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RESEARCH ARTICLE

Association of Forced Vital Capacity with the Developmental Gene *NCOR2*

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Abstract

Background

Forced Vital Capacity (FVC) is an important predictor of all-cause mortality in the absence of chronic respiratory conditions. Epidemiological evidence highlights the role of early life factors on adult FVC, pointing to environmental exposures and genes affecting lung development as risk factors for low FVC later in life. Although highly heritable, a small number of genes have been found associated with FVC, and we aimed at identifying further genetic variants by focusing on lung development genes.

Methods

Per-allele effects of 24,728 SNPs in 403 genes involved in lung development were tested in 7,749 adults from three studies (NFBC1966, ECRHS, EGEA). The most significant SNP for the top 25 genes was followed-up in 46,103 adults (CHARGE and SpiroMeta consortia) and 5,062 children (ALSPAC). Associations were considered replicated if the replication p-value survived Bonferroni correction ($p < 0.002$; $0.05/25$), with a nominal p-value considered as suggestive evidence. For SNPs with evidence of replication, effects on the expression levels of nearby genes in lung tissue were tested in 1,111 lung samples (Lung eQTL consortium), with further functional investigation performed using public epigenomic profiling data (ENCODE).

Results

NCOR2-rs12708369 showed strong replication in children ($p = 0.0002$), with replication unavailable in adults due to low imputation quality. This intronic variant is in a strong transcriptional enhancer element in lung fibroblasts, but its eQTL effects could not be tested due to low imputation quality in the eQTL dataset. *SERPINE2*-rs6754561 replicated at nominal level in both adults ($p = 0.036$) and children ($p = 0.045$), while *WNT16*-rs2707469 replicated at nominal level only in adults ($p = 0.026$). The eQTL analyses showed association of *WNT16*-rs2707469 with expression levels of the nearby gene *CPED1*. We found no statistically significant eQTL effects for *SERPINE2*-rs6754561.

Conclusions

We have identified a new gene, *NCOR2*, in the retinoic acid signalling pathway pointing to a role of vitamin A metabolism in the regulation of FVC. Our findings also support *SERPINE2*, a COPD gene with weak previous evidence of association with FVC, and suggest *WNT16* as a further promising candidate.

Introduction

Forced vital capacity (FVC), a spirometric measure routinely used in clinical practice to approximate vital capacity, is increasingly recognised as an important parameter beyond its diagnostic and prognostic role in restrictive lung diseases. Unlike the ratio of forced expiratory volume in 1 second (FEV_1) to FVC, an indicator of airway obstruction, FVC is a strong predictor of all-cause mortality in asymptomatic adults without chronic respiratory conditions[1]. Although the origins of a low FVC in the general population are poorly understood, there is a strong link to poverty[2], and in particular to low socio-economic status in early life[3]. Endemic vitamin A deficiency is associated with low FVC, and maternal supplementation with vitamin A before, during and after pregnancy, improves FVC in offspring[4]. Low FVC has also been associated with early exposure to particulate air pollution[5]. The deviation of an individual's FVC values (and lung function in general) from the population mean has been shown to remain stable over time, with future values being predicted by early measurements ("tracking")[6], which means that early life and genetic effects that manifest in childhood will influence the individual's whole FVC life trajectory. Taken together, this evidence highlights the role of early life factors on adult FVC, which points to environmental exposures and genes

affecting the development of the lung. Severe defects in lung development lead to neonatal death, but milder structural or functional defects could affect lung function and increase susceptibility to lung diseases that become clinically detectable during childhood or later life, including asthma and COPD[7]. This is supported by experimental work on *in-vitro* and animal models of lung function and disease[8].

Knowledge of the genetics of FVC is still limited. Biological candidates for FVC, mainly related to host defense, inflammatory pathway, pulmonary surfactant and oxidative stress, have been evaluated in candidate-gene association studies, but replication has been difficult. New candidates for FVC have been provided by genome-wide association (GWA) studies, the largest being a recent meta-analysis from the joint CHARGE and SpiroMeta consortia on 52,253 individuals, with replication of the top associations in 24,840 individuals[9]. It identified eight loci, of which six new (*EFEMP1*, *BMP6*, *MIR129-2-HSD17B12*, *PRDM11*, *WVVOX*, *KCNJ2*), and two previously associated with FEV₁ and FEV₁/FVC (*GSTCD* and *PTCH1*). The eight loci explain 1.8% of FVC variation, and yet FVC heritability (proportion of FVC variation attributable to genetic factors) is estimated around 40–60% by familial aggregation and twin studies[10, 11] and, more recently, genome-wide data[12].

Available GWA datasets represent an invaluable resource to test hypotheses about the role of genetic pathways involved in specific pathophysiological mechanisms. We hypothesised that focusing on genes lying in pathways related to lung development could help identify new candidates for FVC and further our understanding of the underlying biological mechanisms.

Materials and Methods

We evaluated the effect on FVC of 403 genes (24,728 SNPs) related to lung development in two stages. In Stage 1, all SNPs were tested for association with FVC in a meta-analysis of three European adult studies (ECRHS[13], NFBC1966[14], EGEA[15]). For replication in adults (CHARGE and SpiroMeta consortia)[9] and children (ALSPAC[16]) in Stage 2, we selected the best signal for the top 25 genes, defined as the SNP with the lowest meta-analysis p-value which satisfied the following criteria: minor allele frequency >0.05 and imputation quality (imputation R²) >0.7 in all three studies; low between-study heterogeneity defined as I² <30%, with I² representing the percentage of total variation in effect estimates across studies due to heterogeneity rather than chance.

The rationale for limiting our replication analysis to the best signal for the top 25 genes was to maximise the probability of successful replication in children, where the sample size was only 5,062. With this sample size, testing for replication of 25 SNPs gives a power of about 80% to detect a variant explaining 0.3% of FVC residual variance, at a Bonferroni corrected p-value threshold of 0.002 (0.05/25). This assuming that genetic effects in children may be slightly stronger than in adults, where the variance explained by the eight loci previously identified[9] was 1.8%, an average of 0.23% per SNP.

Selection of candidate genes and SNPs

Two experts in lung development, a basic scientist (C.H.D.) and a clinician scientist (M.H.), compiled a list of genes involved in lung development, first independently and then through agreement. The selection of genes was based on their knowledge of the topic, mainly using genetic evidence from animal models[8, 17, 18]. This initial list was extended to include additional genes suggested by: 1) pathways information obtained from KEGG[19]—relevant genes lying in the same pathways as those in the initial list; 2) information from published literature identified using HuGE Navigator[20]—genes considered as associated with lung development in previous genetic association studies. When in doubt about which genes to select from large

Table 1. Characteristics of studies in Stage 1. N = number of subjects included in the analyses.

Study	N	Country	Sex[% male]	Age (years)		Height (cm) [Mean (SD)]	FVC (ml) [Mean (SD)]
				Absolute Range	Mean (SD)		
NFBC1966	5,218	Finland	47.9%	31–31	31 (0)	171.2 (9.2)	4,718 (987)
ECRHS	1,662	Spain, United Kingdom, France, Germany, Sweden, Norway, Switzerland, Estonia	47.5%	19.7–48.1	34.0 (7.1)	170.5 (9.5)	4,552 (1,031)
EGEA	869	France	46.1%	18.0–76.5	38.5 (12.6)	168.6 (8.5)	4,239 (982)

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gene families, those with higher gene expression in foetal lung were chosen, with information retrieved from the Human U133A/GNF1H Gene Atlas database using BioGPS[21].

The final list included 403 genes (S1 Table). According to NCBI gene definition, we retrieved SNPs within 2 kb upstream and 500 bp downstream of each gene, using the R package NCBI2R (<http://cran.r-project.org/web/packages/NCBI2R>). We identified 24,728 SNPs for which imputed data (based on HapMap release 22) were available for all three studies in Stage 1 (S1 Table).

Study populations

Stage 1. Below and in Table 1 we briefly describe the three studies, with details on spirometry and genotyping methods summarised in S2 and S3 Tables.

The Northern Finland Birth Cohort 1966 (NFBC1966) is a birth-cohort study in the provinces of Oulu and Lapland that recruited pregnant women with an expected date of delivery in 1966. A total of 12,231 children were recruited and followed-up in adulthood[14], with 6,033 participating in the clinical follow-up at 31 years. Of these, 5,218 individuals with GWA and spirometry data were included in this study.

The European Community Respiratory Health Survey (ECRHS) is an international cohort study designed to identify risk factors for asthma[13] that started in 1992–1994, with follow-up performed twice in the following 20 years. Included in this study are 1,662 subjects from the first survey (ECRHS I, age 20–48) with GWA and spirometry data available, recruited from 16 centres that used random sampling frameworks.

The Epidemiological study on the Genetics and Environment of Asthma (EGEA), which combines a case-control and a family-based study of asthma, was conducted in 1991–1995 (EGEA1), with follow-up after 12 years (EGEA2, 2003–2007)[15]. The study included 388 nuclear families, ascertained by one or two asthmatic adult or paediatric probands, and 415 population-based controls, totalling 2,120 subjects. This analysis only includes 869 non-asthmatic adults, using spirometry data from EGEA1 for subjects ≥ 18 year old at baseline and EGEA2 for those < 18 in EGEA1.

Stage 2. The joint CHARGE (Cohorts for Heart and Aging Research in Genomic Epidemiology) and SpiroMeta consortia performed a GWA investigation of FVC in 52,253 individuals of European ancestry from 26 studies[9], which included ECRHS and NFBC1966. Included here are 46,103 individuals from 24 studies, after subtracting the contribution of ECRHS and NFBC1966. New effect estimates and standard errors were derived by taking a weighted difference between the original fixed-effect meta-analysis estimate and the pooled estimate of ECRHS and NFBC1966.

The Avon Longitudinal Study of Parents and their Children (ALSPAC) is a birth cohort study consisting initially of 14,541 women and their children recruited in the county of Avon, UK, in the early 1990s[16]. Included in this study are 5,062 white European children (50.3%

male) of 8–9 years of age with GWA and spirometry data. Their mean height was 132.6 cm (standard deviation, SD: 5.8) and mean FVC 1,931 ml (SD: 319).

Statistical analyses

Stage 1. Study-specific estimates for the three studies were obtained assuming an additive mode of inheritance. In ECRHS, linear regression analyses of the effects of the SNPs on FVC (in ml) were adjusted for age, age², height, sex, centre, and first four ancestry principal components to control for residual population stratification. In NFBC1966, all subjects were 31 year olds and linear regression analyses were only adjusted for height, sex and first two principal components. In the family-based EGEA, the regression analyses were performed using linear mixed models to account for family structure, adjusting for age, age², height, sex and first two principal components.

Inverse-variance weighted meta-analysis of the three studies using a fixed effect model was performed on a total of 7,749 individuals.

The association analyses for NFBC1966 were carried out using SNPTTEST[22], while the analyses for ECRHS and EGEA and the meta-analysis were performed using R, version 3.0.1 (www.R-project.org).

Stage 2. Individual cohorts within CHARGE and SpiroMeta performed GWA analyses for FVC (ml) using linear regression adjusted for age, age², height and sex (plus height² and weight for CHARGE), as well as centre and/or principal components if appropriate[9].

In ALSPAC, linear regression analyses on FVC (ml) were performed adjusting for age, age², height and sex. Principal components were not included since no evidence of population stratification was found in the study.

Replication of a SNP was defined based on evidence from Stage 2 only, rather than on combined evidence from Stage 1 and Stage 2, since this protects against the winner's curse, an upwards bias typical of the screening stage[23]. We considered a SNP replicated if the effect estimate was in the same direction as in Stage 1 and the one-side p-value survived Bonferroni correction for multiple testing ($p < 0.002$) in either adults or children. We considered replication evidence as suggestive if the p-value was significant only at nominal level.

Lung eQTL data

For SNPs with evidence of replication, we investigated their effects on the expression of nearby genes (genes within 100 kb up and downstream from the SNP) in lung samples from the Lung QTL consortium. This includes data on 1,111 individuals undergoing lung surgery, recruited at Laval University (n = 409), University of British Columbia (n = 339) and University of Groningen (n = 363)[24].

Gene expression and genotyping profiles were obtained using a custom Affymetrix array (GEO platform GPL10379) and the Illumina Human1M-Duo BeadChip array, respectively. Expression values were extracted using the Robust Multichip Average method[25] implemented in the Affymetrix Power Tools software. Expression values were analysed with a robust regression model adjusted for age, sex and smoking status, using the R statistical package MASS (*rlm* function).

Genetic associations were performed in PLINK 1.9. A fixed-effect meta-analysis was used to pool the results across the three sites.

Results

Stage 1 study-specific and meta-analysis results are reported in Table 2 for the best SNP of the top 25 genes, and in S1 Table for all 24,728 SNPs. Replication could only be performed for 24 SNPs, since no data were available for *EYAI* rs12549242 or any proxy (defined as a SNP with

Table 2. Results for the best SNP of the top 25 genes in Stage 1: NFBC 1966, ECRHS, EGEE, and meta-analysis. Chr: chromosome; EA: effect allele; EAF: effect allele frequency, calculated as weighted average across the three studies; β (standard error, SE): estimate of the per-allele effect on FVC (ml); I^2 : magnitude of the between-study heterogeneity of effect estimates

SNP	Gene	Chr	Position	EA	EAF	NFBC1966(N = 5,218)			ECRHS(N = 1,662)			EGEE(N = 869)			Meta-analysis			
						β	SE	P	β	SE	P	β	SE	P	β	SE	P	I^2 (%)
rs2820472	WLS	1	68,694,307	C	0.70	31.0	11.3	0.0061	30.3	21.7	0.1621	-17.5	32.0	0.5851	26.5	9.6	0.0055	4
rs832169	PKP1	1	201,256,771	A	0.17	29.2	15.8	0.0646	50.0	22.4	0.0257	41.1	31.9	0.1984	36.9	12.0	0.0021	0
rs7527525	ACTN2	1	236,902,560	C	0.33	20.0	11.7	0.0875	33.6	20.2	0.0960	63.7	28.0	0.0238	28.1	9.5	0.0032	8
rs3905417	CTNNA2	2	80,181,443	A	0.23	29.9	12.2	0.0144	30.4	24.4	0.2127	45.7	34.0	0.1796	31.4	10.4	0.0025	0
rs6754561	SERPINE2	2	224,839,696	C	0.30	-29.6	12.2	0.0151	-23.0	19.3	0.2350	-11.4	26.8	0.6707	-25.6	9.6	0.0077	0
rs11926758	RARB	3	25,552,252	G	0.94	52.5	22.9	0.0219	51.3	37.7	0.1734	48.8	48.1	0.3119	51.7	18.1	0.0044	0
rs11716871	TP63	3	189,582,501	A	0.92	-55.4	19.1	0.0037	-32.5	34.0	0.3404	-36.1	47.2	0.4444	-48.4	15.7	0.0021	0
rs4712047	SIRT5	6	13,590,185	A	0.66	34.0	11.2	0.0024	9.9	22.7	0.6622	27.9	32.3	0.3882	29.2	9.6	0.0023	0
rs2722322	SFRP4	7	37,948,714	A	0.15	51.7	15.4	0.0008	36.8	24.3	0.1298	16.3	35.3	0.6437	43.7	12.2	0.0003	0
rs17172023	GLI3	7	42,245,499	C	0.78	36.4	13.8	0.0082	25.2	24.4	0.3034	10.2	33.8	0.7644	31.1	11.3	0.0060	0
rs1049337	CAV1	7	116,200,587	C	0.70	33.6	11.6	0.0038	28.1	19.8	0.1562	-10.3	29.3	0.7254	27.8	9.5	0.0034	0
rs2707469	WNT16	7	120,976,886	A	0.83	34.2	14.3	0.0168	23.6	25.9	0.3611	34.5	39.1	0.3787	32.0	11.9	0.0073	0
rs12549242	EYA1	8	72,216,430	C	0.14	-38.6	16.1	0.0167	-53.8	24.3	0.0267	-72.2	43.4	0.0972	-45.8	12.8	0.0004	0
rs2812427	DLG5	10	79,553,236	A	0.67	33.3	11.2	0.0029	14.8	19.8	0.4548	63.1	28.5	0.0274	32.4	9.2	0.0004	0
rs1994450	PDGFD	11	103,797,349	A	0.13	-41.9	18.0	0.0201	-64.3	29.3	0.0283	2.1	38.9	0.9573	-41.3	14.3	0.0038	0
rs12708369	NCOR2	12	124,875,577	C	0.56	25.1	11.5	0.0291	46.0	20.7	0.0263	-9.1	30.1	0.7626	26.1	9.5	0.0062	13
rs11865499	KAT8	16	31,132,250	A	0.69	33.7	11.3	0.0029	20.4	19.9	0.3061	6.2	28.9	0.8304	27.9	9.3	0.0027	0
rs1880756	CRHR1	17	43,826,666	C	0.58	-26.5	10.6	0.0122	-28.6	19.4	0.1418	-5.4	27.8	0.8472	-24.8	8.8	0.0049	0
rs948589	SMAD4	18	48,586,184	A	0.91	-47.2	19.0	0.0131	-56.4	34.4	0.1073	-78.5	50.7	0.1227	-52.2	15.8	0.0010	0
rs2425024	MMP24	20	33,844,938	A	0.66	25.0	11.3	0.0274	23.9	19.4	0.2184	55.7	26.9	0.0390	28.4	9.2	0.0020	0
rs6061580	CDH4	20	60,058,986	C	0.92	-60.4	22.3	0.0067	-41.5	37.3	0.2657	-7.6	47.0	0.8717	-48.7	17.7	0.0060	0
rs2051179	RUNX1	21	36,326,553	A	0.45	-32.0	10.9	0.0032	-15.7	18.8	0.4038	-16.1	25.9	0.5336	-26.6	8.8	0.0026	0
rs730265	CLDN14	21	37,871,886	A	0.15	-25.8	15.6	0.0973	-41.8	24.2	0.0837	-55.2	33.7	0.1020	-33.7	12.2	0.0057	0
rs2871029	CLDN5	22	19,513,930	A	0.14	31.4	15.1	0.0375	66.5	27.6	0.0161	-10.3	40.0	0.7969	34.6	12.6	0.0060	24
rs5749524	TIMP3	22	33,224,285	C	0.89	49.7	17.0	0.0035	22.4	29.3	0.4452	58.5	38.1	0.1259	44.9	13.7	0.0011	0

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linkage disequilibrium, LD, $R^2 > 0.8$) in CHARGE and SpiroMeta and ALSPAC. In Stage 2, one gene showed strong replication in children, *NCOR2*, with replication unavailable for adults due to low imputation quality; other two genes showed suggestive evidence of replication, one in both adults and children, *SERPINE2*, and the other in adults but not in children, *WNT16* (Table 3). The regional association plots for their lead SNP are presented in S1 Fig.

NCOR2-rs12708369 replicated in ALSPAC children with an effect of 26.9 ml/allele (95% confidence interval: 12.0 to 41.8) and a p-value well below Bonferroni correction ($p = 0.0002$). The estimate was very similar to that of Stage 1 (26.1; 7.5 to 44.7), suggesting a relatively stronger effect in children given their lower FVC, although the confidence intervals are wide and conclusions as to a difference in effect sizes cannot be deduced. In line with this, the proportion of FVC residual variance explained by this SNP was much higher in children than in adults from Stage 1, 0.65% vs. 0.11%. Replication of *NCOR2*-rs12708369 could not be performed in adults because of low imputation quality (imputation $R^2 = 0.4$) and no proxy available. Using publicly available epigenomic profiling data (ChIP-seq) from ENCODE[26] via the UCSC Genome Browser (<http://genome.cse.ucsc.edu>), we found that the intronic variant *NCOR2*-rs12708369 is in a region with regulatory function in lung tissue. The SNP is located within a DNase I hypersensitivity site, in a strong enhancer element with histone mark H3K27ac indicating active chromatin in lung fibroblasts. Unfortunately neither *NCOR2*-rs12708369 nor any proxy could be tested in the lung eQTL analysis due to failed imputation quality control.

Table 3. Replication findings for the best SNP of the top 25 genes. Chr: chromosome; EA: effect allele; EAF: effect allele frequency; β (standard error, SE): per-allele effect on FVC (ml); Repl P: one-side replication p-value, calculated and reported only for estimates in the same direction as the original ones; I^2 : between-study heterogeneity; Imp R^2 = imputation quality R^2 (for CHARGE and SpiroMeta: average imputation R^2 across studies)

SNP	Gene	Chr	EA	EAF	STAGE 1meta-analysis(N = 7,749)			STAGE 2								
								CHARGE and SpiroMeta meta-analysis(N = 46,103—Adults)					ALSPAC(N = 5,062—Children)			
					β	SE	P	β	SE	Repl P	I ² (%)	Imp R ²	β	SE	Repl P	Imp R ²
rs2820472	WLS	1	C	0.70	26.5	9.6	0.0055	0.7	4.7	0.444	35	0.92	1.6	8.3	0.423	0.97
rs832169	PKP1	1	A	0.17	36.9	12.0	0.0021	-7.0	4.9	/	23	0.85	-2.1	8.3	/	0.94
rs7527525	ACTN2	1	C	0.33	28.1	9.5	0.0032	-4.2	4.6	/	24	0.71	11.2	7.2	0.059	0.90
rs3905417	CTNNA2	2	A	0.23	31.4	10.4	0.0025	2.5	5.2	0.312	0	0.95	13.2	9.0	0.071	0.99
rs6754561	SERPINE2	2	C	0.30	-25.6	9.6	0.0077	-7.1	3.9	0.036*	0	0.96	-12.0	7.1	0.045*	1.00
rs11926758	RARB	3	G	0.94	51.7	18.1	0.0044	-4.1	7.4	/	26	0.98	3.1	12.2	0.401	0.99
rs11716871	TP63	3	A	0.92	-48.4	15.7	0.0021	17.8	7.5	/	0	0.86	-14.4	12.5	0.125	0.98
rs4712047	SIRT5	6	A	0.66	29.2	9.6	0.0023	0.6	4.7	0.447	18	0.72	-4.7	8.5	/	0.70
rs2722322	SFRP4	7	A	0.15	43.7	12.2	0.0003	-1.7	5.1	/	18	0.94	12.0	8.8	0.088	1.00
rs17172023	GLI3	7	C	0.78	31.1	11.3	0.0060	-9.6	5.3	/	27	0.84	8.4	10.0	0.202	0.75
rs1049337	CAV1	7	C	0.70	27.8	9.5	0.0034	-4.5	5.1	/	35	0.69	0.3	7.4	0.484	1.00
rs2707469	WNT16	7	A	0.83	32.0	11.9	0.0073	10.0	5.2	0.026*	6	0.92	11.8	9.4	0.105	0.90
rs2812427	DLG5	10	A	0.67	32.4	9.2	0.0004	4.5	4.1	0.138	0	0.95	2.1	7.1	0.382	1.00
rs1994450	PDGFD	11	A	0.13	-41.3	14.3	0.0038	-1.7	5.5	0.380	0	0.76	-10.7	9.6	0.132	0.79
rs12708369	NCOR2	12	C	0.56	26.1	9.5	0.0062	NA ¹	NA ¹	NA ¹	38	0.38	26.9	7.6	0.0002**	0.78
rs11865499	KAT8	16	A	0.69	27.9	9.3	0.0027	4.2	4.6	0.181	30	0.84	10.6	7.5	0.078	1.00
rs1880756	CRHR1	17	C	0.58	-24.8	8.8	0.0049	-5.0	4.0	0.108	15	0.96	2.9	7.0	/	1.00
rs948589	SMAD4	18	A	0.91	-52.2	15.8	0.0010	8.8	6.7	/	0	0.96	-14.7	12.2	0.114	1.00
rs2425024	MMP24	20	A	0.66	28.4	9.2	0.0020	3.3	4.0	0.205	0	0.96	-11.3	7.1	/	1.00
rs6061580	CDH4	20	C	0.92	-48.7	17.7	0.0060	9.0	8.6	/	3	0.73	2.6	13.9	/	0.92
rs2051179	RUNX1	21	A	0.45	-26.6	8.8	0.0026	-3.8	3.8	0.159	26	0.94	-5.9	6.7	0.188	0.97
rs730265	CLDN14	21	A	0.15	-33.7	12.2	0.0057	-3.0	7.2	0.338	20	0.50	8.0	8.0	/	0.99
rs2871029	CLDN5	22	A	0.14	34.6	12.6	0.0060	-0.7	5.8	/	47	0.90	6.0	9.7	0.269	1.00
rs5749524	TIMP3	22	C	0.89	44.9	13.7	0.0011	2.0	6.0	0.371	0	0.94	1.4	10.4	0.448	1.00

* Nominal significance ($p < 0.05$)

** Significance after Bonferroni correction ($p < 0.002$)

¹ Results not available: the SNP had a very low average imputation R^2 (0.38) and no proxies ($LD R^2 > 0.80$) were available

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SERPINE2-rs6754561, a variant located 133 bp downstream from the gene, replicated at nominal level in adults from the CHARGE and SpiroMeta consortia (-7.1 ml/allele; $p = 0.036$), where there was no heterogeneity across the 24 studies ($I^2 = 0\%$), and ALSPAC children (-12.0 ml/allele; $p = 0.045$). The proportion of FVC residual variance explained was only 0.01% in adults, but 0.11% in children (0.09% in adults from Stage 1). *SERPINE2*-rs6754561 did not show association with the expression of *SERPINE2* or any nearby genes in the lung eQTL dataset.

The intronic variant *WNT16*-rs2707469 replicated at nominal level in adults (10.0 ml/allele; $p = 0.026$; $I^2 = 6\%$), but not in children (11.8 ml/allele; $p = 0.105$). The proportion of FVC residual variance explained was only 0.01% in adults from the CHARGE and SpiroMeta consortia (0.10% in Stage 1). This variant is in a conserved region and is located in a DNase I hypersensitivity site in lung fibroblasts. *WNT16*-rs2707469 was not associated with *WNT16* expression but showed suggestive evidence of an effect on a nearby gene, *CPEDI*, with the FVC-lowering

allele G associated with higher *CPED1* mRNA expression levels ($p = 0.087$; $I^2 = 0\%$; [S2 Fig](#)). We investigated this further and found that the effect on *CPED1* expression was stronger ($p = 0.004$; $I^2 = 0\%$) for a SNP in high LD with *WNT16*-rs2707469 ($R^2 = 0.94$), rs2536166 ([S2 Fig](#)).

Discussion

By testing the association of FVC with genes related to lung development, we have identified a new gene, *NCOR2*, in the retinoic acid signalling pathway pointing to a role of vitamin A metabolism in the regulation of FVC. Our study also provides support for *SERPINE2*, a gene which has previously shown weak evidence of association with FVC, and suggests *WNT16* as a promising candidate requiring further investigation.

NCOR2 (nuclear receptor corepressor 2), also known as *SMRT* (silencing mediator of retinoid and thyroid hormone), is a potent regulator of retinoid and thyroid hormone signalling. Nuclear receptors are ligand-activated transcription factors that regulate many developmental and physiological processes. Retinoic acid is the biologically active metabolite of vitamin A (retinol) which has a well described role in organogenesis and epithelial homeostasis directing growth, patterning and differentiation of many organs including the lung[27]. *NCOR2* is a transcriptional “platform” protein that acts as a repressive co-regulatory factor for multiple transcription factor pathways. Publicly available data retrieved from BioGPS[21] (Human U133A/GNF1H Gene Atlas database) show that the expression of *NCOR2* in the adult lung is very high and that the gene is also expressed in foetal lung. In this study we found an association of *NCOR2* (rs12708369) with FVC in adults, which strongly replicated in children. Replication in adults from the CHARGE and SpiroMeta consortia could not be performed due to low imputation quality and no data on proxies available either. The *NCOR2*-rs12708369 intro- nic variant is in a strong transcriptional enhancer element in lung fibroblasts and may therefore affect gene expression levels[28], although we were not able to test this due to the same problem of low imputation quality in the Lung eQTL dataset. The replication of *NCOR2* in children and the known central developmental roles of retinoic acid and thyroid hormone signalling during alveologenesis[29] suggest that this gene may influence lung growth and ultimately FVC. Although retinoic acid has also been postulated to have a role in ongoing alveolar maintenance and regeneration[30], in our study the *NCOR2*-rs12708369 effect in adults could be estimated only in Stage 1 mostly based on 31-year olds, so potential effects on FVC decline would not have been detected. Interestingly, another related gene, the *RARB* encoding the retinoic acid receptor beta, was selected in Stage 1, although it could not be replicated possibly due to the low minor allele frequency of its selected SNP (rs11926758; MAF = 0.06). This gene has been previously associated with measures of airway obstruction in adults and children (FEV₁/FVC)[31, 32], and in infants (V'maxFRC)[33]. Overall our findings point to a role of vitamin A/thyroid metabolism in the regulation of FVC, and suggest the importance of further research investigating genes in related pathways as well as gene-environment interactions with vitamin A intake.

SERPINE2 is a member of a gene family encoding serpins, highly conserved proteins that help maintain tissue integrity by controlling the activity of proteases in diverse biological processes, in particular by inhibiting serine proteases such as trypsin. *SERPINE2* has a known link to airway obstruction, with strong evidence of association with COPD[34] and some evidence of association with childhood asthma[35]. Our findings support an association with a marker of lung restriction too, FVC, in both adults and children, in line with previous findings of an association with FVC in children that could not be replicated[36]. *SERPINE2*-rs6754561 showed no effect on the expression of *SERPINE2* or nearby genes in the lung. However, although the Lung eQTL dataset represents the largest eQTL mapping study of human lung

samples currently available, weak to moderate effects on gene expression may not have been detected due to insufficient statistical power. Cellular heterogeneity in lung tissue may also impair the detection of cell type-specific eQTL[37].

We also found suggestive evidence of an association of *WNT16* with FVC in adults. *WNT16* belongs to a family of genes encoding 19 Wnt ligands, secreted signalling proteins involved in many developmental processes. Although Wnts are critical for normal lung development[18, 38], Wnt16 has not been previously studied in relation to lung function and disease. In addition to lung development, evidence from mouse models suggests that Wnt16 plays a role in tissue repair[39] and in the response to cellular damage[40]. The *WNT16*-rs2707469 intronic variant is in a conserved region with regulatory function in lung fibroblasts. This variant showed no eQTL effect on *WNT16* in the lung, but an effect on a nearby gene, *CPED1* (cadherin-like and PC-esterase domain containing 1). *CPED1* has both a cadherin-like domain, thought to have a carbohydrate binding function, and a PC-esterase domain, predicted to modify cell surface biomolecules like glycoproteins. It is possible that Wnt16, which is a glycoprotein containing carbohydrates, could bind to, and/or be modified by, *CPED1*.

By focusing on genetic pathways related to lung development, which represent highly plausible candidates for low FVC, our study identifies a novel gene and proposes two further promising candidates which had not been identified in the previous GWA meta-analysis[9]. This shows how a comprehensive hypothesis-driven approach can complement hypothesis-free GWA analyses in identifying variants which failed to reach the strict significance level needed to protect against false positives in genome-wide investigations (typically 5×10^{-8}). However, we did miss the association of one of the genes we tested, *PTCH1*, a gene which has shown association with FVC in the previous GWA meta-analysis[9] and had been identified before as associated with FEV₁/FVC[32, 41]. The three SNPs previously identified in *PTCH1* had non-significant p-values in our Stage 1 analysis, most likely due to their relatively low minor allele frequency (MAF between 0.08 and 0.10), which made our analysis underpowered to detect them.

In conclusion, this study identifies *NCOR2* as a new gene for FVC, indicating the importance of further research into the role of vitamin A intake/supplementation and its interactions with related genes in the regulation of FVC. Our findings also suggest other biological pathways as promising candidates for future investigation. We might expect genes involved in lung development to show stronger effects in childhood, and the relatively large replication estimate of the effect of *NCOR2*-rs12708369 in children seems to support this. We speculate that future investigation of genes involved in lung development in larger samples of children and young adults could identify further genetic variants associated with FVC through their effect on lung growth and maximum level attained.

Supporting Information

S1 Fig. Regional association plots for *NCOR2* rs12708369, *SERPINE2* rs6754561 and *WNT16* rs2707469.

(DOC)

S2 Fig. Forest plots for the meta-analyses of lung gene expression levels of *CPED1* associated with *WNT16* variants.

(DOC)

S1 Table. Stage 1 study-specific and meta-analysis results for all the 24,728 SNPs in the 403 genes.

(XLSX)

S2 Table. Spirometry methods for studies in Stage 1.
(DOC)

S3 Table. Genotyping and imputation methods for studies in Stage 1.
(DOC)

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References

1. Burney PG, Hooper R. Forced vital capacity, airway obstruction and survival in a general population sample from the USA. *Thorax*. 2011; 66(1):49–54. Epub 2010/10/29. doi: [10.1136/thx.2010.147041](https://doi.org/10.1136/thx.2010.147041) PMID: [20980245](https://pubmed.ncbi.nlm.nih.gov/20980245/).
2. Hegewald MJ, Crapo RO. Socioeconomic status and lung function. *Chest*. 2007; 132(5):1608–14. Epub 2007/11/14. doi: [10.1378/chest.07-1405](https://doi.org/10.1378/chest.07-1405) PMID: [17998360](https://pubmed.ncbi.nlm.nih.gov/17998360/).
3. Bartley M, Kelly Y, Sacker A. Early life financial adversity and respiratory function in midlife: a prospective birth cohort study. *American journal of epidemiology*. 2012; 175(1):33–42. Epub 2011/12/06. doi: [10.1093/aje/kwr284](https://doi.org/10.1093/aje/kwr284) PMID: [22138040](https://pubmed.ncbi.nlm.nih.gov/22138040/).
4. Checkley W, West KP Jr, Wise RA, Baldwin MR, Wu L, LeClerq SC, et al. Maternal vitamin A supplementation and lung function in offspring. *The New England journal of medicine*. 2010; 362(19):1784–94. Epub 2010/05/14. doi: [10.1056/NEJMoa0907441](https://doi.org/10.1056/NEJMoa0907441) PMID: [20463338](https://pubmed.ncbi.nlm.nih.gov/20463338/).

5. Kulkarni N, Pierse N, Rushton L, Grigg J. Carbon in airway macrophages and lung function in children. *The New England journal of medicine*. 2006; 355(1):21–30. Epub 2006/07/11. doi: [10.1056/NEJMoa052972](https://doi.org/10.1056/NEJMoa052972) PMID: [16822993](https://pubmed.ncbi.nlm.nih.gov/16822993/).
6. Twisk JW, Staal BJ, Brinkman MN, Kemper HC, van Mechelen W. Tracking of lung function parameters and the longitudinal relationship with lifestyle. *The European respiratory journal*. 1998; 12(3):627–34. Epub 1998/10/08. PMID: [9762791](https://pubmed.ncbi.nlm.nih.gov/9762791/).
7. Shi W, Bellusci S, Warburton D. Lung development and adult lung diseases. *Chest*. 2007; 132(2):651–6. Epub 2007/08/19. doi: [10.1378/chest.06-2663](https://doi.org/10.1378/chest.06-2663) PMID: [17699136](https://pubmed.ncbi.nlm.nih.gov/17699136/).
8. Krauss-Etschmann S, Bush A, Bellusci S, Brusselle GG, Dahlen SE, Dehmel S, et al. Of flies, mice and men: a systematic approach to understanding the early life origins of chronic lung disease. *Thorax*. 2013; 68(4):380–4. Epub 2012/07/12. doi: [10.1136/thoraxjnl-2012-201902](https://doi.org/10.1136/thoraxjnl-2012-201902) PMID: [22781122](https://pubmed.ncbi.nlm.nih.gov/22781122/).
9. Loth DW, Soler Artigas M, Gharib SA, Wain LV, Franceschini N, Koch B, et al. Genome-wide association analysis identifies six new loci associated with forced vital capacity. *Nature genetics*. 2014; 46(7):669–77. doi: [10.1038/ng.3011](https://doi.org/10.1038/ng.3011) PMID: [24929828](https://pubmed.ncbi.nlm.nih.gov/24929828/); PubMed Central PMCID: PMC4140093.
10. Palmer LJ, Knuiman MW, Divitini ML, Burton PR, James AL, Bartholomew HC, et al. Familial aggregation and heritability of adult lung function: results from the Busselton Health Study. *The European respiratory journal*. 2001; 17(4):696–702. Epub 2001/06/13. PMID: [11401066](https://pubmed.ncbi.nlm.nih.gov/11401066/).
11. Hallberg J, Iliadou A, Anderson M, de Verdier MG, Nihlen U, Dahlback M, et al. Genetic and environmental influence on lung function impairment in Swedish twins. *Respiratory research*. 2010; 11:92. Epub 2010/07/08. doi: [10.1186/1465-9921-11-92](https://doi.org/10.1186/1465-9921-11-92) PMID: [20604964](https://pubmed.ncbi.nlm.nih.gov/20604964/); PubMed Central PMCID: PMCPmc2914039.
12. Klimentidis YC, Vazquez AI, de Los Campos G, Allison DB, Dransfield MT, Thannickal VJ. Heritability of pulmonary function estimated from pedigree and whole-genome markers. *Frontiers in genetics*. 2013; 4:174. Epub 2013/09/24. doi: [10.3389/fgene.2013.00174](https://doi.org/10.3389/fgene.2013.00174) PMID: [24058366](https://pubmed.ncbi.nlm.nih.gov/24058366/); PubMed Central PMCID: PMCPmc3766834.
13. Burney PG, Luczynska C, Chinn S, Jarvis D. The European Community Respiratory Health Survey. *The European respiratory journal*. 1994; 7(5):954–60. Epub 1994/05/01. PMID: [8050554](https://pubmed.ncbi.nlm.nih.gov/8050554/).
14. Rantakallio P. The longitudinal study of the northern Finland birth cohort of 1966. *Paediatric and perinatal epidemiology*. 1988; 2(1):59–88. Epub 1988/01/01. PMID: [2976931](https://pubmed.ncbi.nlm.nih.gov/2976931/).
15. Kauffmann F, Dizier MH, Annesi-Maesano I, Bousquet J, Charpin D, Demeais F, et al. EGEA (Epidemiological study on the Genetics and Environment of Asthma, bronchial hyperresponsiveness and atopy)—descriptive characteristics. *Clinical and experimental allergy: journal of the British Society for Allergy and Clinical Immunology*. 1999; 29 Suppl 4:17–21. Epub 2000/01/21. PMID: [10641560](https://pubmed.ncbi.nlm.nih.gov/10641560/).
16. Boyd A, Golding J, Macleod J, Lawlor DA, Fraser A, Henderson J, et al. Cohort Profile: the 'children of the 90s'—the index offspring of the Avon Longitudinal Study of Parents and Children. *International journal of epidemiology*. 2013; 42(1):111–27. doi: [10.1093/ije/dys064](https://doi.org/10.1093/ije/dys064) PMID: [22507743](https://pubmed.ncbi.nlm.nih.gov/22507743/); PubMed Central PMCID: PMC3600618.
17. Kho AT, Bhattacharya S, Mecham BH, Hong J, Kohane IS, Mariani TJ. Expression profiles of the mouse lung identify a molecular signature of time-to-birth. *American journal of respiratory cell and molecular biology*. 2009; 40(1):47–57. Epub 2008/07/31. doi: [10.1165/rcmb.2008-0048OC](https://doi.org/10.1165/rcmb.2008-0048OC) PMID: [18664640](https://pubmed.ncbi.nlm.nih.gov/18664640/); PubMed Central PMCID: PMCPmc2606946.
18. Morrissey EE, Hogan BL. Preparing for the first breath: genetic and cellular mechanisms in lung development. *Developmental cell*. 2010; 18(1):8–23. Epub 2010/02/16. doi: [10.1016/j.devcel.2009.12.010](https://doi.org/10.1016/j.devcel.2009.12.010) PMID: [20152174](https://pubmed.ncbi.nlm.nih.gov/20152174/); PubMed Central PMCID: PMCPmc3736813.
19. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M. KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic acids research*. 2012; 40(Database issue):D109–14. doi: [10.1093/nar/gkr988](https://doi.org/10.1093/nar/gkr988) PMID: [22080510](https://pubmed.ncbi.nlm.nih.gov/22080510/); PubMed Central PMCID: PMC3245020.
20. Yu W, Gwinn M, Clyne M, Yesupriya A, Khoury MJ. A navigator for human genome epidemiology. *Nature genetics*. 2008; 40(2):124–5. Epub 2008/01/30. doi: [10.1038/ng0208-124](https://doi.org/10.1038/ng0208-124) PMID: [18227866](https://pubmed.ncbi.nlm.nih.gov/18227866/).
21. Wu C, Orozco C, Boyer J, Leglise M, Goodale J, Batalov S, et al. BioGPS: an extensible and customizable portal for querying and organizing gene annotation resources. *Genome biology*. 2009; 10(11):R130. Epub 2009/11/19. doi: [10.1186/gb-2009-10-11-r130](https://doi.org/10.1186/gb-2009-10-11-r130) PMID: [19919682](https://pubmed.ncbi.nlm.nih.gov/19919682/); PubMed Central PMCID: PMCPmc3091323.
22. Marchini J, Howie B, Myers S, McVean G, Donnelly P. A new multipoint method for genome-wide association studies by imputation of genotypes. *Nature genetics*. 2007; 39(7):906–13. Epub 2007/06/19. doi: [10.1038/ng2088](https://doi.org/10.1038/ng2088) PMID: [17572673](https://pubmed.ncbi.nlm.nih.gov/17572673/).
23. Thomas DC, Casey G, Conti DV, Haile RW, Lewinger JP, Stram DO. Methodological Issues in Multi-stage Genome-wide Association Studies. *Statistical science: a review journal of the Institute of Mathematical Statistics*. 2009; 24(4):414–29. Epub 2010/07/08. PMID: [20607129](https://pubmed.ncbi.nlm.nih.gov/20607129/); PubMed Central PMCID: PMCPmc2895324.

24. Hao K, Bosse Y, Nickle DC, Pare PD, Postma DS, Laviolette M, et al. Lung eQTLs to help reveal the molecular underpinnings of asthma. *PLoS genetics*. 2012; 8(11):e1003029. Epub 2012/12/05. doi: [10.1371/journal.pgen.1003029](https://doi.org/10.1371/journal.pgen.1003029) PMID: [23209423](https://pubmed.ncbi.nlm.nih.gov/23209423/); PubMed Central PMCID: PMCPmc3510026.
25. Irizarry RA, Hobbs B, Collin F, Beazer-Barclay YD, Antonellis KJ, Scherf U, et al. Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics (Oxford, England)*. 2003; 4(2):249–64. Epub 2003/08/20. doi: [10.1093/biostatistics/4.2.249](https://doi.org/10.1093/biostatistics/4.2.249) PMID: [12925520](https://pubmed.ncbi.nlm.nih.gov/12925520/).
26. Birney E, Stamatoyannopoulos JA, Dutta A, Guigo R, Gingeras TR, Margulies EH, et al. Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature*. 2007; 447(7146):799–816. Epub 2007/06/16. doi: [10.1038/nature05874](https://doi.org/10.1038/nature05874) PMID: [17571346](https://pubmed.ncbi.nlm.nih.gov/17571346/); PubMed Central PMCID: PMCPmc2212820.
27. Cunningham TJ, Duester G. Mechanisms of retinoic acid signalling and its roles in organ and limb development. *Nature reviews Molecular cell biology*. 2015; 16(2):110–23. Epub 2015/01/07. doi: [10.1038/nrm3932](https://doi.org/10.1038/nrm3932) PMID: [25560970](https://pubmed.ncbi.nlm.nih.gov/25560970/).
28. Corradin O, Scacheri PC. Enhancer variants: evaluating functions in common disease. *Genome medicine*. 2014; 6(10):85. Epub 2014/12/05. doi: [10.1186/s13073-014-0085-3](https://doi.org/10.1186/s13073-014-0085-3) PMID: [25473424](https://pubmed.ncbi.nlm.nih.gov/25473424/); PubMed Central PMCID: PMCPmc4254432.
29. Massaro D, Massaro GD. Lung development, lung function, and retinoids. *The New England journal of medicine*. 2010; 362(19):1829–31. Epub 2010/05/14. doi: [10.1056/NEJMe1002366](https://doi.org/10.1056/NEJMe1002366) PMID: [20463343](https://pubmed.ncbi.nlm.nih.gov/20463343/).
30. Hind M, Giltthorpe A, Stinchcombe S, Maden M. Retinoid induction of alveolar regeneration: from mice to man? *Thorax*. 2009; 64(5):451–7. Epub 2009/04/30. doi: [10.1136/thx.2008.105437](https://doi.org/10.1136/thx.2008.105437) PMID: [19401491](https://pubmed.ncbi.nlm.nih.gov/19401491/).
31. Kreiner-Moller E, Bisgaard H, Bonnelykke K. Prenatal and postnatal genetic influence on lung function development. *The Journal of allergy and clinical immunology*. 2014; 134(5):1036–42.e15. Epub 2014/05/27. doi: [10.1016/j.jaci.2014.04.003](https://doi.org/10.1016/j.jaci.2014.04.003) PMID: [24857373](https://pubmed.ncbi.nlm.nih.gov/24857373/).
32. Soler Artigas M, Loth DW, Wain LV, Gharib SA, Obeidat M, Tang W, et al. Genome-wide association and large-scale follow up identifies 16 new loci influencing lung function. *Nature genetics*. 2011; 43(11):1082–90. doi: [10.1038/ng.941](https://doi.org/10.1038/ng.941) PMID: [21946350](https://pubmed.ncbi.nlm.nih.gov/21946350/); PubMed Central PMCID: PMCPmc3267376.
33. Collins SA, Lucas JS, Inskip HM, Godfrey KM, Roberts G, Holloway JW. HHIP, HDAC4, NCR3 and RARB polymorphisms affect fetal, childhood and adult lung function. *The European respiratory journal*. 2013; 41(3):756–7. Epub 2013/03/05. doi: [10.1183/09031936.00171712](https://doi.org/10.1183/09031936.00171712) PMID: [23456936](https://pubmed.ncbi.nlm.nih.gov/23456936/); PubMed Central PMCID: PMCPmc3691629.
34. Demeo DL, Mariani TJ, Lange C, Srisuma S, Litonjua AA, Celedon JC, et al. The SERPINE2 gene is associated with chronic obstructive pulmonary disease. *American journal of human genetics*. 2006; 78(2):253–64. Epub 2005/12/17. doi: [10.1086/499828](https://doi.org/10.1086/499828) PMID: [16358219](https://pubmed.ncbi.nlm.nih.gov/16358219/); PubMed Central PMCID: PMCPmc1380249.
35. Himes BE, Klanderman B, Ziniti J, Senter-Sylvia J, Soto-Quiros ME, Avila L, et al. Association of SERPINE2 with asthma. *Chest*. 2011; 140(3):667–74. Epub 2011/03/26. doi: [10.1378/chest.10-2973](https://doi.org/10.1378/chest.10-2973) PMID: [21436250](https://pubmed.ncbi.nlm.nih.gov/21436250/); PubMed Central PMCID: PMCPmc3168857.
36. Kerkhof M, Boezen HM, Granell R, Wijga AH, Brunekreef B, Smit HA, et al. Transient early wheeze and lung function in early childhood associated with chronic obstructive pulmonary disease genes. *The Journal of allergy and clinical immunology*. 2014; 133(1):68–76 e1–4. doi: [10.1016/j.jaci.2013.06.004](https://doi.org/10.1016/j.jaci.2013.06.004) PMID: [23886569](https://pubmed.ncbi.nlm.nih.gov/23886569/).
37. Dimas AS, Deutsch S, Stranger BE, Montgomery SB, Borel C, Attar-Cohen H, et al. Common regulatory variation impacts gene expression in a cell type-dependent manner. *Science (New York, NY)*. 2009; 325(5945):1246–50. Epub 2009/08/01. doi: [10.1126/science.1174148](https://doi.org/10.1126/science.1174148) PMID: [19644074](https://pubmed.ncbi.nlm.nih.gov/19644074/); PubMed Central PMCID: PMCPmc2867218.
38. Yates LL, Dean CH. Planar polarity: A new player in both lung development and disease. *Organogenesis*. 2011; 7(3):209–16. Epub 2011/10/28. doi: [10.4161/org.7.3.18462](https://doi.org/10.4161/org.7.3.18462) PMID: [22030785](https://pubmed.ncbi.nlm.nih.gov/22030785/); PubMed Central PMCID: PMCPmc3243034.
39. Rai MF, Schmidt EJ, McAlinden A, Cheverud JM, Sandell LJ. Molecular insight into the association between cartilage regeneration and ear wound healing in genetic mouse models: targeting new genes in regeneration. *G3 (Bethesda, Md)*. 2013; 3(11):1881–91. Epub 2013/09/05. doi: [10.1534/g3.113.007302](https://doi.org/10.1534/g3.113.007302) PMID: [24002865](https://pubmed.ncbi.nlm.nih.gov/24002865/); PubMed Central PMCID: PMCPmc3815053.
40. Binet R, Ythier D, Robles AI, Collado M, Larrieu D, Fonti C, et al. WNT16B is a new marker of cellular senescence that regulates p53 activity and the phosphoinositide 3-kinase/AKT pathway. *Cancer research*. 2009; 69(24):9183–91. Epub 2009/12/03. doi: [10.1158/0008-5472.can-09-1016](https://doi.org/10.1158/0008-5472.can-09-1016) PMID: [19951988](https://pubmed.ncbi.nlm.nih.gov/19951988/).
41. Hancock DB, Eijgelsheim M, Wilk JB, Gharib SA, Loehr LR, Marcianti KD, et al. Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function. *Nature genetics*. 2010; 42(1):45–52. Epub 2009/12/17. doi: [10.1038/ng.500](https://doi.org/10.1038/ng.500) PMID: [20010835](https://pubmed.ncbi.nlm.nih.gov/20010835/); PubMed Central PMCID: PMCPmc2832852.