J Physiol (Lond) 492:419-430.

Knot HJ, Standen NB and Nelson MT (1998): Ryanodine receptors regulate arterial diameter and wall $[Ca^{2+}]$ in cerebral arteries of rat via Ca^{2+} -dependent K⁺ channels. J Physiol (Lond) 508:211-221.

Kojima M, Aoki K, Asano M, Fujimoto S and Matsuda T (1991): Malfunction of arterial sarcoplasmic reticulum leading to faster and greater contraction induced by high-potassium depolarization in young spontaneously hypertensive rats. J Hypertens 9:783-788.

Korkor AB (1987): Reduced binding of $[{}^{3}H]$ -1,25-dihydroxyvitamin D₃ in the parathyroid glands of patients with chronic renal failure. New Engl J M ed 316:1573-1577.

Kotchen TA, Ott CE, Whitescarver SA, Resnick LM, Gertner JM and Blehschmidt NG (1989): Calcium and calcium regulating hormones in the "prehypertensive" Dahl salt sensitive rat (calcium and salt sensitive hypertension). Am J Hypertension 2:747-753.

Kubo T, Taguchi K and Ueda M (1998): L-type calcium channels in vascular smooth muscle cells from spontaneously hypertensive rats: effects of calcium agonist and antagonist. Hypertens Res 21:33-37.

Kubo T, Saito E, Hosokawa H, Ibusuki T, Kambe T and Fukumori R (1999): Local renin-angiotensin system and mitogen-activated protein kinase activation in rat aorta. Eur J Pharmacol 365:103-110.

Kuchan MJ and Frangos JA (1993): Shear stress regulates endothelin-1 release via protein kinase C and cGMP in cultured endothelial cells. Am J Physiol 264:H150-H156.

Kuriyama S, Kaguchi Y, Nakamura K, Hashimoto T and Sakai O (1992): Effect of serum on cell membrane Na-K transport of vascular smooth muscle in culture: a comparative study between normotensive and hypertensive rats. Pharmacol Res 25:155-165.

Kurokawa K (2001): Salt, kidney and hypertension: why and what to learn from genetic analyses? Nephron 89:369-376.

Küng CF and Lüscher TF (1995): Different mechanisms of endothelial dysfunction with ageing and hypertension in rat aorta. Hypertension 25:194-200.

Küng CF, Moreau P, Takase H and Lüscher TF (1995): L-NAME hypertension alters endothelial and smooth muscle function in rat aorta: prevention by trandolapril and verapamil. Hypertension 26:744-751.

Kwan CY and Daniel EE (1982): Arterial muscle membrane abnormalities of hydralazine-treated spontaneously hypertensive rats. Eur J Pharmacol 82:187-190.

Lacy PS, Pilkington G, Hanvesakul R, Fish HJ, Boyle JP and Thruston H (2000): Evidence against potassium as an endothelium-derived hyperpolarizing factor in rat mesenteric small arteries. Br J Pharmacol 129:605-611.

Lamb FS, Moreland RS and Webb RC (1988): Calcium and contractile responses to ouabain and potassium-free solution in aorta from spontaneously hypertensive rats. J Hypertens 6:821-828.

Law MR, Frost CD and Wald NJ (1991): By how much does dietary salt reduction lower blood pressure? III--Analysis of data from trials of salt reduction. BMJ 302:819-824.

Lee CH, Poburko D, Sahota P, Sandhu J, Ruehlmann DO and van Breemen C (2001): The mechanism of phenylepinephrine-mediated $[Ca^{2+}]$ oscillations underlying tonic contraction in the rabbit inferior vena cava. J Physiol (Lond) 534:641-650.

Lee CH, Poburko D, Kuo KH, Seow CY and van Breemen C (2002): Ca²⁺ oscillations, gradients, and homeostasis in vascular smooth muscle. Am J Physiol Heart Circ Physiol 282:H1571-H1583.

Levitsky DO, Clergue M, Lambert F, Souponitskaya MV, Le Jemtel TH, Lecarpentier Y and Lompré A-M (1993): Sarcoplasmic reticulum calcium transport and Ca^{2+} -ATPase gene expression in thoracic and abdominal aortas of normotensive and spontaneously hypertensive rats. J Biol Chem 268:8325-8331.

Li JS and Schiffrin EL (1994): Resistance artery structure and neuroeffector mechanisms in hypertension induced by inhibition of nitric oxide synthase. Am J Hypertens 7:996-1004.

Li JS, Sventek P and Schiffrin EL (1996): Effect of antihypertensive treatment and N^{ω} -nitro-L-arginine methyl ester on cardiovascular structure in deoxycorticosterone acetate-salt hypertensive rats. J Hypertens 14:1331-1340.

Li JS, Touyz RM and Schiffrin EL (1998): Effects of AT1 and AT2 angiotensin receptor antagonists in angiotensin II-infused rats. Hypertension 31:487-492.

Li PL and Campbell WB (1997): Epoxyeicosatrienoic acids activate K^+ channels in coronary smooth muscle through a guanine nucleotide binding protein. Circ Res 80:877-884.

Li YC, Kong J, Wei M, Chen ZF, Liu SQ and Cao LP (2002): 1,25-Dihydroxyvitamin D_3 is a negative endocrine regulator of the renin-angiotensin system. J Clin Invest 110(2):22-239.

Lifton RP, Gharavi AG and Geller DS (2001): Molecular mechanisms of human hypertension. Cell 104:545-556.

Ligtenberg G, Blankestijn PJ, Oey PL, Klein IH, Dijkhorst-Oei LT, Boomsma F, Wieneke GH, van Huffelen AC and Koomans HA (1999): Reduction of sympathetic hyperactivity by enalapril in patients with chronic renal failure. N Engl J Med 340:1321-1328.

Lindpaintner K, Kreutz R and Ganten D (1992): Genetic variation in hypertensive and 'control' strains. What are we controlling for anyway? Hypertension 19:428-430.

Linz W, Wiemer G, Gohlke P, Unger T and Scholkens BA (1995): Contribution of kinins to the cardiovascular actions of angiotensin-converting enzyme inhibitors. Pharmacol Rev 47:25-49.

Liu Y, Jones AW and Sturek M (1994): Increased barium influx and potassium current in stroke-prone spontaneously hypertensive rats. Hypertension 23:1091-1095.

Liu Y, Fredricks KT, Roman RJ and Lombard JH (1997a): Response of resistance arteries to reduced PO_2 and vasodilators during hypertension and elevated salt intake. Am J Physiol 273:H869-H877.

Liu Y, Pleyte K, Knaus HG and Rusch NJ (1997b): Increased expression of Ca^{2+} -sensitive K⁺ channels in aorta of hypertensive rats. Hypertension 30:1403-1409.

Llach F (1995): Secondary hyperparathyroidism in renal failure: the trade-off hypothesis revisited. Am J Kidney Dis 25:663-679.

Llach F and Forero FV (2001): Secondary hyperparathyroidism in chronic renal failure: pathogenic and clinical aspects. Am J Kidney Dis 38:S20-S33.

Locatelli F, Cannata-Andía JB, Drücke TB, Hörl WH, Fouque D, Heimburger O and Ritz E (2002): Management of disturbances of calcium and phosphate metabolism in chronic renal insufficiency, with emphasis on the control of hyperphosphataemia. Nephrol Dial Transplant 17:723-731.

Lohmeier TE (2001): The sympathetic nervous system and long-term blood pressure regulation. Am J Hypertens 14:147S-154S.

Lombard JH, Sylvester FA, Phillips SA and Frisbee JC (2003): High-salt diet impairs vascular relaxation mechanisms in rat middle cerebral arteries. Am J Physiol Heart Circ Physiol 284:H1124-H1133.

London GM, Guerin AP, Marchais SJ, Pannier B, Safar ME, Day M and Metivier F (1996): Cardiac and arterial interactions in end-stage renal disease. Kidney Int 50:600-608.

London GM (2000): Alterations of arterial function in end-stage renal disease. Nephron 84:111-118.

London GM (2003): Cardiovascular disease in chronic renal failure: pathophysiological aspects. Semin Dial 16:85-94.

Lopez-Hilker S, Dusso AS, Rapp NS, Martin KJ and Slatopolsky E (1990): Phosphorus restriction reverses hyperparathyroidism in uremia independent of changes in calcium and calcitriol. Am J Physiol 259:F432-F437.

Luckhoff A, Pohl U, Mulsch A and Busse R (1988): Differential role of extra- and intracellular calcium in the release of EDRF and prostacyclin from cultured endothelial cells. Br J Pharmacol 95:189-196.

Luik AJ, Spek JJ, Charra B, van Bortel LM, Laurent G and Leunissen KM (1997): Arterial compliance in patients on long-treatment-time dialysis. Nephrol Dial Transplant 12:2629-2632.

Luke RG (1998): Chronic renal failure - a vasculopathic state. N Engl J Med 339:841-843.

Lüscher TF and Noll G (1995): The pathogenesis of cardiovascular disease: role of the endothelium as a target and mediator. Atherosclerosis 118:81-90.

Lüscher TF and Vanhoutte PM (1986): Endothelium-dependent contractions to acetylcholine in the aorta of the spontaneously hypertensive rat. Hypertension 8:344-348.

Lüscher TF, Boulanger CM, Dohi Y and Yang Z (1992): Endothelium-derived contracting factors. Hypertension 19:117-130.

Lüscher TF, Oemar BS, Boulanger CM and Hahn AWA (1993a): Molecular and cellular biology of endothelin and its receptors-Part I. J Hypertens 11:7-11

Lüscher TF, Oemar BS, Boulanger CM and Hahn AWA (1993b): Molecular and cellular biology of endothelin and its receptors-Part II. J Hypertens 11:121-126.

Lüscher TF, Seo B and Bühler FR (1993c): Potential role of endothelin in hypertension. Controversy on endothelin in hypertension. Hypertension 21:752-757.

MacAllister RJ, Rambausek MH, Vallance P, Williams D, Hoffmann KH and Ritz E (1996): Concentration of dimethyl-L-arginine in the plasma of patients with end-stage renal failure. Nephrol Dial Transplant 11:2449-2452.

MacGregor GA (1998): Salt: blood pressure, the kidney, and other harmful effects. Nephrol Dial Transplant 13:2471-2479.

MacGregor GA and Sever PS (1996): Salt--overwhelming evidence but still no action: can a consensus be reached with the food industry? CASH (Consensus Action on Salt and Hypertension). BMJ 312:1287-1289.

Mancia G, Grassi G, Giannattasio C and Seravalle G (1999): Sympathetic activation in the pathogenesis of hypertension and progression of organ damage. Hypertension 34:724-728.

Manjeet S and Sim MK (1987): Decreased Na⁺K⁺ATPase activity in the aortic endothelium and smooth muscle of the spontaneously hypertensive rats. Clin Exp Hypertens 9:797-812.

Marchenko SM and Sage SO (1996): Calcium-activated potassium channels in the endothelium of intact rat aorta. J Physiol 492:53-60.

Marescau B, Nagels G, Possemiers I, De Broe ME, Because I, Billiouw J-M, Lornoy W and De Deyn PP (1997): Guanidino compounds in serum and urine of nondialyzed patients with chronic renal insufficiency. Metabolism 46:1024-1031.

Marco MP, Craver L, Betriu A, Belart M, Fibla J and Fernández E (2003): Higher impact of mineral metabolism on cardiovascular mortality in a European hemodialysis population. Kidney Int 63(Suppl 85):S111-S114.

Marks AR (1992): Calcium channels expressed in smooth muscle. Circulation 86:61-67.

Martens JR and Gelband CH (1996): Alterations in rat interlobar artery membrane potential and K⁺ channels in genetic and nongenetic hypertension. Circ Res 79:295-301.

Martin-Mateo MC, del Canto-Jafiez E and Barrero-Martinez MJ (1998): Oxidative stress and enzyme activity in ambulatory renal patients undergoing continuous peritoneal dialysis. Ren Fail 20:117-124.

Martonosi AN, Jona I, Molnar E, Seidler NW, Buchet R and Varga S (1990): Emerging views on the structure and dynamics of the Ca^{2+} -ATPase in sarcoplasmic reticulum. FEBS Lett 268:365-370.

Masugi F, Ogihara T, Hasegawa T and Kumahara Y (1987): Ouabain-like and non-ouabain-like factors in plasma of patients with essential hypertension. Clin Exp Hypertens 9:1233-1242.

Matsubara BB, Matsubara LS, Zornoff LA, Franco M and Janicki JS (1998): Left ventricular adaptation to chronic pressure overload induced by inhibition of nitric oxide synthase in rats. Basic Res Cardiol 93:173-181.

McCarron DA (1997): Role of adequate dietary calcium intake in the prevention and management of saltsensitive hypertension. Am J Clin Nutr 65(Suppl 1):712S-716S.

McCarron DA, Lucas PA, Shneidman RJ, LaCour B and Drueke T (1985): Blood pressure development of the spontaneously hypertensive rat after concurrent manipulations of dietary Ca^{2+} and N^+ . Relation to intenstinal Ca^{2+} fluxes. J Clin Invest 76(3):1147-1154.

McCarron DA, Morris C and Cole C (1982): Dietary calcium in human hypertension. Science 217:267-269.

McCarron DA, Morris CD, Henry HJ and Stanton JL (1984): Blood pressure and nutrient intake in the United

States: an analysis of the Health and Nutrition Examination Survey I. Science 224:1392-1398.

McCarron DA and Reusser ME (1999): Finding consensus in the dietary calcium-blood pressure debate. J Am Coll Nutr 18:398S-405S.

McCarron DA, Yung NN, Ugoretz BA and Krutzik S (1981): Disturbances of calcium metabolism in the spontaneously hypertensive rat. Hypertension 3:1165-1167.

McCulloch AI, Bottrill FE, Randall MD and Hiley CR (1997): Characterization and modulation of EDHFmediated relaxations in the rat isolated superior mesenteric arterial bed. Br J Pharmacol 120:1431-1438.

McDonough AA, Leong PK and Yang LE (2003): Mechanisms of pressure natriuresis: how blood pressure regulates renal sodium transport. Ann N Y Acad Sci 986:669-677.

Mcguire JJ, Ding H and Triggle CR (2001): Endothelium-derived relaxing factors: a focus on endothelium-derived hyperpolarizing factor(s). Can J Physiol Pharmacol 79:443-470.

Meffert S, Stoll M, Steckelings UM, Bottari SP and Unger T (1996): The angiotensin II AT₂ receptor inhibits proliferation and promotes differentiation in PC12W cells. Mol Cell Endocrinol 122:59-67.

Merke J, Hugel U, Zlotkowski A, Szabo A, Bommer J, Mall G and Ritz E (1987): Diminished parathyroid 1,25(OH)₂D₃ receptor in experimental uremia. Kidney Int 32:350-353.

Mihai R and Farndon JR (2000): Parathyroid disease and calcium metabolism. Br J Anaesthesia 85:29-43.

Minneman KP (1988): Alpha 1-adrenergic receptor subtypes, inositol phosphates, and sources of cell Ca²⁺. Pharmacol Rev 40:87-119.

Mishra SK and Hermsmeyer K (1994): Selective inhibition of T-type Ca2+ channels by Ro 40-5967. Circ Res 75:144-148.

Mombouli JV, Illiano S, Nagao T, Scott-Burden T and Vanhoutte PM (1992): Potentiation of bradykinininduced relaxations by perindoprilat in canine coronary arteries involves both nitric oxide and endotheliumderived hyperpolarizing factor. Circ Res 71:137-144.

Moncada S and Higgs A (1993): The L-arginine-nitric oxide pathway. N Engl J Med 329:2002-2012.

Monteith GR, Kaple EP, Chen S and Roufogalis BD (1996): Plasma membrane calcium pump-mediated calcium efflux and bulk cytosolic free calcium in cultured aortic smooth muscle cells from spontaneously hypertensive and Wistar-Kyoto normotensive rats. J Hypertens 14:435-442.

Monteith GR, Kaple EP, Kuo TH and Roufogalis BD (1997): Elevated plasma membrane and sarcoplasmic reticulum Ca^{2+} pump mRNA levels in cultured aortic smooth muscle cells from spontaneously hypertensive rats. Biochem Biophys Res Comm 230:344-346.

Morishita R, Gibbons G, Ellison KE and Dzau VJ (1994): Evidence for direct local effect of Ang II in vascular hypertrophy. J Clin Invest 94:978-984.

Morris STW, McMurray JJV, Spiers A and Jardine AG (2001): Impaired endothelial function in isolated human uremic resistance arteries. Kidney Int 60:1077-1082.

Morrisey JJ and Klahr S (1999): Effect of AT2 receptor blockade on the pathogenesis of renal fibrosis. Am J Physiol 276:F39-F45.

Mosterd A, D'Agostini RB, Silbershartz H, Sytkowski PA, Kannel WB, Grobbee DE and Levy D (1999): Trends in the prevalence of hypertension, antihypertensive therapy, and left ventricular hypertrophy from 1950 to 1989. N Engl J Med 340:1221-1227.

Mourad JJ, Girerd X, Boutouyrie P, Laurent S, Safar M and London G (1997): Increased stiffness of radial artery wall material in end-stage renal disease. Hypertension 30:1425-1430.

Muiesan ML, Salvetti M, Monteduro C, Rizzoni D, Zulli R, Corbellini C, Brun C and Agabiti-Rosei E (1999): Effect of treatment on flow-dependent vasodilatation of the brachial artery in essential hypertension. Hypertension 33:575-580.

Mulrow PJ and Franco-Saenz R (1996): The adrenal renin-angiotensin system: a local hormonal regulator of aldosterone production. J Hypertens 14:173-176.

Mulvany MJ (1999): Vascular remodelling of resistance vessels: can we define this? Cardiovasc Res 41:9-13.

Mulvany MJ and Halpern W (1977): Contractile properties of small arterial resistance vessels in spontaneously hypertensive and normotensive rats. Circ Res 41:19-26.

Mulvany MJ, Persson AEG and Andersen J (1991): No persistent effect of angiotensin converting enzyme inhibitor treatment in Milan hypertensive rats despite regression of vascular structure. J Hypertens 9:589-593.

Mupanomunda MM, Ishioka N and Bukoski RD (1999): Interstitial Ca^{2+} undergoes dynamic changes sufficient to stimulate nerve-dependent Ca^{2+} -induced relaxation. Am J Physiol 276:H1035-H1042.

Mäkynen H, Kähönen M, Arvola P, Wu X, Wuorela H and Pörsti I (1996): Endothelial function in deoxycorticosterone-NaCl hypertension: effect of calcium supplementation. Circulation 93:1000-1008.

Nagao T and Vanhoutte PM (1992): Hyperpolarisation as a mechanism for endothelium-dependent relaxations in the porcine coronary artery. J Physiol 445:355-367.

Nahorski SK, Wilcox RA, Mackrill JJ and Chaliss RAJ (1994): Phosphoinositide-derived second messangers and the regulation of Ca^{2+} in vascular smooth muscle. J Hypertens 12:133-143.

Nakajima M, Hutchinson HG, Fujinaga M, Hayashida W, Morishita R, Zhang L, Horiuchi M, Pratt RE and Dzau VJ (1995): The angiotensin II type 2 (AT₂) receptor antagonizes the growth effect of the AT₁ receptor: gain-of-function study using gene transfer. Proc Natl Acad Sci USA 92:10663-10667.

Nava E and Lüscher TF (1995): Endothelium-derived vasoactive factors in hypertension: nitric oxide and endothelin. J Hypertens 13:S39-S48.

Navar LG (1997): The kidney in blood pressure regulation and development of hypertension. Med Clin North Am 81:1165-1198.

Nelson MT (1993): Ca^{2+} -activated potassium channels and ATP-sensitive potassium channels as modulators of vascular tone. Trends Cardiovasc Med 3:54-60.

Nelson MT, Patlak JB, Worley JF and Standen NB (1990): Calcium channels, potassium channels, and voltage dependence of arterial smooth muscle tone. Am J Physiol 259:C3-C18.

Nelson MT and Quayle JM (1995): Physiological roles and properties of potassium channels in arterial smooth muscle. Am J Physiol 268:C799-C822.

Nelson MT, Cheng H, Rubart M, Santana LF, Bonev AD, Knot HJ and Lederer WJ (1995): Relaxation of arterial smooth muscle by calcium sparks. Science 270:633-637.

Neusser M, Tepel M, Golinski P, Holthues J, Spieker C, Zhu Z and Zidek W (1994): Different calcium storage pools in vascular smooth muscle cells from spontaneously hypertensive and normotensive Wistar-Kyoto rats. J Hypertens 12:533-538.

Nguyen PV, Yang X-P, Li G, Deng LY, Flückiger J-P and Schiffrin EL (1993): Contractile responses and signal transduction of endothelin-1 in aorta and mesenteric vasculature of adult spontaneously hypertensive rats. Can J Physiol Pharmacol 71:473-483.

Ni Z and Vaziri ND (2001): Effect of salt loading on nitric oxide synthase expression in normotensive rats. Am J Hypertens 14:155-163.

NIH Consensus Panel (1994). NIH Consensus Conference. Optimal calcium intake. NIH consensus development panel on optimal calcium intake. JAMA 272:1942-1948.

Nilius B and Droogmans G (2001): Ion channels and their functional role in vascular endothelium. Physiol Rev 81:1415-1459.

Nio Y, Matsubara H, Murasawa S, Kanasaki M and Inada M (1995): Regulation of gene transcription of angiotensin II receptor subtypes in myocardial infarction. J Clin Invest 95:46-54.

Nishizuka Y (1995): Protein kinase C anad lipid signaling for sustained cellular responses. FASEB 9:484-496.

Noll G, Wenzel RR, Schneider M, Oesch V, Binggeli C, Shaw S, Weidman P and Luscher TF (1996): Increased activation of sympathetic nervous system and endothelin by mental stress in normotensive offspring of hypertensive parents. Circulation 93:866-869.

Noma A (1983): ATP-regulated K^+ channels in cardiac muscle. Nature 305:147-148.

Noris M, Benigni A, Boccardo P, Aiello S, Gaspari F, Todeschini M, Figliuzzi M and Remuzzi G (1993): Enhanced nitric oxide synthesis in uremia: Implications for platelet dysfunction and dialysis hypotension. Kidney Int 44:445-450.

Numaguchi K, Egashira K, Sakata M, Shimokawa H and Takeshita A (1996): Coronary vascular ATP-sensitive potassium channels are activated to a greater extent in spontaneously hypertensive rats than in Wistar-Kyoto rats. J Hypertens 14:183-189.

Nussberger J, Cugno M, Amstutz C, Cicardi M, Pellacani A and Agostoni A (1998): Plasma bradykinin in angio-oedema. Lancet 251:1693-1697.

O'Donnell ME and Owen NE (1994): Regulation of ion pumps and carriers in vascular smooth muscle. Physiol Rev 74:683-721.

Ohara Y, Peterson TE and Harrison DG (1993): Hypercholesterolemia increases endothelial superoxide anion production. J Clin Invest 91:2546-2551.

Ohkubo N, Matsubara H, Nozawa Y, Mori Y, Murasawa S, Kijima K, Maruyama K, Masaki H, Tsutumi Y, Shibazaki Y, Iwasaka T and Inada M (1997): Angiotensin type 2 receptors are re-expressed by cardiac fibroblasts from failing myopathic hamster hearts and inhibit cell growth and fibrillar collagen metabolism. Circulation 96:3954-3962.

Ohya Y, Setoguchi M, Fujii K, Nagao T, Abe I and Fujishima M (1996): Impaired action of levcromakalim on ATP-sensitive K^+ channels in mesenteric artery cells from spontaneously hypertensive rats. Hypertension 27:1234-1239.

Okamura K, Kondo J, Yoshino M, Ishikawa K, Asano H, Hashimoto H and Ito T (1992): Enalapril reduces the enhanced 1,2-diacylglycerol content and RNA synthesis in spontaneously hypertensive rat hearts before established hypertension. Mol Cell Biochem 112:15-21.

Okazaki T, Zajac JD, Igarashi T, Ogata E and Kronenberg HM (1991): Negative regulatory elements in the human parathyroid hormone gene. J Biol Chem 266:21903-21910.

Omland T, Lie RT, Aakvaag A, Aarsland T and Dickstein K (1994): Plasma endothelin determination as a prognostic indicator of 1-year mortality after acute myocardial infarction. Circulation 89:1573-1579.

Orlov S, Resink TJ, Bernhardt J and Bühler FR (1992): Na^+-K^+ pump and Na^+-K^+ co-transport in cultured vascular smooth muscle cells from spontaneously hypertensive and normotensive rats: baseline activity and regulation. J Hypertens 10:733-740.

Orlov SN, Taurin S, Tremblay J and Hamet P (2001): Inhibition of Na+,K+ pump affects nucleic acid synthesis and smooth muscle cell proliferation via elevation of the [Na+]i/[K+]I ratio: possible implication in vascular remodelling. J Hypertens 19:1559-1565.

Oshima T, Young EW and McCarron DA (1991): Abnormal platelet and lymphocyte calcium handling in prehypertensive rats. Hypertension 18:111-115.

Palmer RM, Ashton DS and Moncada S (1988). Vascular endothelial cells synthesize nitric oxide from Larginine. Nature 333:664-666.

Panfilov VV and Reid JL (1994): Brain and autonomic mechanisms in hypertension. J Hypertens 12:337-343.

Panza JA, Quyyumi AA, Brush Jr. JE and Epstein SE (1990): Abnormal endothelium-dependent vascular relaxation in patients with essential hypertension. N Engl J Med 323:22-27.

Papageorgiu P and Morgan KG (1991): Intracellular free Ca^{2+} is elevated in hypertrophic aortic muscle from hypertensive rats. Am J Physiol 260:H507-H515.

Paterno R, Faraci FM and Heistad DD (1996): Role of $Ca^{(2+)}$ -dependent K⁺ channels in cerebral vasodilatation induced by increases in cyclic GMP and cyclic AMP in the rat. Stroke 27:1603-1607.

Peng H, Matchkov V, Ivarsen A, Aalkjaer C and Nilsson H (2001): Hypothesis for the initiation of vasomotion. Circ Res 88:810-815.

Perry PA and Webb RC (1991): Agonist-sensitive calcium stores in arteries from steroid hypertensive rats. Hypertension 17:603-611.

Peters H and Noble NA (1996): Dietary L-Arginine in renal disease. Semin Nephrol 16:567-575.

Peuler JD, Morgan DA and Mark AL (1987): High calcium diet reduces blood pressure in Dahl salt sensitive rats by neural mechanisms. Hypertension 9:III159-III165.

Pidgeon GB, Lewis LK, Yandle TG, Richards AM and Nicholls MG (1996): Endogenous ouabain, sodium balance and blood pressure. J Hypertens 14:169-171.

Pörsti I, Wuorela H, Arvola P, Mammi P, Nurmi AK, Koistinaho J, Laippala P and Vapaatalo H (1991): Effects of calcium supplementation and deoxycorticosterone on plasma atrial natriuretic peptide and electrolyte excretion in spontaneously hypertensive rats. Acta Physiol Scand 141:343-350.

Porter VA, Bonev AD, Knot HJ, Heppner TJ, Stevenson AS, Kleppisch T, Lederer WJ and Nelson MT (1998): Frequency modulation of Ca²⁺ sparks is involved in regulation of arterial diameter by cyclic nucleotides. Am J Physiol 274:C1346-C1355.

Prasad A, Zhu J, Halcox JP, Waclawiw MA, Epstein SE and Quyyumi AA (2002): Predisposition to atherosclerosis by infections: role of endothelial dysfunction. Circulation 106:184 -190.

Preston RA (1999): Renoprotective effects of antihypertensive drugs. Am J Hypertens 12:19-32.

Prior HM, Webster N, Quinn K, Beech DJ and Yates MS (1998): K(+)-induced dilation of a small renal artery: no role for inward rectifier K+ channels. Cardiovasc Res 37:780-790.

Quast U, Guillon J-M and Cavero I (1994): Cellular pharmacology of potassium channel openers in vascular smooth muscle. Cardiovasc Res 28:805-810.

Quayle JM, Nelson MT and Standen NB (1997): ATP-sensitive and inwardly rectifying potassium channels in smooth muscle. Physiol Rev 77:1165-1232.

Quilley J, Fulton D and McGiff JC (1997): Hyperpolarizing factors. Biochem Pharmacol 54:1059-1070.

Rabelink TJ and Koomans HA (1997): Endothelial function and the kidney. An emerging target for cardiovascular therapy. Drugs 53:S11-S19.

Radaelli A, Mircoli L, Mori I, Mancia G and Ferrari AU (1998): Nitric oxide dependent vasodilation in young spontaneously hypertensive rats. Hypertension 32:735-739.

Rahn KH (1998): Renal function in treated and untreated hypertension. J Hum Hypertens 12:599-601.

Rahn KH, Barenbrock M and Hausberg M (1999): The sympathetic nervous system in the pathogenesis of hypertension. J Hypertens 17:S11-S14.

Randall MD and McCulloch AI (1995): The involvement of ATP-sensitive potassium channels in β -adrenoceptor-mediated vasorelaxation in the rat isolated mesenteric arterial bed. Br J Pharmacol 115:607-612.

Rayson BM and Gilbert MT (1992): Regulation of Na⁺,K⁺-ATPase in hypertension. Semin Nephrol 12:72-75.

Redondo J, Peiró C, Rodriguez-Mañas L, Salaices M, Marín J and Sánchez-Ferrer CF (1995): Endothelial stimulation of sodium pump in cultured vascular smooth muscle. Hypertension 26:177-185.

Reid JL (1994): Hypertension and the brain. Br Med Bull 50:371-380.

Rembold CM (1992): Regulation of contraction and relaxation in arterial smooth muscle. Hypertension 20:129-137.

Resnick LM, Laragh JH, Sealey JE and Alderman MH (1983): Divalent cations in essential hypertension. Relations between serum ionized calcium, magnesium and plasma renin activity. N Engl J Med 309:888-891.

Ribeiro MO, Antunes E, De Nucci G, Lovisolo SM and Zatz R (1992): Chronic inhibition of nitric oxide synthesis, a new model of arterial hypertension. Hypertension 20:298-303.

Ridker PM, Hennekens CH, Roitman-Johnson B and Allen J (1998): Plasma concentration of soluble intercellular adhesion molecule 1 and risks of future myocardial infarction in apparently healthy men. Lancet 351:88-92.

Rinaldi G and Bohr D (1989): Endothelium-mediated spontaneous response in aortic rings of deoxycorticosterone acetate-hypertensive rats. Hypertension 13:256-261.

Riordan JF (1995): Angiotensin II: Biosynthesis, molecular recognition, and signal transduction. Cell Mol Neurobiol 15:637-651.

Ritter JM, Barrow SE, Doktor HS, Stratton PD, Edwards JS, Henry JA and Gould S (1993): Thromboxane A₂ receptor antagonism and synthase inhibition in essential hypertension. Hypertension 22:197-203.

Ritz E and Tarng DC (2001): Renal disease in type 2 diabetes. Nephrol Dial Transplant 16:S11-S18.

Rizzoni D, Porteri E, Castellano M, Bettoni G, Muiesan ML, Tiberio G, Giulini SM, Rossi G, Bernini G and Agabiti-Rosei E (1998): Endothelial dysfunction in hypertension is independent from the etiology and from vascular structure. Hypertension 31:335-341.

Roman RJ, Maier KG, Sun CW, Harder DR and Alonso-Galicia M (2000): Renal and cardiovascular actions of 20-hydroxyeicosatetraenoic acid and epoxyeicosatrienoic acids. Clin Exp Pharmacol Physiol 27:855-865.

Rostand SG and Drücke TB (1999): Parathyroid hormone, vitamin D, and cardiovascular disease in chronic renal failure. Kidney Int 56:383-392.

Rothermund L and Paul M (1998): Hypertension and the renin-angiotensin system – evidence from genetic and transgenic studies. Basic Res Cardiol 93:1-6.

Rubanyi GM (1993): The role of endothelium in cardiovascular homeostasis and diseases. J Cardiovasc Pharmacol 22:1-14.

Ruegg JC (1999): Smooth muscle: PKC-induced Ca^{2+} sensitisation by myosin phosphatase inhibition. J Physiol 520:3.

Ruehlmann DO, Lee CH, Poburko D and van Breemen C (2000): Asynchronous Ca^{2+} waves in intact venous smooth muscle. Circ Res 86:e72-e79.

Ruoff GE (1998): The impact of nonsteroidal anti-inflammatory drugs on hypertension: alternative analgesics for patients at risk. Clin Ther 20:376-387

Rusch NJ and Liu Y (1997): Potassium channels in hypertension: homeostatic pathways to buffer arterial contraction. J Lab Clin Med 130:245-251.

Sada T, Koike H, Ikeda M, Sato K, Ozaki H and Karaki H (1990): Cytosolic free calcium of aorta in hypertensive rats. Chronic inhibition of angiotensin converting enzyme. Hypertension 16:245-251.

Sakamoto H, Nakamura T, Akuzawa N, Masuda H, Sumino H, Saito Y, Ohyama Y, Kurashina T, Tamura J and Kurabayashi M (2002): Reciprocal expression of vascular endothelial growth factor and nitric oxide synthase by coronary arterial wall cells during inhibition of nitric oxide synthesis in rats. Nephron 92:472-474.

Sakata K, Shirotani M, Yoshida H and Kurata C (1999): Cardiac sympathetic nervous system in early essential hypertension assessed by 1231-MIBG. J Nucl Med 40:6-11.

Sanders PW (1996): Salt-sensitive hypertension: lessons from animal models. Am J Kidney Dis 28:775-782.

Sandow SL and Hill CE (2000): Incidence of myo-endothelial gap junctions in the proximal and distal mesenteric arteries of the rat is suggestive of a role in endothelium-derived hyperpolarizing factor-mediated responses. Circ Res 86:341-346.

Schiffrin EL (2001): A critical review of the role of endothelial factors in the pathogenesis of hypertension. J Cardiovasc Pharmacol 38:S3-S6.

Schiffrin EL (1995a): Endothelin in hypertension. Curr Opin Cardiol 10:485-494.

Schiffrin EL (1995b): Endothelin: Potential role in hypertension and vascular hypertrophy. Hypertension 25:1135-1143.

Schiffrin EL (1997): Resistance arteries as endpoints in hypertension. Blood Press Suppl 2:24-30.

Schiffrin EL (2001a): A critical review of the role of endothelial factors in the pathogenesis of hypertension. J Cardiovasc Pharmacol 38:S3-S6.

Schiffrin EL (2001b): Role of endothelin-1 in hypertension and vascular disease. Am J Hypertens 14:83S-89S.

Schmid C, Castrop H, Reitbauer J, Della Bruna R and Kurtz A (1997): Dietary salt intake modulates angiotensin II type 1 receptor gene expression. Hypertension 29:923-929.

Schmidt RF and Baylis C (2000): Total nitric oxide production is low in patients with chronic renal disease. Kidney Int 58:1261-1266.

Schimieder RE, Weilprecht H, Schobel H, John S, Weidinger G, Gatzka C and Veelken R (1997): Is endothelial function of the radial artery altered in human essential hypertension? Am J Hypertens 10:323-331.

Shah J and Jandhyala BS (1995): Age-dependent alterations in Na+,K(+)-ATPase activity in the central nervous system of spontaneously hypertensive rats: relationship to the development of high blood pressure. Clin Exp Hypertens 17(5):751-767.

Shimokawa H, Yasutake H, Fujii K, Owada MK, Nakaike R, Fukumoto Y, Takayanagi Y, Nagao T, Egashira K, Fujishima M and Takeshita A (1996): The importance of the hyperpolarizing mechanism increases as the vessel

size decrease in endothelium-dependent relaxations in rat mesenteric circulation. J Cardiovasc Pharmacol 28:703-711.

Shoji T, Emoto M, Tabata T, Kimoto E, Shinohara K, Maekawa K, Kawagishi T, Tahara H, Ishimura E and Nishizawa Y (2002): Advanced atherosclerosis in predialysis patients with chronic renal failure. Kidney Int 61:2187-2192.

Silver J, Kilav R and Naveh-Many T (2002): Mechanisms of secondary hyperparathyroidism. Am J Physiol Renal Physiol 283:F367-F376.

Siewert-Delle A, Ljungman S, Andersson OK and Wilhelmsen L (1998): Does treated primary hypertension lead to end-stage renal disease? A 20-year follow-up of the Primary Prevention Study in Göteborg, Sweden. Nephrol Dial Transplant 13:3084-3090.

Siragy HM and Carey RM (1996): The subtype-2 (AT2) angiotensin receptor regulates renal cyclic guanosine 3',5'-monophosphate and AT1 receptor-mediated prostaglandin E2 production in conscious rats. J Clin Invest 97:1978-1982.

Siragy HM, Inagami T, Ichiki T and Carey RM (1999): Sustained hypersensitivity to angiotensin II and its mechanism in mice lacking the subtype-2 (AT2) angiotensin receptor. Proc Natl Acad Sci USA 96:6506-6510.

Slatopolsky E (2003): New developments in hyperphosphatemia management. J Soc Am Nephrol 14:S297-S299.

Slatopolsky E, Brown A and Dusso A (1999): Pathogenesis of secondary hyperparathyroidism. Kidney Int 56:S14-S19.

Slatopolsky E, Brown A and Dusso A (2001): Role of phosphorus in the pathogenesis of secondary hyperparathyroidism. Am J Kidney Dis 37:S54-S57.

Sleight P (1991): Role of the baroreceptor reflexes in circulatory control, with particular reference to hypertension. Hypertension 18(5Suppl):III31-III34.

Somlyo AP, Wu X, Walker LA and Somlyo AV (1999): Pharmacomechanical coupling: the role of calcium, G-proteins, kinases and phosphatases. Rev Physiol Biochem Pharmacol 134:201-234.

Song Y and Simard JM (1995): β -Adrenoceptor stimulation activates large-conductance Ca²⁺-activated K⁺ channels in smooth muscle cells from basilar artery of guinea pig. Plügers Arch 430:983-993.

Souza HC, Ballejo G, Salgado MC, Da Silva VJ and Salgado HC (2001): Cardiac sympathetic overactivity and decreased baroreflex sensitivity in L-NAME hypertensive rats. Am J Physiol Heart Circ Physiol 280:H844-H850.

Spedding M and Paoletti R (1992): Classification of calcium channels and the sites of action of drugs modifying channel function. Pharmacol Rev 44:363-376.

Spieker C, Heck D, Zidek W and Vetter H (1986): Ca^{2+} metabolism in arteries of spontaneously hypertensive rats: assessment by proton-induced X-ray emission. J Hypertens 4:122-125.

Standen NB and Quayle JM (1998): K^+ channel modulation in arterial smooth muscle. Acta Physiol Scan 164:549-557.

Stauss HM (2002): Baroreceptor reflex function. Am J Physiol Regulatory Integrative Comp Physiol 283:R284-R286.

Steinvinkel P, Heimburger O, Paultre F, Diczfalusy U, Wang T, Berglund L and Jogestrand T (1999): Strong association between malnutrition, inflammation and atherosclerosis in chronic renal failure. Kidney Int 55:1899-1911.

Stock P, Liefeldt L, Paul M and Ganten D (1995): Local renin-angiotensin systems in cardiovascular tissues: localization and functional role. Cardiology 86:2-8.

Stoll M, Steckelings M, Paul M, Bottari SP, Metzger R and Unger T (1995) The angiotensin AT2-receptor mediates inhibition of cell proliferation in coronary endothelial cells. J Clin Invest 95:651-657.

Storm DS, Stuenkel EL and Webb RC (1992): Calcium channel activation in arterioles from genetically hypertensive rats. Hypertension 20:380-388.

Stull JT, Gallagher PJ, Herring BP and Kamm KE (1991): Vascular smooth muscle contractile elements. Cellular regulation. Hypertension 17:723-732.

Sugiyama T, Yoshizumi M, Takaku F and Yazaki Y (1990): Abnormal calcium handling in vascular smooth muscle of spontaneously hypertensive rats. J Hypertens 8:369-375.

Sunano S, Watanabe H, Tanaka S, Sekiguchi F and Shimamura K (1999): Endothelium-derived relaxing, contracting and hyperpolarizing factors of mesenteric arteries of hypertensive and normotensive rats. Br J Pharmacol 126:709-716.

Suo M, Kalliovalkama J, Pörsti I, Jolma P, Tolvanen JP, Vuolteenaho O and Ruskoaho H (2002): N(G)-nitro-Larginine methyl ester-induced hypertension and natriuretic peptide gene expression: inhibition by angiotensin II type 1 receptor antagonism. J Cardiovascular Pharmcol 40:478-486.

Svetky LP, Simons-Morton D, Vollmer WM, Appel LJ, Conlin PR, Ryan DH, Ard J, Kennedy BM for the DASH Research Group (1999): Effects of dietary patterns on blood pressure. Subgroup analysis of the Dietary Approaches to Stop Hypertension (DASH) randomised clinical trial. Arch Intern Med 159:285-293.

Sylverster FA, Stepp DW, Frisbee JC and Lombard JH (2002): High-salt diet depresses acetylcholine reactivity proximal to NOS activation in cerebral arteries. Am J Physiol Heart Circ Physiol 283:H353-H363.

Szabo A, Merke J, Beier E, Mall G and Ritz E (1989): 1,25(OH)2 vitamin D3 inhibits parathyroid cell proliferation in experimental uremia. Kidney Int 35:1049-1056.

Taddei S, Virdis A, Mattei P and Salvetti A (1993): Vasodilation to acetylcholine in primary and secondary forms of human hypertension. Hypertension 21:929-933.

Taddei S, Virdis A, Mattei P, Ghiadoni L, Sudano I and Salvetti A (1996): Defective L-arginine-nitric oxide pathway in offspring of essential hypertensive patients. Circulation 94:1298-1303.

Taddei S, Virdis A, Mattei P, Ghiadoni L, Fasolo CB, Sudano I and Salvetti A (1997a): Hypertension causes premature ageing of endothelial function in humans. Hypertension 29:736-743.

Taddei S, Virdis A, Ghiadoni L, Magagna A and Salvetti A (1997b): Cyclooxygenase inhibition restores nitric oxide activity in essential hypertension. Hypertension 29:274-279.

Taddei S, Virdis A, Ghiadoni L and Salvetti A (2000): Vascular effects of endothelin-1 in essential hypertension: relationship with cyclooxygenase-derived endothelium-dependent contracting factors and nitric oxide. J Cardiovasc Pharmacol 35:S37-S40.

Taddei S, Virdis A, Ghiadoni L, Sudano I, Magagna A and Salvetti A (2001): Role of endothelin in the control of peripheral vascular tone in human hypertension. Heart Fail Rev 6:277-285.

Takami S, Wong ZY, Stebbing M and Harrap SB (1999): Linkage analysis of endohthelial nitric oxide synthase gene with human blood pressure. J Hypertens 17:1431-1436.

Takase H, Dohi Y, Kojima M and Sato K (1994): Changes in the endothelial cyclo-oxygenase pathway in resistance arteries of spontaneously hypertensive rats. J Cardiovasc Pharmacol 23:326-330.

Takahashi E and Berk BC (1998): MAP kinases and vascular smooth muscle function. Acta Physiol Scand 164:611-621.

Takemoto M, Egashira K, Usui M, Numaguchi K, Tomita H, Tsutsui H, Shimokawa H, Sueshi K and Takeshita A (1997): Important role of tissue angiotensin-converting enzyme activity in the pathogenesis of coronary vascular and myocardial structural changes induced by long-term blockade of nitric oxide synthesis in rats. J Clin Invest 99:278-287.

Takuwa Y (1996): Regulation of vascular smooth muscle contraction. The roles of Ca^{2+} , protein kinase C and myosin light chain phosphatase. Jpn Heart J 37:793-813.

Tamura H, Hopp L, Kino M, Tokushige A, Searle BM, Khalil F and Aviv A (1986): Na^+-K^+ regulation in cultured vascular smooth muscle cell of the spontaneously hypertensive rat. Am J Physiol 250:C939-947.

Tanoue A, Koba M, Miyawaki S, Koshimizu TA, Hosoda C, Oshikawa S and Tsujimoto G (2002): Role of the alpha1D-adrenegric receptor in the development of salt-induced hypertension. Hypertension 40:101-106.

Taubes G (1998): The (political) science of salt. Science 281:898-901.

Taylor SG, Southerton JS, Weston AH and Baker JR (1988): Endothelium-dependent effects of acetylcholine in rat aorta: a comparison with sodium nitroprusside and cromakalim. Br J Pharmacol 94:853-863.

Tesfamariam B and Ogletree ML (1995): Dissociation of endothelial cell dysfunction and blood pressure in SHR. Am J Physiol 269:H189-H194.

Thambyrajah J, Landray MJ, McGlynn FJ, Jones HJ, Wheeler DC and Townend JN (2000): Abnormalities of endothelial function in patients with predialysis renal failure. Heart 83:205-209.

Thogersen AM, Jansson J, Boman K, Nilsson TK and Weinehall L (1998): High plasminogen activator inhibitor and tissue plasminogen activator levels in plasma precede a first acute myocardial infarction in both men and women: evidence for the fibrinolytic system as an independent primary risk factor. Circulation 98:2241-2247.

Thompson LE, Rinaldi GJ and Bohr DF (1990): Decreased activity of the sodium-calcium exchanger in tail artery of stroke-prone spontaneously hypertensive rats. Blood Vessels 27:197-201.

Thompson SG, Kienast J, Pyke SD, Haverkate F and van de Loo JCW (1995): Hemostatic factors and the risk of myocardial infarction or sudden death in patients with angina pectoris. N Engl J Med 332:635-641.

Thuraisingham RC and Raine AEG (1999): Maintenance of normal agonist-induced endothelium-dependent relaxation in uraemic and hypertensive resistance vessels. Nephrol Dial Transplant 14:70-75.

Timmermans PB, Wong PC, Chiu AT, Herblin WF, Benfield P, Carini DJ, Lee RJ, Wexler RR, Saye JA and Smith RD (1993): Angiotensin II receptors and angiotensin II receptor antagonists. Pharmacol Rev 45:205-251.

Tolvanen JP, Wu X, Kahonen M, Sallinen K, Makynen H, Pekki A and Porsti I (1996): Effect of celiprolol therapy on arterial dilatation in experimental hypertension. Br J Pharmacol 119:1137-1144.

Tolvanen JP, Mäkynen H, Wu X, Hutri-Kähönen N, Ruskoaho H, Karjala K and Pörsti I (1998): Effects of calcium and potassium supplements on arterial tone in vitro in spontaneously hypertensive rats. Br J Pharmacol 124:119-128.

Touz RM (2003): The role of angiotensin II in regulating vascular structural and functional changes in hypertension. Curr Hypertens Rep 5:155-164.

Touyz RM, Deng L-Y, He G and Schiffrin EL (1999): Angiotensin II stimulates DNA and protein synthesis in vascular smooth muscle cells from human peripheral resistance arteries: role of extracellular signal-regulated kinases. J Hypertens 7:907-917.

Touyz RM and Schiffrin EL (2000): Signal transduction mechanisms mediating the physiological and pathophysiological actions of Ang II in vascular smooth muscle cells. Pharmacol Rev 52:639-672.

Touyz RM, Xiao-Hua W, He G, Salomon S and Schiffrin EL (2002): Increased Angiotensin II-mediated Src signaling via epidermal growth factor receptor transactivation is associated with decreased C-terminal Src kinase activity in vascular smooth muscle cells from spontaneously hypertensive rats. Hypertension 39:479-485.

Tribulova N, Okruhlicova L, Bernatova I and Pechanova O (2000): Chronic disturbances in NO production results in histochemical and subcellular alterations of the rat heart. Physiol Res 49:77-88.

Tsuru H, Tanimitsu N and Hirai T (2002): Role of perivascular sympathetic nerves and regional differences in the features of sympathetic innervation of the vascular system. Jpn J Pharmacol 88:9-13.

Tuomilehto J, Jousilahti P, Rastenyte D, Moltchanov V, Tanskanen A, Pietinen P and Nissinen A (2001): Urinary sodium excretion and cardiovascular mortality in Finland: a prospective study. Lancet 357:848-851.

Urata K, Kinoshita A, Misono K, Bumpus FM and Husain A (1990): Identification of a highly specific chymase as the major angiotensin-forming enzyme in the human heart. J Biol Chem 265:22348-22382.

Ushio-Fukai M, Abe S, Kobayashi S, Nishimura J and Kanaide H (1993): Effects of isoprenaline on cytosolic calcium concentrations and on tension in the porcine coronary artery. J Physiol 462:679-696.

Usui M, Ichiki T, Katoh M, Egashira K and Takeshita A (1998): Regulation of angiotensin II receptor expression by nitric oxide in rat adrenal gland. Hypertension 32:527-533.

Usui M, Egashira K, Kitamoto S, Koyanagi M, Katoh M, Kataoka C, Shimokawa H and Takeshita A (1999): Pathogenic role of oxidative stress in vascular angiotensin-converting enzyme activation in long-term blockade of nitric oxide synthesis in rats. Hypertension 34:546-551.

Vallance P, Leone A, Calver A, Collier J and Moncada S (1992): Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. Lancet 339:572-575.

Van Breemen C and Saida K (1989): Cellular mechanism regulating $[Ca^{2+}]_i$ smooth muscle. Annu Rev Physiol 51:315-329.

Van den Bemd GJ, Pols HA and van Leeuwen JP (2000): Anti-tumor effects of 1,25-dihydroxyvitamin D_3 and vitamin D analogs. Curr Pharm Des 6:717-732.

Van de Voorde J, Vanheel B and Leusen I (1992): Endothelium-dependent relaxation and hyperpolarization in aorta from control and renal hypertensive rats. Circ Res 70:1-8.

Van Guldener C, Lambert J, Janssen MJ, Donker AJ and Stehouwer CD (1997): Endothelium-dependent vasodilation and distensibility of large arteries in chronic haemodialysis patients. Nephrol Dial Transplant 12 Suppl 2:14-18.

Van Leer EM, Seidell JC and Kromhout D (1995): Dietary calcium, potassium, magnesium and blood pressure in the Netherlands. Int J Epidemiol 24:1117-1123.

Van Vliet BN, Hall JE, Lohmeier TE and Mizelle HL (1996): Renal circulation, in Bennet T, Gardiner S (eds): Nervous control of blood vessels. London, Harwood Academic Publishers, 371-433.

Van Zwieten PA (1997): Endothelial dysfunction in hypertension. A critical evaluation. Blood Press Suppl 2:67-70.

Vane JR and Botting RM (1993): Formation by the endothelium of prostacyclin, nitric oxide and endothelin. J Lipid Mediat 6:395-404.

Vanhoutte PM (1993): Is endothelin involved in the pathogenesis of hypertension. Hypertension 21:747-751.

Vanhoutte PM, Boulanger CM, Illiano SC, Nagao T, Vidal M and Mombouli J-V (1993): Endotheliumdependent effects of converting-enzyme inhibitors. J Cardiovasc Pharmacol 22:10-16. Vanhoutte PM and Mombouli J-V (1996): Vascular endothelium: Vasoactive mediators. Prog Cardiovasc Dis 39:229-238.

Vaskonen T (2003): Dietary minerals and modification of cardiovascular risk factors. J Nutr Biochem 14:492-506.

Vaziri ND, Ni Z, Oveisi F, Liang K and Pandian R (2002): Enhanced nitric oxide inactivation and protein nitration by reactive oxygen species in renal insufficiency. Hypertension 39:135-141.

Vaziri ND, Oveisi F and Ding Y (1998): Role of increased oxygen free radical activity in the pathogenesis of uremic hypertension. Kidney Int 53:1748-1754.

Verbeke M, Van de Voorde J, de Ridder L and Lameire N (1994): Functional analysis of vascular dysfunction in cyclosporin treated rats. Cardiovasc Res 28:1152-1156.

Vila E, Tabernero A and Ivorra MD (1993): Inositol phosphate formation and contractile response linked to alpha 1-adrenoceptor in tail artery and aorta from spontaneously hypertensive and Wistar-Kyoto rats. J Cardiovasc Pharmacol 22:191-197.

Vita JA and Keaney JF Jr (2002): Endothelial function: a barometer for cardiovascular risk? Circulation 106:640-642.

Vita JA and Loscalzo J (2002): Shouldering the risk factor burden: infection, atherosclerosis, and the vascular endothelium. Circulation 106:164-166.

Wagner OF, Christ G, Wojta J, Vierhapper H, Parzer S, Nowotny PJ, Schneider B, Waldhäusl W and Binder BR (1992): Polar secretion of endothelin-1 by cultured endothelial cells. J Biol Chem 267:16066-16068.

Walsh MP (1994): Regulation of vascular smooth muscle tone. Can J Physiol Pharmacol 72:919-936.

Wang R and Wu L (1997): The chemical modification of K_{Ca} channels by carbon monoxide in vascular smooth muscle cells. J Biol Chem 272:8222-8226.

Wang XL, Mahaney MC, Sim AS, Wang J, Wang J, Blangero J, Almasy L, Badenhop RB and Wilcken DE (1997): Genetic contribution of the endothelial constitutive nitric oxide synthase gene to plasma nitric oxide levels. Arterioscler Thromb Vasc Biol 17:3147-3153.

Wang Y and Bukoski RD (1998): Differential distribution of the pervascular Ca²⁺ receptor in rat arteries. Br J Pharmacol 125:1397-1404.

Warrens AN, Cassidy MJ, Takahashi K, Ghatei MA and Bloom SR (1990): Endothelin in renal failure. Nephrol Dial Transplant 5:418-422.

Weber AA, Zucker TP, Hasse A, Bonisch D, Wittpoth M and Schror K (1998): Antimitogenic effects of vasodilatory prostaglandins in coronary artery smooth muscle cells. Basic Res Cardiol 93:54-57.

Weinberger MH, Miller JZ, Luft FC, Grim CE and Fineberg NS (1986): Definitions and characteristics of sodium sensitivity and blood pressure resistance. Hypertension 6:II127-II134.

Wellman GC, Cartin L, Eckman DM, Stevenson AS, Saundry CM, Lederer WJ and Nelson MT (2001): Membrane depolarization, elevated Ca(2+) entry, and gene expression in cerebral arteries of hypertensive rats. Am J Physiol Heart Circ Physiol 281:H2559-H2567.

Widlansky ME, Gokce N, Keaney JF and Vita JA (2003): The clinical implications of endothelial dysfunction. J Am Coll Cardiol 42:1149-1160.

Winder SJ, Allen BG, Clement-Chomienne O and Walsh MP (1998): Regulation of smooth muscle actin-myosin interaction and force by calponin. Acta Physiol Scand 164:415-426.

Wright JW, Krebs LT, Stobb JW and Harding JW (1995): The angiotensin IV system: Functional implications. Front Neuroendocrinol 16:23-52.

Wyss JM and Carlson SH (1999): The role of the central nervous system in hypertension. Curr Hypertens Rep 1:246-253.

Ying WZ and Sanders PW (2002): Increased dietary salt activates rat aortic endothelium. Hypertension 39:239-244.

Young EW, Bukoski RD and McCarron DA (1988): Calcium metabolism in experimental hypertension. Proc Soc Exp Biol Med 187:123-141.

Zatz R and Baylis C (1998): Chronic nitric oxide inhibition model six years on. Hypertension 32:958-964.

Zemel MB (2001): Calcium modulation of hypertension and obesity: mechanisms and implications. J Am Coll Nutr 20:428S-435S.

Zhao H, Shimokawa H, Uragami-Harasawa L, Igarashi H and Takeshita A (1999): Long-term vascular effects of Nomega-nitro-L-arginine methyl ester are not soley mediated by inhibition of endothelial nitric oxide synthesis in the rat mesenteric artery. J Cardiovasc Pharmacol 33:554-566.

Zhu Z, Neusser M, Tepel M, Spieker C, Golinski P and Zidek W (1994): Effect of Na,K-ATPase inhibition on cytosolic free calcium ions in vascular smooth muscle cells of spontaneously hypertensive and normotensive rats. J Hypertens 12:1007-1012.

Zicha J and Kunes J (1999): Ontogenetic aspects of hypertension development: analysis in the rat. Physiol Rev 79:1227-1282.

Zygmunt PM and Hoggestatt ED (1996): Role of potassium channels in endothelium-dependent relaxation resistant to nitroarginine in the rat hepatic artery. Br J Pharmacol 117:1600-1606.

ORIGINAL COMMUNICATIONS