



## Phosphorylation of NFATC1 at PIM1 target sites is essential for its ability to promote prostate cancer cell migration and invasion

### Citation

Eerola, S. K., Santio, N. M., Rinne, S., Kouvonen, P., Corthals, G. L., Scaravilli, M., ... Koskinen, P. J. (2019). Phosphorylation of NFATC1 at PIM1 target sites is essential for its ability to promote prostate cancer cell migration and invasion. *CELL COMMUNICATION AND SIGNALING*, 17(1), [148].  
<https://doi.org/10.1186/s12964-019-0463-y>

### Year

2019

### Version

Publisher's PDF (version of record)

### Link to publication

[TUTCRIS Portal \(http://www.tut.fi/tutcris\)](http://www.tut.fi/tutcris)

### Published in

CELL COMMUNICATION AND SIGNALING

### DOI

[10.1186/s12964-019-0463-y](https://doi.org/10.1186/s12964-019-0463-y)

### License

CC BY

### Take down policy


If you believe that this document breaches copyright, please contact [cris.tau@tuni.fi](mailto:cris.tau@tuni.fi), and we will remove access to the work immediately and investigate your claim.

RESEARCH

Open Access



# Phosphorylation of NFATC1 at PIM1 target sites is essential for its ability to promote prostate cancer cell migration and invasion

Sini K. Eerola<sup>1,2</sup>, Niina M. Santio<sup>1</sup>, Sanni Rinne<sup>1</sup>, Petri Kouvonen<sup>3</sup>, Garry L. Corthals<sup>3</sup>, Mauro Scaravilli<sup>2,4</sup>, Giovanni Scala<sup>2,5</sup>, Angela Serra<sup>2</sup>, Dario Greco<sup>2,5</sup>, Pekka Ruusuvaori<sup>2,6</sup>, Leena Latonen<sup>2,4</sup>, Eeva-Marja Rainio<sup>1</sup>, Tapio Visakorpi<sup>2,7</sup> and Päivi J. Koskinen<sup>1\*</sup> 

## Abstract

**Background:** Progression of prostate cancer from benign local tumors to metastatic carcinomas is a multistep process. Here we have investigated the signaling pathways that support migration and invasion of prostate cancer cells, focusing on the role of the NFATC1 transcription factor and its post-translational modifications. We have previously identified NFATC1 as a substrate for the PIM1 kinase and shown that PIM1-dependent phosphorylation increases NFATC1 activity without affecting its subcellular localization. Both PIM kinases and NFATC1 have been reported to promote cancer cell migration, invasion and angiogenesis, but it has remained unclear whether the effects of NFATC1 are phosphorylation-dependent and which downstream targets are involved.

**Methods:** We used mass spectrometry to identify PIM1 phosphorylation target sites in NFATC1, and analysed their functional roles in three prostate cancer cell lines by comparing phosphodeficient mutants to wild-type NFATC1. We used luciferase assays to determine effects of phosphorylation on NFAT-dependent transcriptional activity, and migration and invasion assays to evaluate effects on cell motility. We also performed a microarray analysis to identify novel PIM1/NFATC1 targets, and validated one of them with both cellular expression analyses and in silico in clinical prostate cancer data sets.

**Results:** Here we have identified ten PIM1 target sites in NFATC1 and found that prevention of their phosphorylation significantly decreases the transcriptional activity as well as the pro-migratory and pro-invasive effects of NFATC1 in prostate cancer cells. We observed that also PIM2 and PIM3 can phosphorylate NFATC1, and identified several novel putative PIM1/NFATC1 target genes. These include the ITGA5 integrin, which is differentially expressed in the presence of wild-type versus phosphorylation-deficient NFATC1, and which is coexpressed with PIM1 and NFATC1 in clinical prostate cancer specimens.

**Conclusions:** Based on our data, phosphorylation of PIM1 target sites stimulates NFATC1 activity and enhances its ability to promote prostate cancer cell migration and invasion. Therefore, inhibition of the interplay between PIM kinases and NFATC1 may have therapeutic implications for patients with metastatic forms of cancer.

**Keywords:** Prostate cancer, Metastatic carcinoma, NFATC1, PIM kinases, Cell motility

\* Correspondence: [paivi.koskinen@utu.fi](mailto:paivi.koskinen@utu.fi)

<sup>1</sup>Department of Biology, University of Turku, Vesilinnantie 5, FI-20500 Turku, Finland

Full list of author information is available at the end of the article



## Background

Prostate cancer is globally one of the most prevalent cancers in men. Locally restricted prostate cancer is usually not fatal, but there is a clear need for effective therapies to prevent or stop progression of local tumors to a metastatic state spreading to bones and other vital organs. Formation of metastases is a multistep process, which includes detachment of cancer cells from the primary tumor, migration, adhesion and invasion of cancer cells into blood or lymph vessels, and infiltration of the cells to secondary sites. Thus, improved understanding of the proteins and signaling pathways that regulate the metastatic growth of cancer cells is essential when developing therapies to treat prostate cancer patients.

NFAT (Nuclear Factor of Activated T cells) transcription factors are ubiquitously expressed in human tissues, where they control cellular processes, such as immune responses [1]. However, one of the family members, NFATC1, has also been shown to act as an oncogene that promotes cancer cell proliferation and transformation [2]. Accordingly, elevated levels as well as increased transcriptional activity of NFATC1 have been detected in both solid cancers and hematological malignancies. NFATC1 has been shown to support cell migration or invasion in multiple types of cancer, such as ovarian, breast and prostate cancer as well as glioblastoma [3–7]. Furthermore, it has been reported to support metastatic behavior of prostate or breast cancer cells via increased osteoclastogenesis [8, 9].

Both the subcellular localization and transcriptional activity of NFAT proteins are post-translationally regulated. Most previously identified phosphorylation sites in NFATC1 have been located to the serine-rich regions (SRRs) and SPXX motifs within the NFAT homology region [10, 11]. Phosphorylation of these sites by kinases such as PKA and GSK3 results in nuclear exit and inactivation of NFATC1. By contrast, dephosphorylation of these sites by the calcium-dependent phosphatase calcineurin leads to nuclear translocation and transcriptional activation.

We have previously shown that the oncogenic PIM1 kinase directly interacts with NFATC1 and phosphorylates it *in vitro* [12]. However, in contrast to other kinases, PIM1 does not affect the subcellular localization of NFATC1, but stimulates its transcriptional activity in both immune and neuronal cells [12, 13]. PIM1 belongs to a family of three serine/threonine-specific kinases, which have partially overlapping expression patterns, but share several functions to support cell proliferation and survival [14–16]. Increased expression of PIM family members has been detected both in hematological malignancies and in solid tumors. In prostate cancer, overexpression of either PIM1 or PIM3 positively correlates with tumor size, aggressiveness and/or poor patient

survival [17–21]. Furthermore, PIM kinases have been linked to regulation of prostate cancer cell motility in several cell-based and animal models, where they have supported cell migration, invasion, tumor angiogenesis and the formation of metastases [4, 16, 22]. As also NFATC1 promotes motility of prostate cancer cells and as PIM-selective inhibitors can block this [4], we now wanted to investigate whether or not PIM-dependent phosphorylation of NFATC1 is important for migration and invasion of prostate cancer cells. Therefore, we identified and mutated the PIM targets sites from NFATC1 and analysed the impact of these mutations in three prostate cancer cell lines, the hormone-insensitive PC-3 and DU-145 cells and the hormone-sensitive LNCaP cells. We also performed a microarray analysis to identify putative phosphorylation-dependent target genes for NFATC1.

## Methods

### Cell culture

The cell culture conditions for prostate epithelial adenocarcinoma cell line PC-3 and the stable cell lines overexpressing human PIM1 have been previously described [22]. DU145 and LNCaP prostate cancer cell lines were obtained from American Type Culture Collection (Manassas, VA) and cultured under recommended conditions. For transient transfections, Eugene 6 or HD reagents (Promega, Fitchburg, WI, USA) were used in 1:2 or 1:3 ratio to DNA according to manufacturer's instructions. All the cell lines were frequently tested for mycoplasma contamination. Viability of cells was analysed by the MTT assay [4] or the AlamarBlue® cell viability assay (Thermo Fisher Scientific, Waltham, MA, USA).

### DNA constructs and cloning

The pcDNA3.1/V5-HisC, pGEX-6P-1 and pTag-RFP vectors expressing wild-type (WT) or kinase-deficient (KD) human PIM1, 2 or 3 or mouse PIM3 have been previously described [23]. The NFAT-luciferase reporter plasmids as well as wild-type (WT), N-terminally truncated (amino acids 1–418), dominant negative (DN, amino acids 410–680) and constitutively active SRR mutant (mSRR) human NFATC1 expression vectors based on pGEX-3X or pBJ5-Flag were kindly provided by the group of G.R. Crabtree (Stanford University, CA, USA) [10, 24]. Truncated NFATC1 was digested from pGEX-3X with PflMI and ligated to pEYFP-C2 (Clontech Laboratories, Mountain View, CA, USA). Full-length NFATC1 was multiplied by PCR from pBJ5-NFATC1-Flag by using a forward primer (5' GCG GTA CCG CCA CCA TGG ACT ACA AGG CA 3') and a reverse primer (5' CCC GGA TCC CTG CGT CTT TAG 3'), digested with KpnI and BamHI, and ligated into pFlag-CMV-2 (Sigma-Aldrich), from where it was further

transferred to pEGFP-C3 (Clontech) by BglI and BamHI digestion, followed by ligation.

#### NFATC1 mutagenesis

The QuikChange™ site-directed mutagenesis kit (Stratagene, Agilent Technologies, Santa Clara, CA, USA) was used to prepare phosphodeficient mutants of NFATC1. Mutations to replace serines or threonines with alanine residues were introduced into ten PIM1 target sites with the help of five different primer pairs (Additional file 1: Table S1), resulting in production of double mutant (DM, two primer pairs, 1–2), triple mutant (TM, three primer pairs, 3–5) or multi mutant (MM, all primer pairs) NFATC1.

#### In vitro kinase assays

GST fusion proteins were produced in the *E. coli* BL21 strain as previously described [25] with minor modifications. Protein production was induced with 0,5 mM IPTG and protease activity was inhibited by Aprotinin (1:200; Sigma-Aldrich) during cell lysis. Proteins were either eluted as fusion proteins or cleaved by the Pre-Scission protease according to manufacturer's protocol (GE Healthcare Life Sciences, Little Chalfont, UK). For in vitro kinase assays, cleaved PIM kinase (0.5 µg) and GST-tagged NFATC1 (amino acids 1–418) fusion protein (1 µg) were mixed prior to addition of the 2x kinase buffer (20 mM Pipes, pH 7.0, 5 mM MnCl<sub>2</sub>, 0.25 mM β-glycerophosphate, 0.4 mM spermine, 10 µM ATP) with 0.5 MBq of [<sup>32</sup>P] adenosine triphosphate. To inhibit PIM kinase activity, samples were pre-treated for 15 min with 10 µM DHPCC-9, a pan-PIM inhibitor, which was kindly provided by P. Moreau (University of Clermont Auvergne, France) and dissolved in 0,1% DMSO. This ATP-competitive pyrrolocarbazole compound selectively inhibits catalytic activities of all PIM family members in vitro [26], in cell-based assays [4] and in mice xenografted with PIM-expressing prostate cancer cells [22]. After 15 to 30 min kinase reactions at 30 °C, samples were heated in 2x Laemmli sample buffer (LSB) for 5 min at 95 °C. Phosphorylated proteins were resolved in SDS-PAGE, stained by Page Blue solution (Thermo Fisher Scientific) and detected by autoradiography.

#### Identification of NFATC1 in vivo phosphorylation sites by mass spectrometry

PC-3 cells were transiently transfected with the pEYFP-NFATC1 expression vector. After 48 h, cells were stimulated with TPA and IM for 1 h prior to cell lysis in RIPA buffer supplemented with complete mini EDTA-free protease inhibitors (Roche, Basel, Switzerland). Protein concentrations were determined by the DC Lowry method (Bio-Rad Laboratories, Inc., Hercules, CA, USA). 1 mg aliquots of proteins were mixed with Chromotek-

GFP-Trap® Magnetic beads (Allele Biotechnology, San Diego, CA, USA), after which GFP-tagged proteins were immunoprecipitated according to manufacturer's protocol, heated in 2x LSB, resolved in 10% Bis-Tris gel (Bio-Rad) and stained with colloidal coomassie blue solution (Thermo Fisher Scientific). NFATC1 protein isolation, trypsin digestion and titanium dioxide enrichment without salt extraction were performed as previously described [27, 28]. Thereafter, samples were analysed by LTQ Orbitrap Velos mass spectrometer (Thermo Fisher Scientific), using the HCD Top 10 method with 10 min gradient and mass value of 300 to 2000.

#### Luciferase assays

To measure NFAT-dependent transcriptional activity, PC-3 cells were transiently transfected with the pGL3-IL-2-luciferase reporter and either pBJ5-NFATC1-Flag or an empty control vector. To stimulate NFATC1 activity and nuclear translocation, cells were treated for 7 h with 15 ng/ml of 12–0-tetradecanoyl-phorbol-13-asetate (TPA; Sigma-Aldrich, St. Louis, MO, USA) in DMSO and 1 µM ionomycin (IM; Merck KGaA, Darmstadt, Germany) in EtOH. To inhibit PIM kinase activity, cells were treated for 24 h with 10 µM DHPCC-9 in 0,1% DMSO. As controls for all chemical compounds, their solvents were used. 24 or 48 h after transfections, cells were collected, lysed in 1% NP-40 buffer by repeated freezing and thawing, and analysed for luciferase activity using the Luminoscan Ascent luminometer (Thermo Fisher Scientific).

To compare activities of wild-type (WT) and multi mutant (MM) NFATC1 in PC-3, DU-145 and LNCaP cell lines, cells were transiently transfected with the pGL3-NFAT-luciferase reporter and either WT or MM pCMV-NFATC1-Flag or an empty control vector. Renilla luciferase (pRLTk; Promega) was co-transfected as an internal transfection efficiency control. Part of the cells were treated with TPA and IM and/or DHPCC-9 as described above. To inhibit calcineurin activity and thereby also nuclear translocation of NFATC1, cells were treated for 24 h with 1 µM cyclosporine A (CsA; Merck) in EtOH. Luciferase assays with four parallel samples were performed on 96-well plates using the Dual-Glo® Luciferase Assay System (Promega) according to manufacturer's protocol. Luciferase activities were measured with the EnVision 2104 Multilabel Reader (Perkin Elmer, Waltham, MA, USA). The results were presented as relative luciferase activity (RLU) corresponding to the firefly luciferase light emission values normalized against renilla luciferase light emission values.

#### Localization assays

To determine the subcellular localizations of wild-type and mutant NFATC1 proteins, PC-3 cells plated on

coverslips were transiently transfected with Flag-tagged expression vectors. After 48 h, cells were fixed, permeabilized and stained with anti-Flag antibody (Sigma-Aldrich) and Alexa Fluor™ 488 labelled anti-mouse secondary antibody (Thermo Fisher Scientific). Samples were imaged and analysed with the Zeiss ApoTome.2 fluorescence microscope and Zen lite 2012 software. Approximately 15 images were taken from each sample.

#### Fluorescence-lifetime imaging method (FLIM)

To visualize interactions between RFP-tagged PIM1 and GFP-tagged NFATC1, PC-3 cells plated on coverslips were transiently transfected with the corresponding expression vectors and/or their empty controls. Part of the samples were treated overnight with DMSO or 10  $\mu$ M DHPCC-9. 48 h after transfection, cell samples were fixed with 4% PFA and mounted with Mowiol. First, physical interactions between tagged proteins were measured by analysing GFP lifetime with Lambert Instruments Fluorescence Lifetime Attachment (LIFA) and LI-FLIM software as previously described [23]. Then colocalization of proteins was imaged by Zeiss LSM 780 confocal microscope and by sequential scanning with ZEN lite 2012 software. Excitation wavelengths were 488 nm (GFP) and 561 nm (RFP), and emission wavelengths 500–535 nm (GFP) and 599–651 nm (RFP). Image analyses were performed with the ImageJ® software (Wayne Rasband, NIH, USA).

#### Wound healing assays

PC-3 or DU-145 cells were transiently transfected with wild-type or mutant NFATC1 expression vectors. 24 h later, samples were treated with either DMSO or 10  $\mu$ M DHPCC-9. To confirm that changes in cell migration were not due to changes in cell proliferation, 15  $\mu$ g/ml of the anti-proliferative agent mitomycin C (Sigma-Aldrich) was used. Scratching of the wounds, microscopy and image analyses of PC-3 cells were performed as previously described [4]. Imaging of DU-145 cells was performed with CM Technologies Cell-IQ (D.I. Biotech, Korea) by using 4x objective and image analysis with the Cell-IQ software 4.3 and scratch wound measurement tool.

#### Boyden chamber invasion assays

One day after transfection, invasiveness of PC-3 cells was analysed using cell culture invasion inserts of 8  $\mu$ m pore size (Corning BioCoat™ Matrigel® Invasion Chamber, Bedford, MA, USA) according to manufacturer's instructions. For this purpose, cells were suspended in DMEM supplemented with 1% BSA (20,000 cells/each chamber) and either DMSO or 10  $\mu$ M DHPCC-9. Conditioned medium from confluent MG-63 human osteosarcoma cells was used as a chemoattractant [29].

Cells were incubated for 48 h, after which insert membranes were fixed for 2 min in methanol and stained for 10 min with 0.2% crystal violet in methanol. Then they were cut out from the inserts and mounted with immersion oil. Invaded cells on the membranes were scanned by the Olympus BX51 scanner with Surveyor software and analysed by automated image analysis. Results were verified by manual counting with the ImageJ® software from 5 random fields of each membrane.

#### Gelatinase activity assay

Gelatinase activity assay was performed with InnoZyme™ gelatinase (MMP-2/MMP-9) fluorogenic activity assay kit (Merck) according to manufacturer's instructions. Medium samples for the assay were collected from the upper chambers of invasion inserts after the invasion assays described above. Samples were incubated at + 37 °C for 3 h protected from light. Fluorescence was then measured with the Envision plate reader (Perkin Elmer) with an excitation wavelength of 320 nm and an emission wavelength of 405 nm.

#### Western blotting

Cells were lysed in 2x LSB and heated at 95 °C for 5 min. Proteins were separated by SDS-PAGE, immobilized onto PVDF-membrane (EDM Millipore, Merck) and incubated overnight with anti-PIM-1 (1:500, 12H8; Santa Cruz Biotechnology, Santa Cruz, CA, USA), anti-PIM-2 (1:1000, D1D2; Cell Signaling Technology, Danvers, MA, USA), anti-PIM-3 (1:1000, D17C9; Cell Signaling Technology), anti-NFATC1 (1:500, Santa Cruz Biotechnology), anti-V5 (1:500, Invitrogen, Carlsbad, CA, USA), anti-Flag (1:500, F1804; Sigma-Aldrich), anti-ACTB (anti- $\beta$ -actin; 1:1000, 13E5, #4970S, Cell signaling Technology), anti-GAPDH (1:50000, Sigma-Aldrich), anti- $\beta$  Tubulin (1:40000, Sigma-Aldrich) or anti-Fibrillarlin (1:1000, Cell Signaling Technology) antibodies. After incubations with secondary antibodies, chemiluminescence reactions were generated using either Amersham™ ECL Plus or ECL Prime reagents (GE Healthcare).

#### Microarray analyses

For microarray analyses, PC-3 cells with or without stable PIM1 overexpression were transiently transfected with wild-type (WT) or multi mutant (MM) NFATC1 expression vectors. At the following day, total RNAs were extracted using the TRIzol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. The samples were then labelled and hybridized using the Agilent whole genome oligo microarray platform on Human Gene Expression v2 4x44K Microarray slides (G4845A; Agilent Technologies, Palo Alto, CA, USA). The slides were scanned on the Agilent C-Scanner and the raw expression values were extracted

using the Agilent Feature Extraction software v. 11.0.1.1. Raw mRNA expression values were imported using `limma read.maimages` function. Low quality probes were filtered using the distribution of negative control probes as a reference. In particular, only probes whose raw expression values were higher than the 90th percentile of negative control probes were retained for successive analysis. Expression values were log<sub>2</sub> transformed, quantile normalized between samples and median aggregated at the gene symbol level using Agilent annotation. A limma-based approach [30] was then applied to estimate the difference in average expression in each comparison. A fold-change cutoff ( $\geq 0.1$ ) and *p*-value of ( $< 0.05$ ) were used to determine differential gene expression.

#### Canonical pathway analysis

IPA (Ingenuity Pathway Analysis, Ingenuity Systems) was used for functional enrichment and detection of pathways with significant alterations based on microarray gene expressions. In canonical pathway analysis  $-\log(p\text{-values})$  over threshold 2.5 were considered significant.

#### Real-time quantitative polymerase chain reaction (qRT-PCR)

*PIM*, *NFATC1* and *ITGA5* expression levels were determined from total RNAs isolated from PC-3 cells as described above. Quantitative real-time PCR was performed using random hexamere primers, Maxima reverse transcriptase (Thermo Scientific), Maxima SYBR Green qPCR Master Mix (Thermo Fischer Scientific) and the CFX96™ Real-Time PCR Detection System (Bio-Rad Laboratories, Inc.). Each sample was run in triplicate, and expression values were normalized against the TATA-binding protein (TBP). Sequences of all primers (Sigma-Aldrich) for qRT-PCR are described in the Additional file 1: Table S2.

#### Gene correlation analyses

Three distinct clinical data sets were used to assess correlations between two different genes in clinical prostate cancer patient samples: The Cancer Genome Atlas (TCGA) - Prostate adenocarcinoma RNA-Sequencing data [31], Integrative Genomic Profiling of Human Prostate Cancer microarray data [32] and Tampere PC sequencing data [33].

#### Statistical analyses

The statistical significance of data from luciferase, wound healing, FLIM and cell viability assays was determined using the two-sided *t*-test. Cell invasion and gelatinase activity data were analysed by using the unpaired two-sided *t*-test or Wilcoxon matched pairs test. In RT-qPCR data validation, *P*-values were determined by the Mann-Whitney U-test. In gene correlation analyses, Pearson

correlation coefficient and *P*-values were determined according to Gaussian populations. In all analyses, a *P*-value  $< 0.05$  was considered statistically significant (\*),  $P < 0.01$  (\*\*) and  $P < 0.001$  (\*\*\*). Error bars represent standard deviation (SD) values in each graph. Statistical analyses were performed using the GraphPad Prism version 5.02 (GraphPad Software, La Jolla, CA, USA).

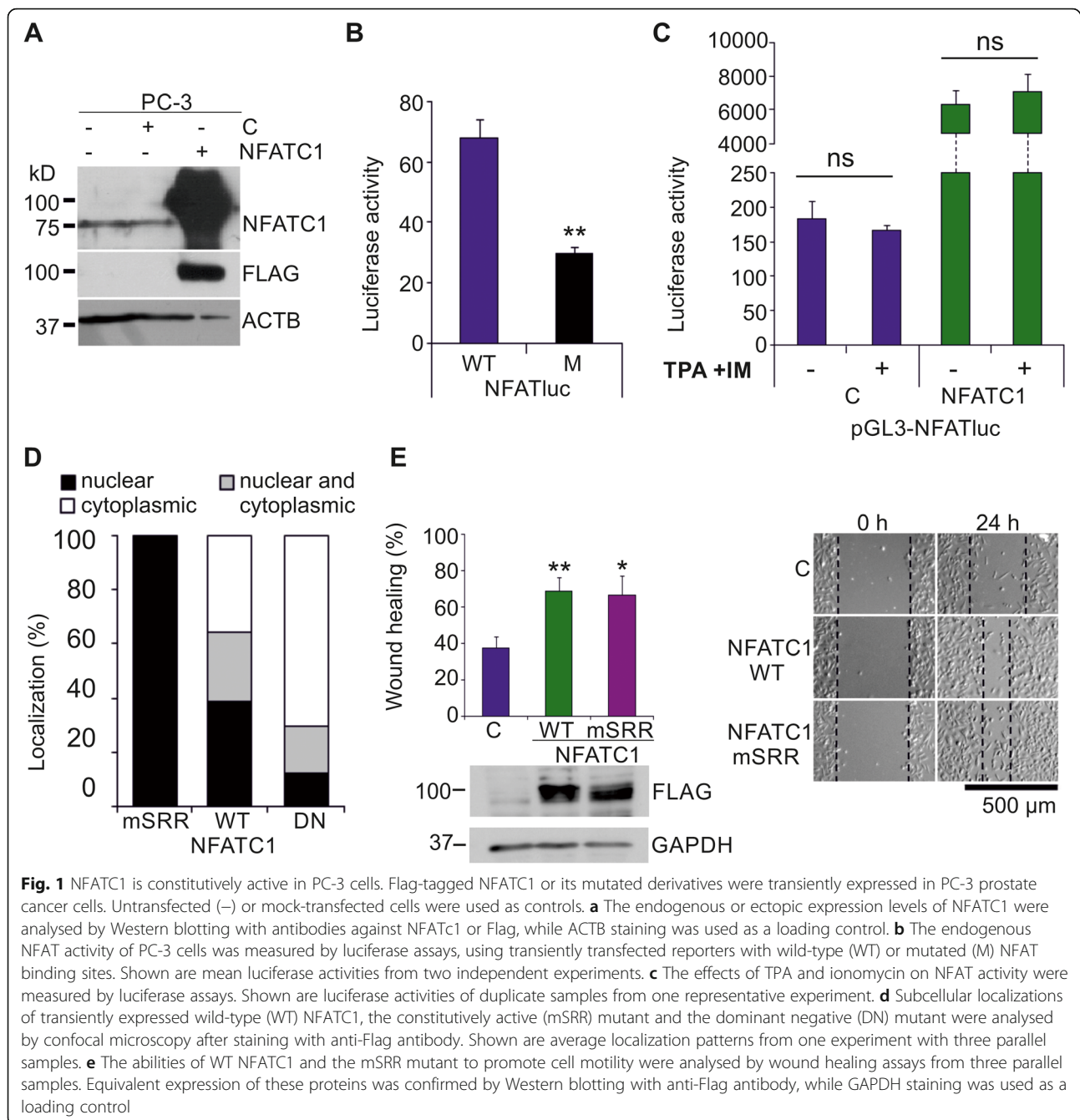
## Results

### NFATC1 is endogenously expressed and constitutively active in PC-3 cells

As we had previously shown both PIM kinases and NFATC1 to be essential for the motility of PC-3 prostate cancer cells [4], we decided to use these cells in order to investigate in more detail the functional interactions between PIM and NFATC1 proteins. When we analysed the basal expression and transcriptional activity of NFATC1 in PC-3 cells, Western blotting with NFATC1 antibodies detected an endogenously expressed protein with the expected size of approximately 75 kDa (Fig. 1a). NFAT-dependent luciferase assays in turn revealed endogenous NFAT activity, which was dependent on the presence of NFAT binding sites (Fig. 1b), and which was enhanced by ectopic overexpression of NFATC1, but not by stimulation of cells with TPA and the calcium ionophore ionomycin (Fig. 1c). This was surprising, since usually the nuclear translocation and activation of NFATC1 is tightly regulated in a calcium- and calcineurin-dependent fashion [1, 2]. To determine the subcellular localization of NFATC1 in PC-3 cells, we transiently expressed there wild-type (WT) or mutant NFATC1 proteins (24; Table 1). While the dominant negative (DN) mutant was mostly retained in the cytoplasm and the constitutively active (mSRR) mutant in the nucleus, the WT protein could be detected in both compartments (Fig. 1d, Additional file 2: Figure S1A), suggesting that it can shuttle between the compartments of PC-3 cells. When we carried out wound healing assays to compare the effects of WT and mSRR NFATC1 on cell migration, we noticed that both of them enhanced cell motility as compared to control cells (Fig. 1e), while no major changes were observed in cell viability (Additional file 2: Figure S1B).

### PIM kinases phosphorylate NFATC1 in several serine and threonine residues

As we had previously shown that the PIM1 kinase phosphorylates NFATC1 and enhances its transcriptional activity [12], we now wanted to identify the as yet unknown PIM1 target sites in NFATC1 and to investigate their physiological roles in more detail. For this purpose, we carried out in vitro kinase assays with GST-tagged PIM-1 and NFATC1 (amino acids 1–418) produced in bacteria, and cell-based assays with YFP-tagged

**Table 1** Different NFATC1 forms and mutants used in the experiments

NFATC1 proteins	Mutated sites	Length
Wild type (WT)	none	full-length
Dominant negative (DN)	none	410–680 aa
Constitutively active (mSRR)	all 11 serines mutated to alanines in the SRR (172–194)	1–418 aa
Double mutant (DM)	S245, S269	full-length
Triple mutant (TM)	S151, S153, T154, S256, S257, S335, T338, T339	full-length
Multi mutant (MM)	S151, S153, T154, S245, S256, S257, S269, S335, T338, T339	full-length

The amino acid substitutions (from serine or threonine to alanine) and other mutations in NFATC1 and the length of each mutant protein used in this study

NFATC1 protein overexpressed in PC-3 cells. When phosphorylated NFATC1-derived peptides were subjected to mass spectrometry analyses, several novel phosphorylation target sites were discovered both from the in vitro samples and from the PC-3 cell-derived samples (Fig. 2a, Additional file 1: Table S3) in addition to those in vivo sites that we had already previously identified from COS-7 cells [27]. However, since more endogenously phosphorylated cellular sites were discovered than in vitro target sites for PIM1, it was evident that many of the in vivo sites were targeted by other kinases.

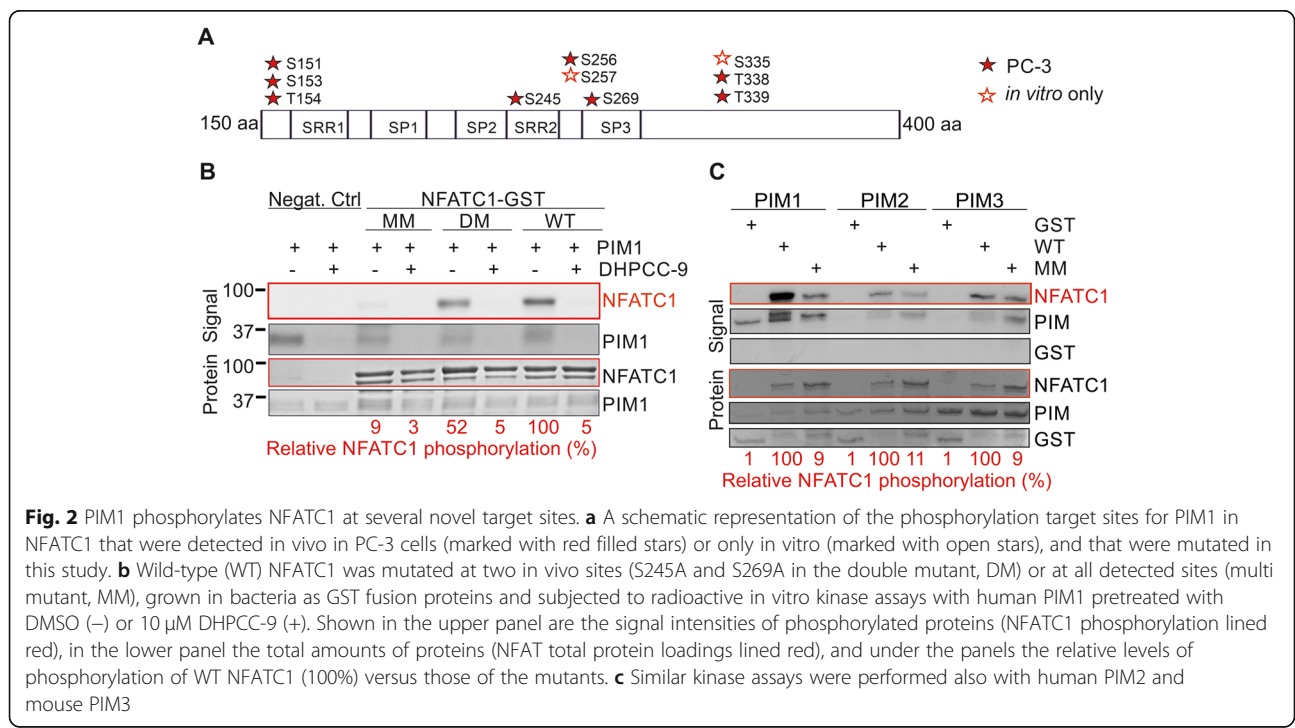
To be able to evaluate the functional impact of phosphorylation at putative PIM1 target sites, we mutated multiple serine or threonine residues in NFATC1 into alanines to create phosphodeficient mutants (Fig. 2a, Table 1, Additional file 1 Table S3). The mutated sites were primarily chosen based on the presence of PIM1 consensus target sequences [34] with basic residues preceding the in vivo phosphosites observed in PC-3 cells. Those were supplemented with close-by sites that had been phosphorylated by PIM1 in vitro. Stepwise mutagenesis resulted in approximately 50% (double mutant, DM) or 90% (multi mutant, MM) reduction in the ability of PIM1 to phosphorylate NFATC1 in vitro, while the previously validated PIM-selective inhibitor DHPCC-9 [4, 26] fully abrogated PIM1 autophosphorylation and PIM1-mediated NFATC1 phosphorylation (Fig. 2b). In addition to PIM1, also PIM2 and PIM3 were able to phosphorylate WT, but not MM NFATC1 in vitro (Fig. 2c).

**Both wild-type and phosphomutant NFATC1 interact with PIM1 in PC-3 cells**

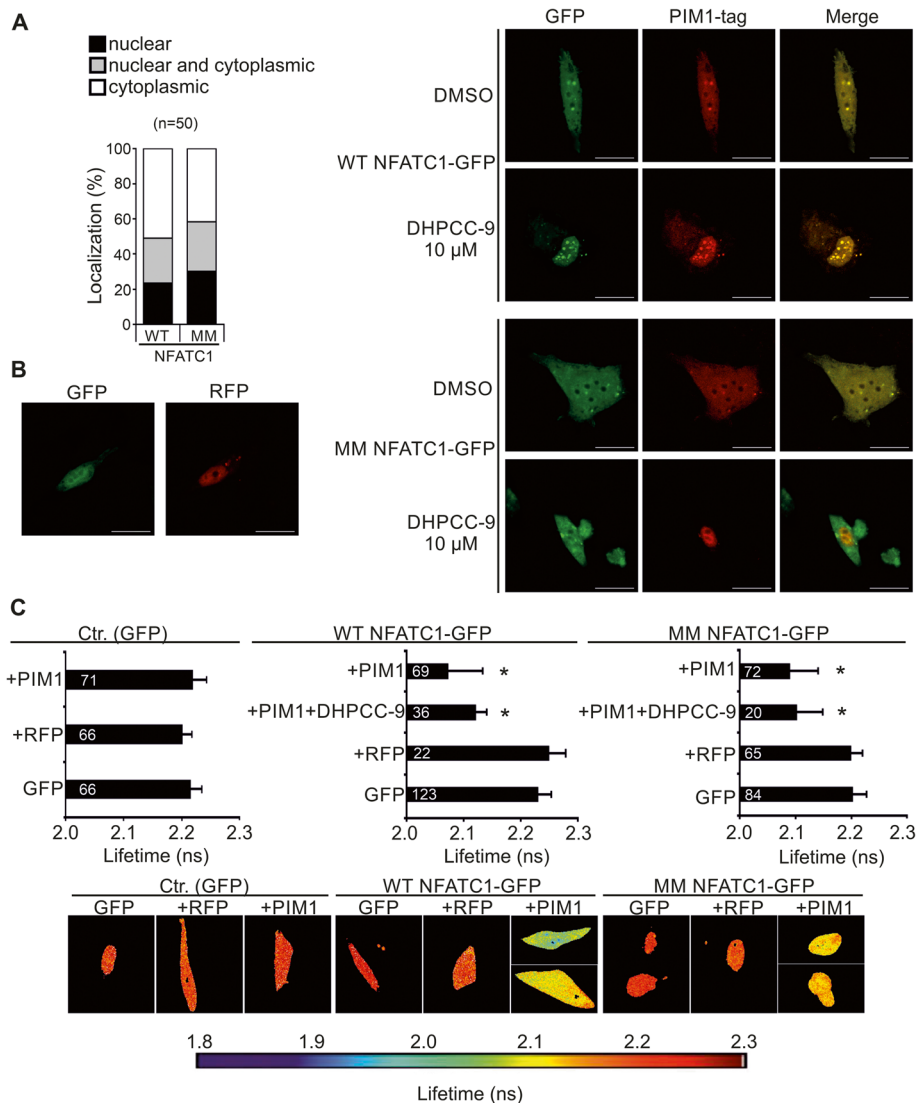
To assess the subcellular localization of WT versus phosphodeficient NFATC1, we transiently expressed them in PC-3 cells, where they showed similar localization patterns in the nucleus, in the cytoplasm or in both (Fig. 3a, Additional file 1: Figure S1C). As we had previously shown that NFATC1 and PIM1 can be co-immunoprecipitated with each other [12], we now wanted to determine whether the mutations in the PIM1 target sites affected either the colocalization or the physical interaction of GFP-tagged NFATC1 proteins with RFP-tagged PIM1. For these purposes, confocal microscopy and the fluorescence-lifetime imaging method (FLIM) were used as in our previous studies [23]. Both WT and MM NFATC1 showed nuclear co-localization (Fig. 3b) as well as interaction (Fig. 3c) with PIM1, as was evident from merged confocal images and from reduced lifetimes of GFP signals, respectively. Furthermore, the PIM inhibitor DHPCC-9 did not have major effects on the localizations or interactions, indicating that PIM-induced phosphorylation was not required there (Fig. 3b, c).

**Phosphorylation by PIM1 promotes NFATC1 activity**

To determine whether phosphorylation affects transcriptional activity of NFATC1, we transiently overexpressed WT and mutant NFATC1 proteins in three different prostate cancer cell lines, PC-3, DU-145 and LNCaP cells. Both PC-3 and DU-145 cells represent hormone-





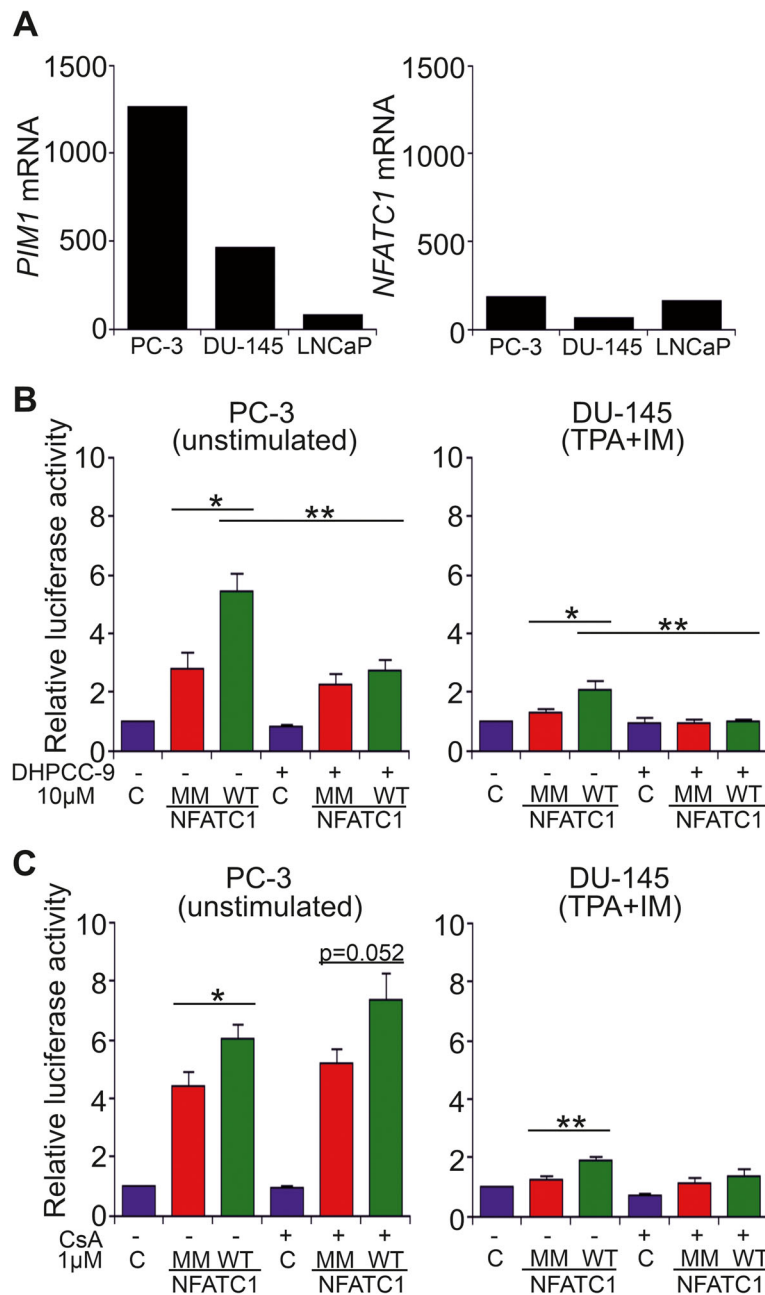


**Fig. 3** PIM1 interacts with NFATC1 in PC-3 cells. **a** Wild-type (WT) or multi mutant (MM) NFATC1 were transiently expressed in PC-3 cells and their subcellular localization patterns were analysed by confocal microscopy after staining with anti-Flag antibody. Shown are means from one experiment with three parallel samples. **b** The physical interactions between RFP-tagged PIM1 and GFP-tagged WT or MM NFATC1 proteins were analysed by fluorescence-lifetime imaging microscopy (FLIM) from samples of transiently transfected PC-3 cells. 24 h after transfection, cells were treated overnight with DMSO or 10  $\mu$ M DHPCC-9. Shown on the left are representative images of negative control cells with expression of empty GFP or RFP vectors, while on the right are single channel or merged images of cells co-transfected with GFP- or RFP-tagged vectors. Scale bar 20  $\mu$ m. **c** Shown are average GFP lifetimes from two independent FLIM experiments along with sample numbers inside the black bars as well as representative images with a heatmap. Lowest negative control (GFP + RFP) value was set as the limit for physical interaction

insensitive tumors, while LNCaP cells are hormone-sensitive, but carry mutated androgen receptors [35]. Based on our previously published RNA-sequencing dataset [36], endogenous *PIM1* mRNA expression levels were relatively high in PC-3 cells, lower in DU-145 cells and lowest in LNCaP cells, while relatively low *NFATC1* mRNA levels were observed for all cell lines (Fig. 4a).

According to data from NFAT-luciferase assays, PC-3 cells had clearly higher basal NFAT activity than DU-

145 or LNCaP cells, although in DU-145 cells the activity could be increased by stimulation with TPA and the calcium ionophore ionomycin (Fig. 4b, Additional file 1: Figure S4A). This suggests that in contrast to PC-3 cells, NFAT nuclear translocation and activation are normally regulated by calcium and calcineurin in DU-145 cells. This conclusion was further supported by the ability of cyclosporin to slightly suppress NFAT activity in stimulated DU-145 cells, but not in any other cell samples (Fig. 4c, Additional file 1: Figure S4B).



**Fig. 4** Effects of PIM-dependent phosphorylation on NFAT activity. **a** The expression levels of *PIM1* and *NFATC1* mRNAs were determined from our previously published RNA-sequencing dataset from PC-3, DU-145 and LNCaP cell lines [36]. **b** The impact of PIM-dependent phosphorylation on NFAT activity was analysed by luciferase assays in PC-3 and DU-145 cells that transiently expressed wild-type (WT) or multi mutant (MM) NFATC1. Cells were treated with either DMSO (-) or 10 µM DHPCC-9 (+). In addition, DU-145 cells had been pre-treated with TPA and IM. Shown are means of relative luciferase activities from two independent experiments with four parallel samples, the results of which had been normalized against the mock-transfected control samples. **c** Similar luciferase assays were performed also with cells treated with either EtOH (-) or 1 µM CsA (+)

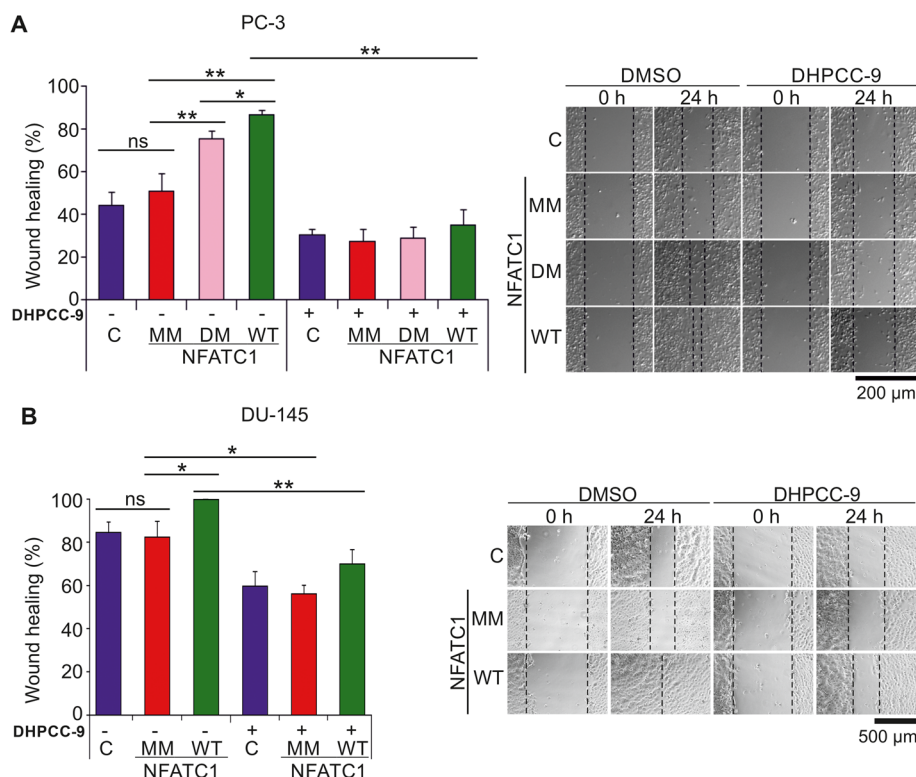
The presence of overexpressed WT NFATC1 strongly enhanced NFAT activity in all three cell lines, while mutations in the PIM1 target sites or treatment of cells with the PIM inhibitor DHPCC-9 resulted in significantly compromised NFAT activities in PC-3 and DU-145 cells, but not in LNCaP cells (Fig. 4b, Additional file 1: Figure S4). These results indicated that full NFAT activity was dependent on phosphorylation of PIM target sites in PC-3 and DU145 cells, but PIM-independent in LNCaP cells, where nearly negligible *PIM* mRNA levels had been observed (Fig. 4a).

**Prostate cancer cell motility is regulated by NFATC1 phosphorylation**

As we had previously shown that PIM inhibition blocks the pro-migratory effects of NFATC1 in PC-3 cells [4], we wanted to investigate the role of NFATC1 phosphorylation in this context. In wound healing assays with PC-3 cells transiently overexpressing WT NFATC1 or phosphomutants, mutations in the PIM target sites significantly reduced the ability of NFATC1 to promote cell migration (Fig. 5a). While there were minor effects by the DM mutant and more pronounced effects by the MM mutant, PIM inhibition by DHPCC-9 completely

blocked cell migration in each case. Similar wound healing experiments were also performed with DU-145 prostate cancer cells, which transiently overexpressed WT NFATC1 or the multi mutant. As in PC-3 cells, mutations in the PIM target sites abolished the ability of NFATC1 to promote cell migration (Fig. 5b). Also DHPCC-9 diminished motility, but less efficiently than in PC-3 cells, which migrated slightly slower than DU-145 cells.

Western blotting was used to confirm equivalent protein levels of NFATC1 and PIM family members in PC-3 cells (Additional file 1: Figure S3A). DHPCC-9 slightly reduced them, but did not significantly affect cell viability (Additional file 1: Figure S3A, B). Similar viability and protein expression data were obtained also from DU-145 cells (Additional file 1: Figure S3C). Additional wound healing assays were performed in PC-3 cells in the presence of mitomycin C to exclude effects of cell proliferation on cell migration (Additional file 1: Figure S3D), but no major differences were observed as compared to its absence (Fig. 5a). More interestingly, the triple mutant (TM) NFATC1 with intact S245 and S269 sites blocked cell migration almost as efficiently as MM lacking them, suggesting that the pro-migratory



**Fig. 5** Lack of PIM1 target sites reduces the ability of NFATC1 to promote migration of prostate cancer cells. Wild-type (WT), double mutant (DM) or multi mutant (MM) NFATC1 were transiently expressed in PC-3 cells (a) or DU-145 cells (b). For wound healing assays, cell layers were scratched 24 h after transfection with a 10 µl pipette tip and the wounded areas were allowed to recover for another 24 h in the presence of either DMSO or 10 µM DHPCC-9. Shown are representative pictures taken at 0 h and 24 h time-points, and average wound healing percentages

effects of NFATC1 were more dependent on phosphorylation of other PIM1 target sites.

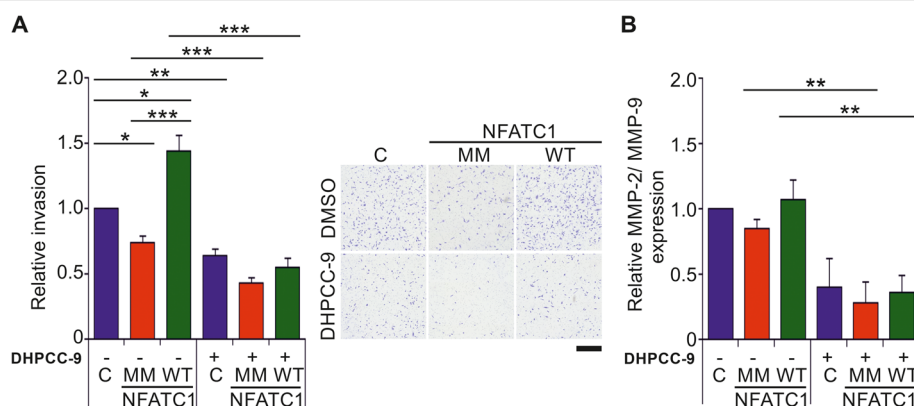
To investigate the role of NFATC1 phosphorylation in cell invasion, we carried out matrigel-based Boyden chamber invasion assays. There WT NFATC1 increased invasion of transiently transfected PC-3 cells through the membranes, while mutations in PIM targets sites in MM NFATC1 or the presence of the PIM inhibitor DHPCC-9 decreased it (Fig. 6a). NFATC1 protein levels were also monitored by western blotting in the invasion experiments (Additional file 1: Figure S3E). No major differences were observed in cell viability, except for a slight increase by MM NFATC1 at the later 72 h time-point (Additional file 1: Figure S3F). As activities of matrix metalloproteinases (MMPs), such as MMP-2 and MMP-9 are needed for cell invasion and may be regulated in an NFAT-dependent fashion also in our cells of interest [2], we analysed the effects of NFATC1 phosphorylation on their expression by gelatinase activity assays. The relative MMP expression levels were slightly, although not significantly reduced by MM NFATC1, while the decrease was more prominent with the PIM inhibitor DHPCC-9 (Fig. 6b). In each case, the MMP enzymatic activities correlated well with data from the invasion assays, suggesting that MMPs are relevant NFATC1 targets, whose activities can be indirectly regulated by PIM kinases.

#### ***ITGA5* is a putative target for phosphorylated NFATC1**

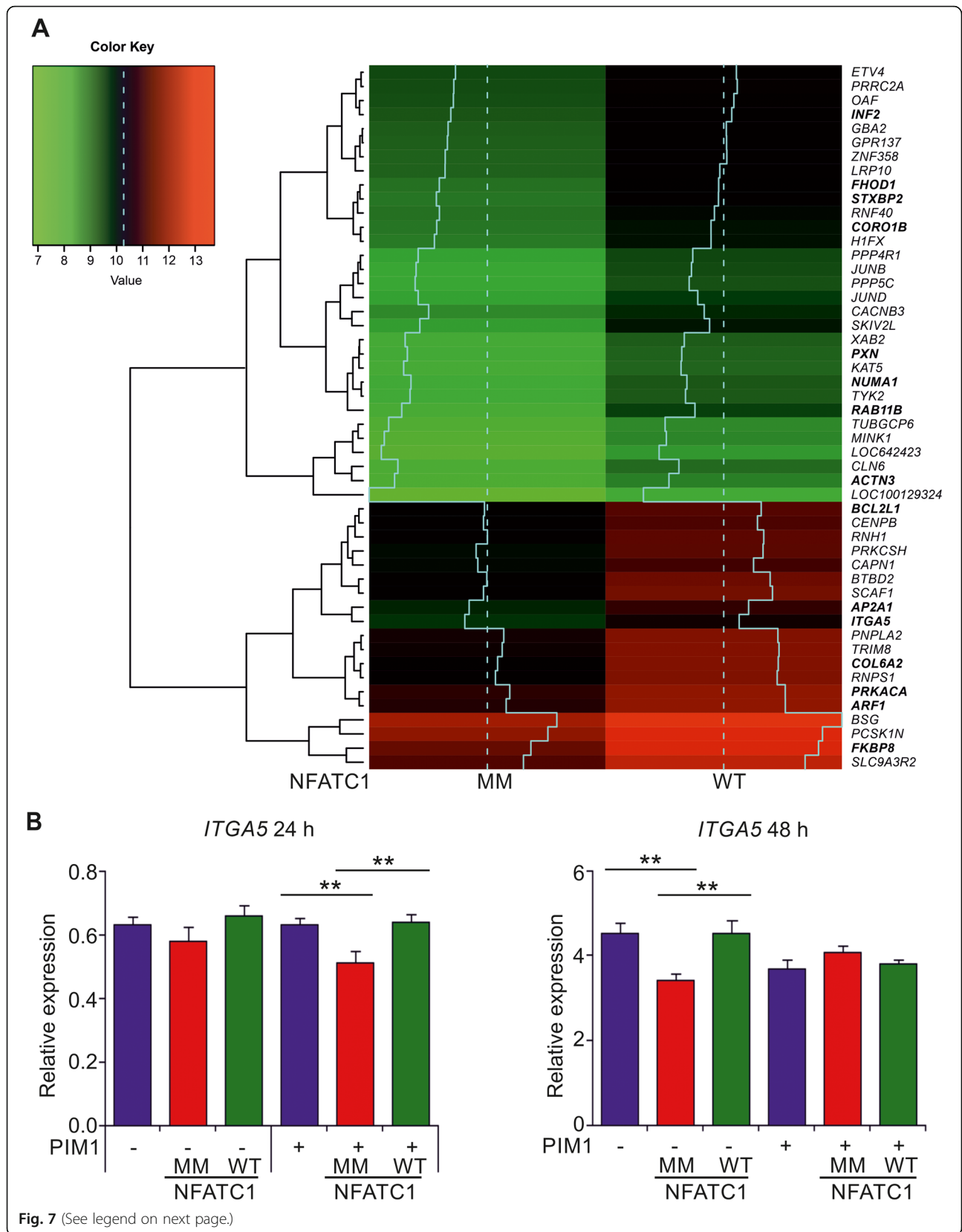
To identify additional targets for the interplay between PIM1 and NFATC1, we designed microarray experiments, where we compared mRNA transcriptomes of PC-3 cells with or without stable overexpression of

PIM1, and with or without transient overexpression of either WT or MM NFATC1. Real-time qPCR was first used to confirm overexpression of PIM1 and/or NFATC1 genes in the cell samples (Additional file 1: Figure S4A-B). With the microarrays, we performed three different types of comparisons: First, we compared parental PC-3 cells to their derivatives that stably expressed PIM1, to identify the genes that are up- or downregulated by elevated PIM1 expression. Secondly, we compared the PC-3 cells that had transiently been transfected with WT or MM NFATC1, to find genes that are controlled by the levels of NFATC1 activity. Finally, we compared PC-3 cells that both stably overexpressed PIM1 and transiently expressed WT or MM NFATC1, to unravel the genes regulated by PIM1-dependent phosphorylation of NFATC1. Genes with altered expression profiles in these three comparisons are listed in Additional file 1: Table S4.

Clustering analyses revealed that the cells overexpressing PIM1 and WT NFATC1 have a different profile as compared to the other samples (Additional file 1: Figure S4C). All the genes listed in Fig. 7a and Additional file 1: Table S4 showed higher mRNA levels in cells with WT NFATC1 than with MM, and their levels were lower also in the other control samples (Additional file 1: Figure S4C). Based on the observed gene expression profiles, we performed a canonical pathway analysis to determine, which cellular functions are primarily affected by the PIM-NFATC1 axis. We discovered five pathways that had significantly been enriched, many of which regulate cell adhesion and motility-related functions, like integrin, paxillin and FAK-signaling pathways (Additional file 2: Figure S6).



**Fig. 6** Lack of PIM1 target sites reduces also the ability of NFATC1 to enhance invasiveness of prostate cancer cells. **a** For invasion assays, PC-3 cells were grown in Boyden chambers in the absence (–) or presence (+) of 10  $\mu$ M DHPCC-9. After 48 h, cells that had invaded through the membranes were fixed, stained with crystal violet and counted. Shown are relative invasion rates from two separate experiments with triplicate samples, the results of which had been normalized against the mock-transfected control samples. Shown are also representative pictures of the effects of wild-type (WT) or multi mutant (MM) NFATC1 on cell invasion after 48 h. Scale bar 500  $\mu$ m. **b** Matrix metalloprotease (MMP) expression levels were measured by gelatinase activity assays from invasion sample media. Shown are relative MMP-2/MMP-9 expression levels from two separate experiments with three parallel samples



(See figure on previous page.)

**Fig. 7** Microarray analysis reveals *ITGA5* as a putative PIM1/NFATC1 target gene. **a** Heatmap of the potential PIM1/NFATC1 target genes found from microarray analysis. Shown are fifty genes with highest log<sub>2</sub> fold changes (logFC ≥ 1 and P-value ≤ 0.05), when PC-3 cells expressing PIM1 plus multi mutant (MM) NFATC1 were compared to cells expressing PIM1 plus wild-type (WT) NFATC1. Dashed line indicates the median of the expression values and solid line shows the expression levels more precisely in a diagrammatic form. Genes listed in bold are reviewed in more detail in the discussion. **b** Relative expression levels of *ITGA5* mRNA were analysed by real-time qPCR from microarray samples (right panel) and from another independent data set (left panel) after transient transfections of WT or MM NFATC1 to PC-3 cells without (–) or with (+) stable PIM1 overexpression. The data were normalized against *TBP* expression levels

To validate the microarray data, we selected integrin alpha 5 (*ITGA5*) for more detailed expression analysis, as it is involved in the regulation of cell adhesion to matrices such as fibronectin [37, 38], and as we have previously connected PIM inhibition to decreased adhesion to fibronectin [39]. When we compared the expression levels of *ITGA5* mRNA between one independent data set (24 h after transfections) with microarray samples (48 h after transfections), we observed decreased expression in cells with MM NFATC1 as compared to control cells or cells with WT NFATC1 (Fig. 7b). These differences resembled those observed in MMP assays (Fig. 6b), and were statistically significant after 24 h in the cells stably overexpressing PIM1 and after 48 h in the control cell line, suggesting a role for PIM1-mediated phosphorylation and activation of NFATC1 in regulating *ITGA5* mRNA expression levels.

Our data prompted us to examine clinical prostate cancer samples for their expression levels of *PIM1*, *NFATC1* and/or *ITGA5* mRNAs. Therefore, we performed pairwise comparisons of these three genes in three independent prostate cancer patient-derived datasets [31–33]. The expression levels of *PIM1* and *ITGA5* or *NFATC1* and *ITGA5* mRNAs positively correlated in all datasets (Additional file 2: Figure S5). Most interestingly, the positive correlation between *NFATC1* and *ITGA5* increased along the Gleason score, with the strongest correlation in prostate cancer patients with Gleason ≥ 8.

## Discussion

Here we have analysed the functional interactions of PIM and NFATC1 proteins in several prostate cancer cell lines. We have identified multiple PIM target sites in NFATC1 that are phosphorylated in vitro and/or in cells, and are essential for the transcriptional activity of NFATC1 as well as for its pro-migratory and pro-invasive effects. By contrast, the physical interactions or colocalization of PIM1 and NFATC1 are not affected by PIM-dependent phosphorylation. In addition to PIM1, also PIM2 and PIM3 can phosphorylate NFATC1, adding it to the growing list of substrates shared by all PIM family members [16].

While our study was in progress, additional kinases targeting NFATC1 were identified. Phosphorylation by

the IκB kinase epsilon (IKKε) was shown to inhibit NFATC1 activity [40], whereas phosphorylation by the DYRK1A kinase increased NFATC1 protein stability by interfering with NFATC1 ubiquitination and degradation [41]. We identified two IKKε target sites (Ser151 and Ser161) and one DYRK1A site (Ser278) as cellular phosphorylation sites of NFATC1 in PC-3 cells, but our multi mutant NFATC1 protein lacked only one of them (Ser 151), suggesting that the effects of the mutant were mostly due to lack of PIM-dependent phosphorylation. This conclusion was further supported by our observations that the double mutant lacking known PKA target sites (Ser245 and Ser269; 11) promotes cell migration nearly as efficiently as wild-type NFATC1, while the triple mutant with intact PKA target sites inhibits cell motility almost as much as the multi mutant.

In this study, we have shown that the PC-3 prostate cancer cells exhibit constitutive NFAT activity. This is in contrast to most cells, where upstream activation of the calcium- and calcineurin-dependent pathway is required to allow NFAT family members to enter the nucleus and stimulate transcription there [1, 2]. This may not be a general feature of prostate cancer cells, since in another hormone-insensitive cell line, DU-145, NFAT activity could be enhanced by the calcium ionophore ionomycin and inhibited by the calcineurin inhibitor cyclosporin A. Yet in both cell lines, the transcriptional as well as pro-migratory activities of NFATC1 were similarly compromised by mutations in the PIM target sites. As the PIM-selective inhibitor DHPCC-9 blocked the activities of NFATC1 even more efficiently, this suggests that it affects additional downstream targets, only some of which are shared by PIM1 and NFATC1.

In our microarray analyses of transfected PC-3 cell samples, we were able to identify novel putative PIM1/NFATC1 target genes, which were more abundantly expressed in the presence of both PIM1 and wild-type NFATC1, but less in cells expressing the multi mutant NFATC1 or in other types of control cells. Thus, expression of all these target genes may be upregulated by PIM1-dependent phosphorylation of NFATC1. The putative PIM1/NFATC1 target genes included one encoding for the known PIM substrate NUMA1 (nuclear mitotic apparatus protein 1 [42]). Otherwise the target genes could be divided into several groups based on the

types of proteins encoded by them, including regulators of transcription, cell cycle, cell survival, cell motility, cell adhesion as well as intracellular trafficking. As expected, there were several genes involved in the NFAT signaling pathway [43], such as those encoding the catalytic subunit alpha of protein kinase A (PRKACA) and the FK506 immunosuppressant-binding immunophilin protein FKBP8. The latter protein also acts as a chaperone for the anti-apoptotic BCL2 protein, the expression levels of which have previously been shown to be upregulated by PIM kinases [44]. In addition, the BCL2 homolog BCL2L1 was listed there as also several genes encoding proteins involved in intracellular trafficking (RAB11B, STXBP2, AP2A1, ARF1). Maybe most interestingly in regard to our data on promotion of prostate cancer cell motility by PIM1 and NFATC1, there were several genes encoding regulators of the cytoskeletal actin network (INF2, FHOD1, ACTN3, CORO1B) and cell adhesion (COL6A2, PXN, ITGA5).

As signaling pathways involving integrins were highly enriched in our canonical pathway analysis, we picked ITGA5 for further expression analyses. Integrins are well-known cellular adhesion receptors that connect cells to the extracellular matrix and have been implicated in multiple steps of tumorigenesis [45]. ITGA5 has an essential role in cell adhesion, migration and tumor invasion [46–48]. Interestingly, previous experiments have linked both PIM and NFAT family members to integrin-mediated cell adhesion or motility. NFATC1 binds to the *ITGB3* promoter in osteoclast precursor cells, while NFATC2 and NFAT5 promote ITGA6/ITGB4-mediated cell invasion in breast cancer [49, 50]. Furthermore, PIM inhibition decreases cell adhesion to collagen and fibronectin matrices via different integrin subunits [38]. While no clear PIM-dependent changes in integrin activity or expression have previously been reported, we now found correlations between *PIM1* or *NFATC1* mRNA expression levels with *ITGA5*, both in PC-3 cells and in prostate cancer patient-derived samples. However, more detailed studies are needed to determine how critical *ITGA5* or other genes identified by the microarray analyses are in mediating the pro-motility effects of PIM and/or NFATC1 proteins.

## Conclusions

In conclusion, we have shown that phosphorylation of PIM1 target sites stimulates the transcriptional activity of NFATC1 and enhances its ability to promote prostate cancer cell migration and invasion. Thereby, the interplay between PIM kinases and NFATC1 may also provide possibilities for therapeutic interventions against metastatic prostate cancer through combinatory approaches involving PIM-selective kinase inhibitors.

## Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s12964-019-0463-y>.

**Additional file 1: Table S1.** Primers for site-directed mutagenesis in NFATC1. **Table S2.** Primers for qRT-PCR. **Table S3.** Novel NFATC1 phosphorylation sites. **Table S4.** Phosphorylation-dependent differences in the expression of PIM/NFATC1 target genes in PC-3 cells.

**Additional file 2: Figure S1.** Lack of PIM target sites does not affect subcellular localization of NFATC1. **Figure S2.** Effects of PIM-dependent phosphorylation on NFAT activity. **Figure S3.** Lack of PIM1 target sites reduces the ability of NFATC1 to promote cancer cell motility. **Figure S4.** Microarray analysis reveals phosphorylation-dependent differences in the expression of PIM/NFATC1 target genes in PC-3 cells. **Figure S5.** Integrin signaling pathway is enriched in PIM1 and NFATC1 expressing cells. **Figure S6.** ITGA5 mRNA expression levels correlates with those of *PIM1* and *NFATC1* in clinical prostate cancer samples.

## Abbreviations

ACTB:  $\beta$ -actin; CsA: Cyclosporine A; DHPCC-9: Pan-PIM inhibitor; DM: double mutant; DN: dominant negative; IM: Ionomycin; ITGA5: Integrin alpha 5; LSB: Laemmli sample buffer; MM: multi mutant; MMP: Matrix metalloproteinase; mSRR: constitutively active mutant of NFATC1; NFAT: Nuclear factor of activated T cells; ns: non-significant difference; TBP: TATA-binding protein; TM: triple mutant; TPA: 12-*o*-tetradecanoyl-phorbol-13-*o*-acetate; WT: wild-type

## Acknowledgements

We thank G.R. Crabtree for the NFATC1 expression and reporter constructs, P. Moreau for DHPCC-9, O. Silvennoinen for the Renilla reporter plasmid, H. Ahlfors for helpful discussions, M. Ruuska, S. Himanen and K. Ikkala for technical assistance, and the Biocenter Finland facilities (Turku Proteomics Facility, Turku Cell Imaging and Cytometry Core, and the Tampere Imaging Facility) for assistance in mass spectrometry and microscopy.

## Authors' contributions

PJK, EMR, SKE and NMS conceived and designed the study, and PJK, EMR, LL and TV supervised it. SKE, NMS, EMR, SR, PK and MS acquired data, and GC, GS, AS and DG helped to analyse and interpret them. SKE, NMS and PJK wrote the manuscript. All authors read and approved the final manuscript.

## Authors' information

Not applicable.

## Funding

This work was supported by grants from the Academy of Finland (grant 287040 to PJK, grants 313921 and 314558 to PR, grants 275151 and 292307 to DG), Sigrid Juselius Foundation, Cancer Society of Finland, Competitive State Research Financing of the Expert Responsibility area of Tampere University Hospital (to TV), the Pirkanmaa Fund of the Finnish Cultural Foundation (grant 0116947–3 to SE) and grant from Paulo Foundation to MS. The funding agencies had no roles in design of the study, in collection, analysis or interpretation of data, or in writing the manuscript.

## Availability of data and materials

The microarray data has been deposited to the Gene Expression Omnibus (GEO, National Center for Biotechnology Information, Bethesda, MD, USA) with series entry number GSE120133, and is available from there: <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE120133>. Other experimental data sets used and analysed during the current study as well as materials prepared are available from the corresponding author on reasonable request.

## Ethics approval and consent to participate

The use of clinical material was approved by the ethical committee of the Tampere University Hospital (TAUH, Tampere, Finland).

## Consent for publication

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

**Author details**

<sup>1</sup>Department of Biology, University of Turku, Vesilinnantie 5, FI-20500 Turku, Finland. <sup>2</sup>Faculty of Medicine and Health Technology, Tampere University and Tays Cancer Center, Tampere University Hospital, Tampere, Finland. <sup>3</sup>Turku Centre for Biotechnology, University of Turku, Turku, Finland. <sup>4</sup>Institute of Biomedicine, University of Eastern Finland, Kuopio, Finland. <sup>5</sup>University of Helsinki, Helsinki, Finland. <sup>6</sup>Signal processing laboratory, Tampere University of Technology, Pori, Finland. <sup>7</sup>Fimlab Laboratories, Tampere, Finland.

Received: 18 June 2019 Accepted: 22 October 2019

Published online: 15 November 2019

**References**

- Rao A, Luo C, Hogan PG. Transcription factors of the NFAT family: regulation and function. *Annu Rev Immunol.* 1997;15:707–47.
- Mancini M, Toker A. NFAT proteins: emerging roles in cancer progression. *Nat Rev Cancer.* 2009;9:810–20.
- Li L, Duan Z, Yu J, Dang HX. NFATc1 regulates cell proliferation, migration, and invasion of ovarian cancer SKOV3 cells in vitro and in vivo. *Oncol Rep.* 2016;36:918–28.
- Santio NM, Vahakoski RL, Rainio EM, Sandholm JA, Virtanen SS, Prudhomme M, et al. Pim-selective inhibitor DHPCC-9 reveals Pim kinases as potent stimulators of cancer cell migration and invasion. *Mol Cancer.* 2010;9:279.
- Seifert A, Rau S, Küllertz G, Fischer B, Santos AN. TCDD induces cell migration via NFATc1/ATX-signaling in MCF-7 cells. *Toxicol Lett.* 2009;184:26–32.
- Wang L, Wang Z, Li J, Zhang W, Ren F, Yue W. NFATc1 activation promotes the invasion of U251 human glioblastoma multiforme cells through COX-2. *Int J Mol Med.* 2015;35:1333–40.
- Liu Y, Liang T, Qiu X, Ye X, Li Z, Tian B, Yan D. Down-regulation of Nfatc1 suppresses proliferation, migration, invasion, and Warburg effect in prostate cancer cells. *Med Sci Monit.* 2019;25:1572–81.
- Kavitha CV, Deep G, Gangar SC, Jain AK, Agarwal C, Agarwal R. Silibinin inhibits prostate cancer cells- and RANKL-induced osteoclastogenesis by targeting NFATc1, NF- $\kappa$ B, and AP-1 activation in RAW264.7 cells. *Mol Carcinog.* 2012;53:169–80.
- Ouyang Z, Guo X, Chen X, Liu B, Zhang Q, Yin Z, Zhai Z, Qu X, Liu X, Peng D, Shen Y, Liu T, Zhang Q. Hypericin targets osteoclast and prevents breast cancer-induced bone metastasis via NFATc1 signaling pathway. *Oncotarget.* 2018;9:1868–84.
- Beals CR, Sheridan CM, Turck CW, Gardner P, Crabtree GR. Nuclear export of NF-ATc enhanced by glycogen synthase kinase-3. *Science.* 1997;275:1930–4.
- Sheridan CM, Heist EK, Beals CR, Crabtree GR, Gardner P. Protein kinase a negatively modulates the nuclear accumulation of NF-ATc1 by priming for subsequent phosphorylation by glycogen synthase kinase-3. *J Biol Chem.* 2002;277:48664–76.
- Rainio EM, Sandholm J, Koskinen PJ. Cutting edge: transcriptional activity of NFATc1 is enhanced by the Pim-1 kinase. *J Immunol.* 2002;168:1524–7.
- Glazova M, Aho TL, Palmethofer A, Murashov A, Scheinin M, Koskinen PJ. Pim-1 kinase enhances NFATc activity and neuroendocrine functions in PC12 cells. *Brain Res Mol Brain Res.* 2005;138:116–23.
- Eichmann A, Yuan L, Bréant C, Alitalo K, Koskinen PJ. Developmental expression of Pim kinases suggests functions also outside of the hematopoietic system. *Oncogene.* 2000;19:1215–24.
- Bachmann M, Möröy T. The serine/threonine kinase Pim-1. *Int J Biochem Cell Biol.* 2005;37:726–30.
- Santio NM, Koskinen PJ. PIM kinases: from survival factors to regulators of cell motility. *Int J Biochem Cell Biol.* 2017;93:74–85.
- Valdman A, Fang X, Pang ST, Ekman P, Egevad L. Pim-1 expression in prostatic intraepithelial neoplasia and human prostate cancer. *Prostate.* 2004;60:367–71.
- Xu Y, Zhang T, Tang H, Zhang S, Liu M, Ren D, Niu Y. Overexpression of PIM-1 is a potential biomarker in prostate carcinoma. *J Surg Oncol.* 2005;92:326–30.
- Cibull TL, Jones TD, Li L, Eble JN, Ann Baldrige L, Malott SR, Luo Y, Cheng L. Overexpression of pim-1 during progression of prostatic adenocarcinoma. *J Clin Pathol.* 2006;59:285–8.
- Van der Poel HG, Zevenhoven J, Bergman AM. Pim1 regulates androgen-dependent survival signaling in prostate cancer cells. *Urol Int.* 2010;84:212–20.
- Qu Y, Zhang C, Du E, Wang A, Yang Y, Guo J, et al. Pim-3 is a critical risk factor in development and prognosis of prostate cancer. *Med Sci Monit.* 2016;22:4254–60.
- Santio NM, Eerola SK, Paatero I, Yli-Kauhalauma J, Anizon F, Moreau P, et al. Pim kinases promote migration and metastatic growth of prostate cancer xenografts. *PLoS One.* 2015;10:e0130340.
- Santio NM, Landor SKJ, Vahtera L, Ylä-Pelto J, Paloniemi E, Imanishi SY, et al. Phosphorylation of Notch1 by Pim kinases promotes oncogenic signaling in breast and prostate cancer cells. *Oncotarget.* 2016a;7:43220–38.
- Beals CR, Clipstone NA, Ho SN, Crabtree GR. Nuclear localization of NF-ATc by a calcineurin-dependent, cyclosporin-sensitive intramolecular interaction. *Genes Dev.* 1997;11:824–34.
- Kiriiazis A, Vahakoski RL, Santio NM, Arnaudova R, Eerola SK, Rainio EM, et al. Tricyclic Benzo[c]azulenes selectively inhibit activities of Pim kinases and restrict growth of Epstein-Barr virus-transformed cells. *PLoS One.* 2013;8:e55409.
- Akué-Gédu R, Rossignol E, Azzaro S, Knapp S, Filippakopoulos P, Bullock AN, et al. Synthesis, kinase inhibitory potencies, and in vitro antiproliferative evaluation of new Pim kinase inhibitors. *J Med Chem.* 2009;52:6369–81.
- Kouvonen P, Rainio EM, Suni V, Koskinen P, Corthals GL. Enrichment and sequencing of phosphopeptides on indium tin oxide coated glass slides. *Mol BioSyst.* 2011;7:1828–37.
- Imanishi SY, Kochin V, Ferraris SE, de Thonel A, Pallari HM, Corthals GL, et al. Reference-facilitated phosphoproteomics: fast and reliable phosphopeptide validation by microLC-ESI-Q-TOF MS/MS. *Mol Cell Proteomics.* 2007;6:1380–91.
- Virtanen SS, Väänänen HK, Härkönen PL, Lakkakorpi PT. Alendronate inhibits invasion of PC-3 prostate cancer cells by affecting the mevalonate pathway. *Cancer Res.* 2002;62:2708–14.
- Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. Limma powers differential expression analyses for RNA-seq and microarray studies. *Nucleic Acids Res.* 2015;43:e47.
- The Cancer Genome Atlas Research Network. The molecular taxonomy of primary prostate Cancer. *Cell.* 2015;163:1011–25.
- Taylor BS, Schultz N, Hieronymus H, Gopalan A, Xiao Y, Carver BS, et al. Integrative genomic profiling of human prostate cancer. *Cancer Cell.* 2010;18:11–22.
- Annala M, Kivinummi K, Tuominen J, Karakurt S, Granberg K, Latonen L, et al. Recurrent SKL-activating rearrangements in ETS-negative prostate cancer. *Oncotarget.* 2015;6:6235–50.
- Palaty CK, Clark-Lewis I, Leung D, Pelech SL. Phosphorylation site substrate specificity determinants for the Pim-1 protooncogene-encoded protein kinase. *Biochem Cell Biol.* 1997;75:153–62.
- Veldscholte J, Ris-Stalpers C, Kuiper GG, Jenster G, Berrevoets C, Claassen E, van Rooij HC, Trapman J, Brinkmann AO, Mulder E. A mutation in the ligand binding domain of the androgen receptor of human LNCaP cells affects steroid binding characteristics and response to anti-androgens. *Biochem Biophys Res Commun.* 1990;173:534–40.
- Ylipää A, Kivinummi K, Kohvakka A, Annala M, Latonen L, Scaravilli M, Kartasalo K, Leppänen SP, Karakurt S, Seppälä J, Yli-Harja O, Tammela TL, Zhang W, Visakorpi T, Nykter M. Transcriptome sequencing reveals PCAT5 as a novel ERG-regulated long noncoding RNA in prostate Cancer. *Cancer Res.* 2015;75:4026–31.
- Desgrosellier JS, Cheresch DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer.* 2010;10:9–22.
- Cai X, Liu C, Zhang TN, Zhu YW, Dong X, Xue P. Down-regulation of FN1 inhibits colorectal carcinogenesis by suppressing proliferation, migration, and invasion. *J Cell Biochem.* 2018;119:4717–28.
- Santio NM, Salmela M, Arola H, Eerola SK, Heino J, Rainio EM, Koskinen P. The PIM1 kinase promotes prostate cancer cell migration and adhesion via multiple signalling pathways. *Exp Cell Res.* 2016b;342:113–24.
- Zhang J, Feng H, Zhao J, Feldman ER, Chen SY, Yuan W, Huang C, Akbari O, Tibbetts SA, Feng P. I $\kappa$ B kinase  $\epsilon$  is an NFATc1 kinase that inhibits T cell immune response. *Cell Rep.* 2016;16:405–18.
- Liu H, Wang K, Chen S, Sun Q, Zhang Y, Chen L, Sun X. NFATc1 phosphorylation by DYRK1A increases its protein stability. *PLoS One.* 2017;12:e0172985.
- Bhattacharya N, Wang Z, Davitt C, McKenzie IF, Xing PX, Magnuson NS. Pim-1 associates with protein complexes necessary for mitosis. *Chromosoma.* 2002;111:80–95.



43. Belinky F, Nativ N, Stelzer G, Zimmerman S, Iny Stein T, Safran M, Lancet D. PathCards: multi-source consolidation of human biological pathways. Database. 2015. <https://doi.org/10.1093/database/bav006>.
44. Lilly M, Sandholm J, Cooper JJ, Koskinen PJ, Kraft A. The PIM-1 serine kinase prolongs survival and inhibits apoptosis-related mitochondrial dysfunction in part through a bcl-2-dependent pathway. *Oncogene*. 1999;18:4022–31.
45. Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. *Nat Rev Cancer*. 2018;12:533–48.
46. Akiyama SK, Yamada SS, Chen WT, Yamada KM. Analysis of fibronectin receptor function with monoclonal antibodies: roles in cell adhesion, migration, matrix assembly, and cytoskeletal organization. *J Cell Biol*. 1989;109:863–75.
47. Qin L, Chen X, Wu Y, Feng Z, He T, Wang L, Liao L, Xu J. Steroid receptor coactivator-1 upregulates integrin  $\alpha_5$  expression to promote breast cancer cell adhesion and migration. *Cancer Res*. 2011;71:1742–51.
48. Wong AW, Paulson QX, Hong J, Stubbins RE, Poh K, Schrader E, Nunez NP. Alcohol promotes breast cancer cell invasion by regulating the Nm23-ITGA5 pathway. *J Exp Clin Cancer Res*. 2011;30:75.
49. Crotti TN, Flannery M, Walsh NC, Fleming JD, Goldring SR, McHugh KP. NFATc1 regulation of the human  $\beta 3$  integrin promoter in osteoclast differentiation. *Gene*. 2006;372:92–102.
50. Jauliac S, Lopez-Rodriguez C, Shaw LM, Brown LF, Rao A, Toker A. The role of NFAT transcription factors in integrin-mediated carcinoma invasion. *Nat Cell Biol*. 2002;4:540–4.

### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

