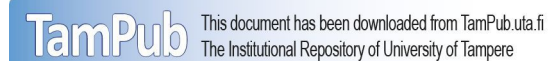


1 **FAIMS analysis of urine gaseous headspace is capable of differentiating ovarian cancer**

2



3 Riikka J Niemi ^{a,*}, Antti N Roine ^b, Emmi Eräviita ^b, Pekka S Kumpulainen ^c, Johanna U Mäenpää

4 ^{a,b}, Niku Oksala ^{b,d}

5 ^aDepartment of Obstetrics and Gynecology, Tampere University Hospital, P.O. Box 2000, 33521

6 Tampere, Finland

7 ^bFaculty of Medicine and Life Sciences, University of Tampere, P.O. Box 100, 33014 Tampere,

8 Finland

9 ^cFaculty of Biomedical Sciences and Engineering, Tampere University of Technology, P.O. Box

10 527, 33101 Tampere, Finland

11 ^dDepartment of Vascular Surgery, Tampere University Hospital, P.O. Box 2000, 33521 Tampere,

12 Finland

13 *Corresponding author: Tampere University Hospital, Department of Obstetrics and Gynecology,

14 P.O. Box 2000 FI-33521 Tampere, Finland. Tel. + 358 3 31169083. E-mail address:

15 riikka.niemi@fimnet.fi

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20

21 **Abstract**

22

23 **Aim:** We hypothesized that field asymmetric waveform ion mobility spectrometry (FAIMS) as a
24 novel artificial olfactory technology could differentiate urine of women with malignant ovarian
25 tumors from controls and women with benign tumors, based on previous findings on the ability of
26 canine olfactory system to “smell” cancer.

27 **Patients and methods:** Preoperative urine samples from 51 women with ovarian tumors, both benign
28 and malignant, and from 18 women with genital prolapse, as controls, were collected. The samples
29 were analyzed by FAIMS device. Data analysis was processed by quadratic data analysis (QDA) and
30 linear discriminant analysis (LDA), and cross-validated using 10-fold cross-validation.

31 **Results:** Thirty-three women had malignant ovarian tumors, of which 18 were high-grade cancers.
32 FAIMS distinguished controls from malignancies with the accuracy of 81.3 % (sensitivity 91.2 %
33 and specificity 63.1 %), and benign tumors from malignancies with the accuracy of 77.3 %
34 (sensitivity 91.5 % and specificity 51.4 %). Moreover, low grade tumors were also separated from
35 high grade cancers and benign ovarian tumors with accuracies of 88.7 % (sensitivity 87.8 % and
36 specificity 89.6 %) and 83.9 % (sensitivity 73.1 % and specificity 92.9 %), respectively.

37 **Conclusions:** This proof of concept-study indicates that the FAIMS from urine has potential to
38 discriminate malignant ovarian tumors from no tumor-bearing controls and benign tumors.

39

40 Key words: FAIMS; ovarian neoplasm; ovarian cancer; VOC; Owlstone Lonestar; urine

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44

45 **Introduction**

46 Annually 22,000 new ovarian cancer (OC) cases are diagnosed in the United States, and the survival
47 rates are poor due to the majority of OCs being detected at advanced stages [1]. While early diagnosis
48 and adequate cytoreductive surgery improve prognosis, there is a need for better preoperative
49 diagnostic methods for ovarian tumors.

50

51 Various ultrasound-based models have been developed for preoperative evaluation of ovarian masses.
52 These include e.g. Risk of Malignancy Index (RMI) [2] and logistic regression analyses and
53 ultrasound-based rules from the International Ovarian Tumor Analysis (IOTA)-study. Although they
54 have relatively high sensitivity and specificity, they are non-applicable for about 20 % of tumors [3].

55

56 Studies on urinary biomarkers for OC are relatively sparse. Urinary protein biomarkers, human
57 epididymis protein 4 (HE4) and mesothelin, have shown to improve the early detection of serous OC
58 compared to serum biomarkers [4]. Metabolite changes related to OC have been discovered as
59 potential biomarkers [5,6], like N¹,N¹²-diacetylspermine in polyamine analyses [7]. In addition,
60 circulating microRNAs have been shown to be abundant in urine of OC patients [8].

61

62 Many diseases are linked to distinct odors caused by volatile organic compounds (VOCs) released
63 into exhaled air, urine, blood and stool [9]. Horvath et al. trained dogs to discriminate OC patients
64 and healthy controls from tissue samples [10] and blood samples from cancer patients [11] with high
65 accuracy. The costly training, limited working capacity and cultural factors have prevented the use
66 of “sniffer dogs” in the clinic. Artificial olfaction with electronic devices could be easier to validate
67 and adopt into clinical practice [9].

68

69 Gas chromatography-mass spectrometry (GC-MS) has been used extensively in analysis of VOCs
70 but it involves complex technology and has high costs. Electronic nose (eNose) technology provides
71 a more economical and simpler way to qualitatively analyze VOCs. The technology mimics the
72 working principle of mammalian olfactory system (Figure 1). Ion mobility spectrometry (IMS) works
73 according to the same principles, providing a qualitative VOC spectrum from the sample. Field
74 asymmetric waveform IMS (FAIMS) is a modern and sensitive variant of IMS providing a high
75 sensitivity and stability [12]. The working principle of FAIMS is illustrated in Figure 2.

76

77 There is mounting evidence of the potential of eNose devices in detection of cancer from various
78 sample media [12]. FAIMS specifically has previously been shown to detect colorectal and pancreatic
79 cancers from urine [13,14]. Detection of OC has been only attempted from cancer tissue [15]. Urine
80 is a promising sampling method since it can be obtained non-invasively.

81

82 We hypothesized that FAIMS would be capable of differentiating the urine of women with OC from
83 benign ovarian tumors and controls.

84

85 **Materials and methods**

86 **Subjects and study design**

87 Between May 2013 and March 2016, 60 women with an adnexal tumor scheduled for surgery gave a
88 morning urine sample in the operation day at the Department of Obstetrics and Gynecology of
89 Tampere University Hospital. They were all postmenopausal, and none of them had an ongoing
90 treatment for cancer. After operation nine tumors were excluded due to their non-ovarian origin or a
91 concurrent malignant tumor. The final sample size after exclusions was 51. Eighteen women
92 scheduled for urinary incontinence or genital prolapse surgery were recruited as controls. The samples
93 were stored at -70°C until analysis. Because of the proof-of-concept nature of the study, no power
94 calculations could be done. The size of the study population was based on the experience from
95 previous studies with similar technology [16].

96

97 The samples were defrosted and analyzed using Owlstone Lonestar (Owlstone Inc, Cambridge,
98 United Kingdom) device which uses FAIMS technique. The sensor was coupled with ATLAS
99 sampling unit (Owlstone Inc, Cambridge, United Kingdom) that standardizes the analytical
100 conditions by controlling the temperature and dilution of the VOCs evaporated from the sample.

101

102 **Protocol of FAIMS**

103 For FAIMS analysis, we used settings previously described by Arasaradnam et al [13]. The step-by-
104 step analysis protocol was as follows:

- 105 1) Urine samples were first thawed at room temperature and analyzed in random order.
- 106 2) A 5 ml urine sample was aliquoted to a 30 ml glass vial and warmed to 40°C.
- 107 3) Once the sample achieved the target temperature, three consecutive scans were conducted to
108 minimize the effect of scan-to-scan variation.

109 4) After the analysis, the sample vial was removed from the sampling unit and a vial of 5 ml of
110 purified water was placed in to the chamber.

111 5) The vapour released from the purified water acts as a cleaning agent that removes the carry-over
112 effect of trace VOCs from the urine sample that are retained in the sensor. Five consecutive scans
113 with purified water were conducted.

114 The next urine sample was placed to the sampling chamber and the process was repeated. To ensure
115 stable and clean carrier gas for the system, we utilized standard pressurized clean air that was cleaned
116 from residual humidity with a silica gel filter and from residual VOCs with activated charcoal filter
117 before entering the system. We used the flow settings recommended by the manufacturer for urine
118 samples: The flow rate over the sample was 500 ml/min, which was mixed to 2000 ml/min stream of
119 clean air for a total flow of 2500 ml/min for the sensor. The FAIMS scanning settings used were also
120 ones provided by the manufacturer: Dispersion field from 0 to 90 % was scanned in 51 steps and
121 compensation voltage from -6 to +6 V was scanned in 512 steps. Each scan contains two ion windows,
122 one for negative and one for positive ions. One window is produced by the negative ions that collide
123 the positive detector and the other is produced by the positive ions that collide the negative detector,
124 respectively. The detectors are illustrated in Figure 2.

125 The ion window is a spectrum that has compensation voltage on the X axis and dispersion field on
126 the Y axis as seen in Figure 3. The compensation voltage is the base voltage between the electric
127 plates in the separation part of the FAIMS sensor. This biases the ion flow either towards negative or
128 positive plate. The dispersion field strength represents the strength of the electrical field between the
129 plates as a percentage of the maximum field that can be created by the system. The ion window is
130 compiled by adjusting the dispersion field strength stepwise and on each step scanning the selected
131 compensation voltage range at each step. The scans were saved on the hard drive of the Lonestar
132 system from which they were transferred to an USB drive for statistical analysis.

133

134 **Statistical methods**

135 The last of the three scans from the urine sample was found to be equal in performance when
136 compared to the average of three scans, and was taken for analysis. One scan consists of a matrix of
137 52,200 measurement values, including both positive and negative ion window. The areas with no
138 response were removed and the remaining signal was downsampled, selecting every other line and
139 column of the scan, leaving 1,536 points for each measurement.

140

141 Forward feature selection with linear discriminant analysis (LDA) and quadratic discriminant
142 analysis (QDA) were utilized to find discriminating features from each group. Both LDA and QDA
143 seek a classifier that is optimal for discrimination of the groups. LDA is a special case of QDA where
144 the covariance of each group is assumed to be equal which results in a linear discriminator whereas
145 QDA allows the covariances to differ which also enables quadratic, parable-shaped discriminators.
146 Because LDA is a simpler method, it is preferred as the first option to test. The results were cross-
147 validated by 10-fold cross-validation to avoid overfitting. In this method, the dataset is divided into
148 10 groups. One group is then excluded from the dataset and the remaining nine groups are used to
149 create the classification parameters as the training set. The excluded group is then classified using
150 these parameters. Since, due to random division for the cross validation, the classification parameters
151 change to a certain extent in every run, the process was repeated 100 times to reduce the effect of
152 variation and to calculate averages and standard deviations for classification results. The analysis was
153 conducted with MATLAB R2017b (MathWorks Inc, Natick, MA, USA).

154

155 **Results**

156 Characteristics of the final study population are presented in Table 1. The averages and standard
157 deviations of the 100 runs of QDA and LDA analysis are given in Table 2. The performances of QDA
158 and LDA seem to be mostly equal yet there is a notable difference in comparisons of benign tumors

159 with low grade vs. high grade malignant tumors, respectively. The data produced by FAIMS is
160 nonlinear by nature [17], and it is likely that nonlinear methods such as QDA yield better results in
161 most cases, especially when the differences between groups are less distinct. By QDA analysis,
162 benign ovarian tumors were distinguished from malignant tumors with sensitivity and specificity of
163 91.5 % and 51.4 %, respectively. However, the specificity improved to 79.7 % when they were
164 compared only to high-grade ovarian cancers. Even low grade ovarian malignancies were
165 discriminated from high grade ovarian cancers with sensitivity of 87.8 % and specificity of 89.6 %,
166 and from benign ovarian tumors with sensitivity of 73.1 % and specificity of 92.9 %, respectively.
167 Figure 3 shows average FAIMS outputs from urine sample of a control and of a woman with ovarian
168 cancer.

169

170 **Discussion**

171 This study provides preliminary evidence that FAIMS analysis of VOCs can discriminate urine
172 samples from OC patients, patients with non-malignant tumors and healthy controls. High grade
173 ovarian cancers seem to be separated from low grade ovarian cancers, benign ovarian tumors and
174 controls.

175

176 The study further demonstrates that OC is associated with distinct odor [18-20]. The fact that this
177 phenomenon is apparent in urine suggests that a systemic process is involved. It is apparent that
178 metastatic, systemic cancer may elicit profound changes in urine composition that may be an
179 indication of decreasing renal function. However, in the case of colorectal cancer, even early stage
180 cancers could be detected [13]. There is in fact mounting body of evidence that cancer releases VOCs
181 to systemic circulation that consequently are released through alveoli to breath and via glomerular
182 filtration to urine [21]. This suggests that breath and urine can be considered alternative sampling
183 methods for some VOCs. The feasibility of FAIMS/IMS has been demonstrated in both sampling

184 sources [13,22]. Reliable sampling from exhaled breath is challenging [23] and the performance of
185 breath VOC analysis in OC seems to be inferior to our results obtained from urine [18,24]. Since
186 urine can be obtained non-invasively, we consider it as a more promising sampling source for VOC
187 analysis in OC.

188

189 VOCs in different sample mediums and cancers seem to have common features, which are related to
190 oxidation such as benzene derivatives [13,18,21]. The metabolic origin and function of most of these
191 VOCs are unclear. They can originate from endogenous and exogenous sources and may thus be a
192 result also from environmental exposure instead of the cancer [21]. In this study we achieved a good
193 discrimination of high grade and low grade cancers. It has been suggested that KRAS and TP3
194 mutations play a role as a watershed in development of high or low grade serous OC, i.e. type I and
195 II OCs [25]. These single mutations have resulted in VOC changes in cellular model [26] that reflect
196 those found in urine in other cancers [13]. We speculate that the VOC alterations concerning various
197 mutations should be studied in future also in ovarian cancer.

198

199 This study must be considered as preliminary, and the results should be verified in larger patient
200 cohorts with this repeatable method. However, there is urgent need for early detection of especially
201 aggressive type II OCs, with an ultimate goal to improve the prognosis of this devastating disease
202 [25]. An important topic in future FAIMS research is to examine if cytoreductive surgery and
203 immunosuppressive therapy have influences on VOC emissions of urine samples. FAIMS technology
204 itself has advantages compared to GC-MS- and eNose implications; the technology by nature is
205 sensitive to trace concentrations of molecules, is considerably more economical than MS-based
206 methods, and does not suffer stability problems of other eNose technologies [27]. In contrast to canine
207 studies, FAIMS is standardized and repeatable, whereas it is almost impossible to replicate research
208 settings of canine studies because of variation in dogs.

209

210 Our study has also limitations. First, the present results cannot as such be generalized to unselected
211 populations, but rather should be considered valid in the setting of tertiary hospitals, as part of the
212 diagnostic work-up of adnexal tumors. Second, the number of analyzed urine samples was quite
213 small. However, the proportions of three patient groups (controls, benign and malignant tumors) were
214 balanced. Third, the considerable number of low malignant potential and borderline ovarian tumors
215 in our study certainly has an influence on our results comparing benign and malignant ovarian tumors,
216 and may have contributed to the rather great deviation seen between comparisons of benign tumors
217 and all or low-grade malignant tumors. However, the comparisons between benign ovarian tumors or
218 controls and high grade ovarian tumors are more accurate and specific. Fourth, the storage time of
219 our samples was several years, which may have reduced the VOC emissions and thus differences
220 between groups, as has been shown in a recent study examining the effect of storage on VOC profiles
221 of urine [28]. In addition, the effects of the diet and possible medications may have had influence on
222 the concentration and composition of urine although the samples were collected in the morning after
223 at least four hours fasting. The fact that the highest discrimination rate was achieved for benign tumors
224 and controls suggests that there is a degree of bias between patient groups. This may also result from
225 the larger and more heterogenous nature of cancer group.

226

227 **Conclusion**

228 According to our results, we propose that the VOC signature of urine of ovarian cancer patients can
229 be recognized by FAIMS and that it has potential for being a non-invasive method in the detection of
230 ovarian malignancy. Our novel study encourages us to examine further possibilities of FAIMS for
231 diagnostics and follow-up of gynecological malignancies.

232

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240 had no role in planning and execution of the study or in in the analysis and writing process of the
241 article.

242 **Conflict of interest statement**

243 RJN, EE and JUM declare no conflicts of interest. NO, PSK and ANR are shareholders of Olfactomics
244 Ltd. which is about to commercialize proprietary technology for the detection of diseases by ion
245 mobility spectrometry.

246 **Ethical conduct of research**

247 All participants gave their informed consent to the study, and the investigation was approved by the
248 Ethic committee of Tampere University Hospital.

249 **Acknowledgements**

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251

252 **References**

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- 322
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- 324

325 **Table 1.** Demographic data of study population

	Malignant tumors		Benign tumors	Controls
n	33		18	18
Age (years)				
Median	64		64	71
(range)	(51-82)		(51-73)	(55-83)
Diagnosis (n)	Low grade cancers (15) - mucinous adenocarcinoma Stage IA and IC (1+1) - endometrioid adenocarcinoma Stage IA (1) - mucinous borderline Stage IA (5) - serous borderline Stage IA (4) - Sertoli-Leydig cell tumor Stage IIC (1) - Granulosa cell tumor Stage IA (2)	High grade cancers (18) - carcinosarcoma Stage IIC (1) - high grade serous adenocarcinoma <ul style="list-style-type: none"> • Stage IC (1) • Stage IIC (1) • Stage III/IV (15) 	Serous cystadenoma (9) Mucinous cystadenoma (1) Fibroma (2) Simple cyst (3) Endometriotic cyst (2) Necrotized cyst (1)	Genital prolapse or urinary incontinence (18)

326

327 **Table 2.** Results of FAIMS signal data and QDA and LDA classification

Classification pairs	QDA			LDA			328
	Accuracy (%) (±2 Std)	Sensitivity (%) (±2 Std)	Specificity (%) (±2 Std)	Accuracy (%) (±2 Std)	Sensitivity (%) (±2 Std)	Specificity (%) (±2 Std)	329
Benign ovarian tumors vs. controls	91.9 (±9.8)	93.4 (±11.4)	90.4 (±14.4)	86.1 (±9.6)	86.0 (±11.2)	86.1 (±12.2)	330
Controls vs. malignant ovarian tumors	81.3 (±8.2)	91.2 (±7.2)	63.1 (±16.0)	81.2 (±5.8)	90.4 (±5.2)	64.3 (±12.8)	331
Controls vs. high grade ovarian cancers	81.9 (±5.2)	89.1 (±2.8)	74.6 (±9.6)	82.1 (±6.0)	88.7 (±3.2)	75.6 (±11.8)	332 333
Benign vs. malignant ovarian tumors	77.3 (±13.8)	91.5 (±6.4)	51.4 (±32.0)	65.9 (±13.8)	87.1 (±9.0)	27.1 (±38.6)	334
Benign ovarian tumors vs. low grade ovarian cancers	83.9 (±23.4)	73.1 (±41.4)	92.9 (±11.4)	59.3 (±7.0)	35.9 (±14.0)	78.8 (±5.8)	335 336
Benign ovarian tumors vs. high grade ovarian cancers	82.5 (±10.0)	85.3 (±15.0)	79.7 (±12.0)	82.5 (±9.6)	85.0 (±15.0)	79.9 (±11.2)	337 338
Low grade vs. high grade ovarian cancers	88.7 (±11.2)	87.8 (±12.8)	89.6 (±16.6)	82.0 (±10.8)	84.3 (±16.0)	79.7 (±13.4)	339

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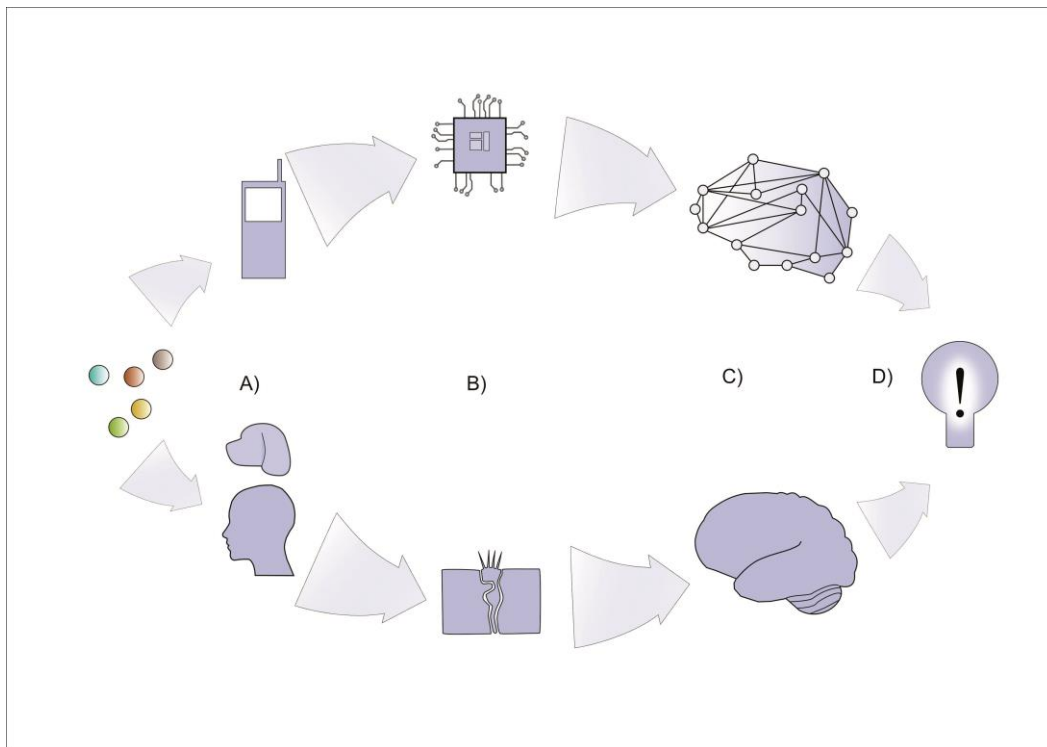
342 **Figure 1.** The working principle of mammary and eNose compared

343 A) VOCs enter a sampling unit where the humidity, the temperature and the concentration of the
344 sample are optimized.

345 B) Optimized sample enters the sensor unit where different VOCs attach to different areas of the
346 sensor and produce electrical currents.

347 C) Electrical currents are referred to a computing system for analysis where they are associated with
348 previously gathered information.

349 D) A result of the analysis is produced.



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358 **Figure 2.** Illustration on the working principle of FAIMS

359 A) Sample vial is placed in to the sampling chamber where VOCs are released from the sample.

360 VOCs are then transferred to the analyzer by clean air flow.

361 B) In the analyzer, VOCs are first ionized by a radioactive isotope and gain electrical charge.

362 C) Ionized VOCs enter separation area where they are alternately exposed to high and low electric

363 fields between the electric plates. The plates also have a baseline compensation voltage that is

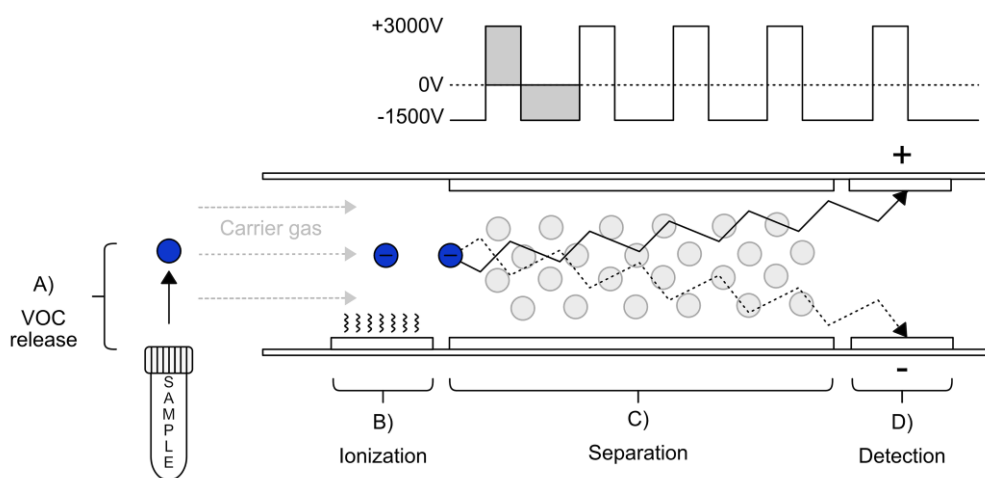
364 periodically adjusted. The different properties of VOCs cause them to travel at different speed in the

365 separation chamber and behave differently in high and low electric fields. This results in separation

366 of the VOCs according to their charge, shape and mass.

367 D) At the last stage of the analysis, VOCs collide with detectors, creating electric currents that create

368 a unique spectrum for each molecular mixture.



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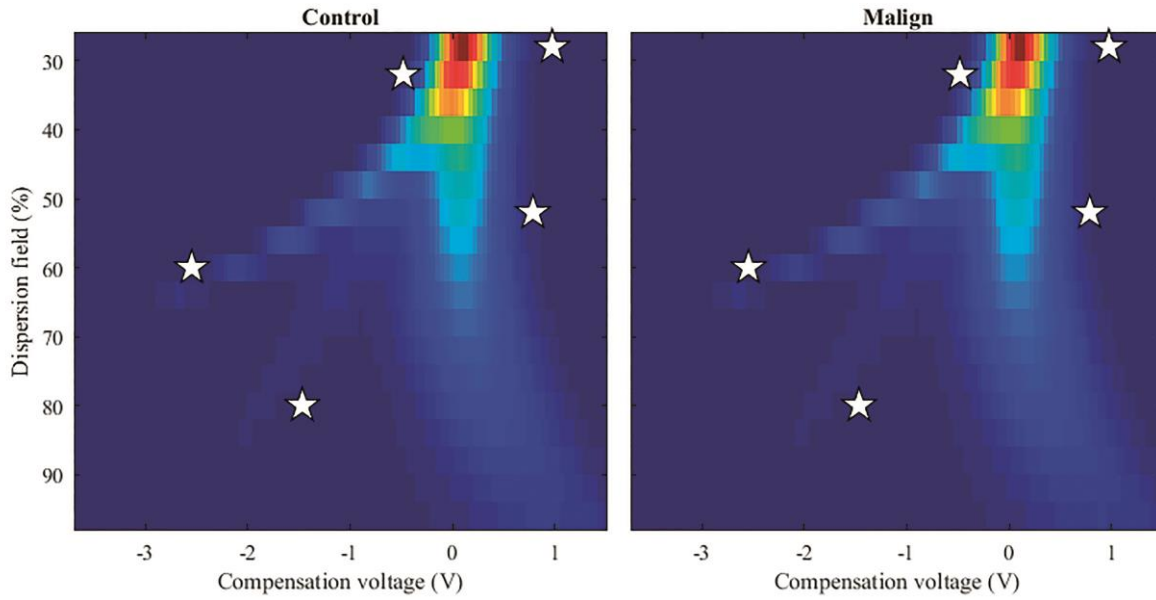
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376 **Figure 3.** Average FAIMS spectrum from a patient with ovarian cancer and from a control
377 Stars indicate the areas of the spectrum that yielded optimum discrimination of the two groups.
378 Compensation voltage is on X-axis and dispersion field strength is on Y-axis.



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