SHORT COMMUNICATION

Safety of Novel Amino-5-laevulinate Photosensitizer Precursors in Photodynamic Therapy on Healthy Human Skin

Noora Neittaanmäki-Perttu¹, Eerika Neittaanmäki², Ilkka Pölönen³, Erna Snellman⁴ and Mari Grönroos⁵

¹Department of Dermatology and Allergology, Helsinki University and Helsinki University Hospital, FIN-00029 Helsinki, ²Department of Pharmacy, Helsinki University, Helsinki, ³Department of Mathematical Information Technology, University of Jyväskylä, ⁴Department of Dermatology, Tampere University and Tampere University Hospital, Tampere, and ⁵Department of Dermatology and Allergology, Päijät-Häme Social and Health Care Group, Lahti, Finland. E-mail: noora.neittaanmaki@fimnet.fi

Accepted Apr 27, 2015; Epub ahead of print May 5, 2015

Photodynamic therapy (PDT) is a highly effective treatment for superficial skin cancers and skin cancer precursors (1, 2). In PDT, the administration of a photosensitizing drug, followed by its activation by a specific wavelength of light matching the absorbance of the sensitizer, leads to a phototoxic reaction destroying the tumour cells (3). Adverse effects include pain during the illumination, and erythema and crusting after treatment. Recently, several attempts have been made to improve the tolerability of PDT (4–6). Several topical porphyrinbased photosensitizer precursors are available for PDT. Currently, amino-5-laevulinate (5-ALA) and its methyl ester (MAL) are widely used. Novel photosensitizer formulations include 5-ALA nanoemulsion (BF-200 ALA) and a long-chain lipophilic 5-aminolaevulinate hexylester (HAL). These novel formulations can be used at low concentrations, which may increase the tolerability and reduce the costs of the treatment (7, 8). This nonsponsored double-blinded pilot study tested the safety of PDT with BF-200 ALA and HAL at 2 different concentrations compared with MAL on healthy human skin.

MATERIALS AND METHODS

The study was approved by the local ethics committee. We included 7 healthy volunteers, aged between 26 and 34 years, with skin phototypes I–II, to receive PDT on the sun-protected sides of their forearms with 4 different photosensitizer precursors: (i) MAL 16% (Metvix®, Galderma, Paris, France), (ii) BF-200 ALA (10% 5-ALA, Ameluz®, Biofrontera, Leverkusen, Germany), (iii) HAL 2%, and (iv) HAL 0.2%. The HAL creams were prepared from commercially available HAL powder (Hex-

vix®, Photocure, Oslo, Norway) by mixing it into a standard cream base (Unguentum M®, Allmirall, Madrid, Spain). The treatment sites were randomized using a web-based validated Research randomizer[©] and kept blinded from the subjects. Areas 2×1082 cm at distances of 5 cm from each other were marked on the skin and gently curettaged. The photosensitizers were weighed to achieve a 0.5 mm layer on the skin (0.5 mm-layer thickness was determined using the formula: treatment area size (mm²)* 0.5 (mg/mm²)) and kept occluded under a lightimpermeable cover for 3 h. Afterwards the drugs were wiped off and areas were irradiated using red light-emitting diode (LED) light (Aktilite CL128, Garderma, Paris, France) to achieve a light dose of 37 J/cm². Pain was recorded before, during and after the illumination using a visual analogue scale (VAS). Fluorescence images were taken before and immediately after the illumination using Wood's light (Philips Burton®, Somerset, USA) and a digital camera (Canon Ixus 10 megapixels). Fluorescence intensity and photobleaching were calculated from the images using the MatLab® (Mathworks, Natick, MA, USA). The fluorescence index in arbitrary units (AU) was calculated by measuring the mean fluorescence of the treated area and dividing it by the fluorescence of untreated skin of the same patient. The severity of the reactions at 1 and 2 days was evaluated from photographs by a blinded observer (NNP) who was unaware of the randomization. In addition, erythema was measured using a spectrometer (DermaSpectrometer®, Cortex Technology, Hadsund, Denmark) (9). Friedman's test and Pearson's correlation were used for statistical analyses.

RESULTS

Mean maximal pain during the illumination was significantly lower for HAL 0.2% and HAL 2% compared with BF-200 ALA and MAL (p=0.043). No significant difference was found in pain due to BF-200 ALA and MAL.

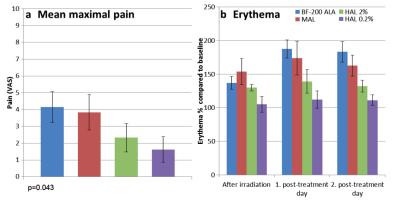


Fig. 1. (a) Mean maximal pain values during the illumination. Hexyl-ester (HAL) 0.2% and HAL 2% caused significantly less pain compared with amino-5-laevulinate (5-ALA) nanoemulsion (BF-200 ALA) and the methyl ester (MAL) of 5-ALA (p=0.043), while no difference was found in the pain between BF-200 ALA and MAL. (b) Erythema% compared with the baseline. Directly after illumination the erythema values differed between all 4 photosensitizers, p=0.0041. During 2 post-treatment days HAL0.2% caused significantly less erythema compared with BF-200 ALA (p=0.003 at the first and p=0.001 at the second post-treatment day) and MAL (p=0.043 at the first and p=0.023 on the second post-treatment day). No significant difference was found between MAL and BF-200 ALA or HAL 2%.

The mean \pm SD maximal pain scores (VAS 1–10) were 4.2 \pm 2.4 for BF-200 ALA, 3.8 \pm 2.8 for MAL, 2.3 \pm 2.2 for 2% HAL and 1.6 \pm 2.0 for 0.2% HAL (Fig. 1a).

The erythema values are shown in Fig. 1b. A significant difference was found in the erythema between HAL0.2% and BF-200 ALA (p=0.003 on the first and p=0.001 on the second post-treatment day), and between HAL0.2% and MAL (p=0.043 on the first and p=0.023 on the second post-treatment day). No significant difference was found between MAL and BF-200 ALA or HAL 2%.

The treatment reactions of the investigated compounds are summarized in Table I. The reactions were similar on the second post-treatment day (Fig. S1¹).

Reduction in the initial protoporphyrin fluorescence induced by the photosensitizers, i.e. photo-bleaching, can predict the treatment efficacy (10). After 3 h occlusion the fluorescence was equal in the BF-200 ALA, MAL and HAL2% groups (ns), but lower in the HAL 0.2% group (p=0.043). Photobleaching was equal with BF-200 ALA, MAL and HAL2%, while significantly lower photobleaching was seen with HAL0.2% (p=0.003 and p=0.023 compared with BF-200 ALA and MAL).

There was a strong positive correlation between photo-bleaching and the clinically assessed reaction severity², and a weaker positive correlation between photobleaching and erythema measured with the spectrometer³. There was no correlation between mean maximal pain and photo-bleaching, erythema or total protoporphyrin IX (PpIX) fluorescence.

DISCUSSION

These results show that low-concentration HAL was better tolerated than BF-200 ALA or MAL with regards to pain and erythema. Interestingly, while better tolerated, HAL2% produced similar fluorescence and photobleaching to that of MAL and BF-200 ALA. The fact that all photosensitizers induced pain and erythema on non-photo-damaged healthy skin indicates that the effect of PDT is not completely specific to cancerous tissues.

A limitation of our trial was the small sample size and the lack of an accurate fluorescence imaging system. As our study was limited to healthy human skin, further research is needed into the safety and efficacy of these novel photosensitizers in cancerous skin.

Previously PDT with MAL20% was less painful and caused less erythema compared with ALA20% on healthy sun-exposed human skin (11). We showed that BF-200 ALA was as well tolerated as MAL. This

Table I. Treatment reactions

	BF-200	MAL	HAL 2%	HAL 0.2%
None	0	1	1	4
Mild	1	2	3	2
Intermediate	2	2	0	1
Severe	4	2	3	0

MAL: amino-5-laevulinate methyl ester; HAL: 5-aminolaevulinate hexylester

may be due to the lower concentration of 5-ALA in this nano-formulation. In UV-exposed mice, HAL2% induced similar epidermal fluorescence to that of MAL 20% (12). Our finding of strong fluorescence and photobleaching with only 2% HAL supports these findings and further indicates the better tolerability of HAL.

These pilot results show better tolerability of low-concentration HAL compared with BF-200 ALA and MAL on healthy human skin. However, further research is required to determine whether the treatment might be sufficiently effective without erythema and pain.

ACKNOWLEDGEMENTS

The authors would like to thank our healthy volunteers for participating in the study and MSc Reeta Neittaanmäki for statistical assistance. The study was supported by the Foundation for Clinical Chemistry Research.

Conflicts of interest. NNP has received travel grants from Biofrontera and Galderma and speaker honoraria from Biofrontera and Desitin Pharma. EN, IP, MG, and ES have no conflicts of interest.

REFERENCES

- Morton CA, Szeimies RM, Sidoroff A, Braathen LR. European guidelines for topical photodynamic therapy part 1: treatment delivery and current indications – actinic keratoses, Bowen's disease, basal cell carcinoma. J Eur Acad Dermatol Venereol 2013; 27: 536–544.
- Morton CA, Szeimies RM, Sidoroff A, Braathen LR. European guidelines for topical photodynamic therapy part 2: emerging indications field cancerization, photorejuvenation and inflammatory/infective dermatoses. J Eur Acad Dermatol Venereol 2013; 27: 672–679.
- 3. Ericson MB, Wennberg AM, Larkö O. Review of photodynamic therapy in actinic keratosis and basal cell carcinoma. Ther Clin Risk Manag 2008; 4: 1–9.
- 4. Wiegell SR, Haedersdal M, Philipsen PA, Eriksen P, Enk CD, Wulf HC. Continuous activation of PpIX by daylight is as effective as and less painful than conventional photodynamic therapy for actinic keratoses; a randomized, controlled, single-blinded study. Br J Dermatol 2008; 158: 740–746.
- 5. Wiegell SR, Petersen B, Wulf HC. Topical corticosteroid reduces inflammation without compromising efficacy of photodynamic therapy for actinic keratoses a randomised clinical trial. Br J Dermatol 2014; 171: 1487–1492.
- Petersen B, Wiegell SR, Wulf HC. Light protection of the skin after photodynamic therapy reduces inflammation: an unblinded randomized controlled study. Br J Dermatol 2014; 171: 175–178.
- 7. Peng Q, Berg K, Moan J, Kongshaug M, Nesland JM.

¹http://www.medicaljournals.se/acta/content/?doi=10.2340/00015555-2131 2 r=0.969 for MAL, p=0.007; r=0.847 for BF-200 ALA, p=ns; r=0.940 for HAL2%, p=0.018; r=0.939 for HAL 0.2%, p=0.022.

 $^{^{3}}$ r=0.850 for MAL, p=0.015; r=0.569 for BF-200 ALA, p=ns; r=0.906 for HAL2%, p=0.005; r=0.312 for HAL 0.2%, p=ns.

- 5-Aminolevulinic acid-based photodynamic therapy: principles and experimental research. Photochem Photobiol 1997; 65: 235–251.
- 8. Maisch T, Santarelli F, Schreml S, Babilas P, Szeimies RM. Fluorescence induction of protoporphyrin IX by a new 5-aminolevulinic acid nanoemulsion used for photodynamic therapy in a full-thickness ex vivo skin model. Exp Dermatol 2010; 19: 302–305.
- 9. Fullerton A, Fischer T, Lahti A, Wilhelm KP, Takiwaki H, Serup J: Guidelines for measurement skin colour and erythema A report from the Standardization Group of the European Society of Contact Dermatitis. Contact Dermatitis 1996; 35: 1–10.
- Tyrrell JS, Campbell SM, Curnow A. The relationship between protoporphyrin IX photobleaching during real-time dermatological methyl-aminolevulinate photodynamic therapy (MAL-PDT) and subsequent clinical outcome. Lasers Surg Med 2010; 42: 613–619.
- 11. Wiegell SR, Stender IM, Na R, Wulf HC. Pain associated with photodynamic therapy using 5-aminolevulinic acid or 5-aminolevulinic acid methylester on tape-stripped normal skin. Arch Dermatol 2003; 139: 1173–1177.
- 12. Togsverd-Bo K, Lerche CM, Philipsen PA, Poulsen T, Wulf HC, Haedersdal M. Porphyrin biodistribution in UV-exposed murine skin after methyl- and hexyl-aminolevulinate incubation. Exp Dermatol 2012; 21: 260–264.