Biological and pathological mechanisms leading to the birth of a small vulnerable newborn

Patricia J Hunter, Toluwalase Awoyemi, Adejumoke I Ayede, R Matthew Chico, Anna L David, Kathryn G Dewey, Christopher P Duggan, Michael Gravett, Andrew J Prendergast, Usha Ramakrishnan, Per Ashorn, Nigel Klein and members of the SVN Steering Committee

UCL Great Ormond Street Institute of Child Health, University College London, London, UK (P J Hunter, PhD, Professor N Klein, MD); Feinberg School of Medicine, Northwestern University, Chicago, IL, USA (T Awoyemi DPhil); Department of Paediatrics, College of Medicine, University of Ibadan and University College Hospital, Ibadan, Nigeria (A I Ayede MSc); Department of Disease Control; Faculty of Infectious & Tropical Diseases; London School of Hygiene & Tropical; London, UK (RM Chico, PhD); Elizabeth Garrett Anderson Institute for Women's Health, University College London, London, UK (Professor A L David, PhD); Department of Nutrition, University of California at Davis, Davis, CA, USA (Professor K G Dewey, PhD); Center for Nutrition, Boston Children's Hospital, Harvard Medical School; Departments of Nutrition and Global Health and Population, Harvard TH Chan School of Public Health; all in Boston, MA, USA, (Professor C P Duggan, MD); Department of Obstetrics & Gynecology, University of Washington, Seattle, WA, USA (Professor M Gravett, MD); Rollins School of Public Health, Emory University, Atlanta, GA, USA (Professor U Ramakrishnan, PhD); Blizard Institute, Queen Mary University of London, London, UK and Zvitambo Institute for Maternal & Child Health Research, Harare, Zimbabwe (Professor A J Prendergast, DPhil); Center for Child, Adolescent, and Maternal Health Research, Faculty of Medicine and Health Technology, Tampere University and Tampere University Hospital, Tampere, Finland, (Professor P Ashorn, MD).

Corresponding author:

Patricia J Hunter, PhD UCL Great Ormond Street Institute of Child Health University College London London, WCIN 1EH, UK

Email: patricia.hunter@ucl.ac.uk

Patricia J Hunter, PhD	Christopher P Duggan, MD
Great Ormond Street Institute of Child Health	Center for Nutrition
University College London	Boston Children's Hospital
London, UK	Harvard Medical School
Email: patricia.hunter@ucl.ac.uk	Departments of Nutrition and Global Health and
	Population
	Harvard TH Chan School of Public Health
	Boston, MA, USA
	Email: Christopher.Duggan@childrens.harvard.edu
Toluwalase Awoyemi, DPhil	Michael Gravett, MD
Feinberg School of Medicine	Departments of Obstetrics & Gynaecology and of Global
Northwestern University	Health
Chicago, IL, USA	University of Washington
Email: toluwalase.awoyemi@northwestern.edu	Seattle, WA, USA
	Email: gravettm@uw.edu
Adejumoke I Ayede, MSc	Andrew J Prendergast, DPhil
Department of Paediatrics	Blizard Institute
College of Medicine	Queen Mary University of London
University of Ibadan	London, UK
And University College Hospital	London, OX
Ibadan, Nigeria	Zvitambo Institute for Maternal & Child Health Research
Email: idayede@yahoo.co.uk	Harare, Zimbabwe
Email: Idayodo e yanoo.co.ux	Email: a.prendergast@qmul.ac.uk
	Linear. a.prenaergaste qinar.ac.ax
R Matthew Chico, PhD	Usha Ramakrishnan, PhD
Department of Disease Control	Hubert Department of Global Health
Faculty of Infectious & Tropical Diseases	Rollins School of Public Health
London School of Hygiene & Tropical	Emory University
London, UK	Atlanta, GA, USA
Email: Matthew.Chico@lshtm.ac.uk	Email: uramakr@emory.edu
Anna L David, PhD	Per Ashorn, MD
Elizabeth Garrett Anderson Institute for Women's	Center for Child, Adolescent, and Maternal Health
Health	Research
University College London	Faculty of Medicine and Health Technology
London, UK	Tampere University and Tampere University Hospital
	Tampere, Finland
National Institute for Health Research University	Email: per.ashorn@tuni.fi
College London Hospital Biomedical Research	
Centre London, UK	
Email: a.david@ucl.ac.uk	
Kathryn G Dewey, PhD	Nigel Klein, MD
Department of Nutrition	Great Ormond Street Institute of Child Health
University of California, Davis	University College London
Davis, CA, USA Email: kgdewey@ucdavis.edu	London, UK
Email. Aguewey @ucuavis.euu	National Institute for Health Research Great Ormond
	Street Hospital Biomedical Research Centre
	London, UK
	Email: n.klein@ucl.ac.uk

Summary

1

- 2 The pathway to a thriving newborn begins pre-conception and continues *in utero* with a
- 3 healthy placenta and the right balance of nutrients and growth factors that are timed and
- 4 sequenced alongside hormonal suppression of labour until a mature infant is ready for birth.
- 5 Optimal nutrition that includes adequate quantities of quality protein, energy, essential fats
- 6 and an extensive range of vitamins and minerals not only supports fetal growth but may also
- 7 prevent preterm birth by supporting the immune system and alleviating oxidative stress.
- 8 Infection, illness, undernourishment, and harmful environmental exposures can alter this
- 9 trajectory leading to an infant who is too small due to either poor growth during pregnancy or
- preterm birth. Systemic inflammation suppresses fetal growth by interfering with growth
- 11 hormone and its regulation of insulin-like growth factors. Evidence supports the prevention
- 12 and treatment of several maternal infections during pregnancy to improve newborn health.
- 13 However, microbes, such as *Ureaplasma* species, that are able to ascend the cervix and cause
- membrane rupture and chorioamnionitis require new strategies for detection and treatment.
- 15 The surge in fetal cortisol late in pregnancy is essential to parturition at the right time, but
- acute or chronically high maternal cortisol levels caused by psychological or physical stress
- may also trigger labour onset prematurely. In every pathway to the small vulnerable newborn,
- there is a possibility to change direction by supporting improved nutrition, protection against
- infection, holistic maternal wellness, and healthy environments.

Keywords

- 21 Preterm birth, fetal growth restriction, small for gestational age, small vulnerable newborn,
- 22 pregnancy, nutrition, infection

23

20

Key messages

1. Factors that influence fetal growth change over course of pregnancy, from the direct exposure to nutrients in maternal fluids during conception, to the formation and function of the placenta, to the timing of bone elongation and fat deposition. Thus, the timing and regulation of nutrient availability is critical in achieving fetal growth potential.

2. Pregnancy is maintained by the active suppression of labour mechanisms by progesterone and other factors and by a long, closed cervix. Thus, there are physical and chemical "barriers" to the initiation of labour and birth that are overcome by signals that the infant is ready to be born. The barriers can be modulated by progesterone insufficiency, diet and environmental contaminants. In addition, high levels of maternal cortisol and severe inflammation can override the barrier leading to preterm labour and birth.

3. Preterm birth and fetal growth restriction may be the endpoints of different pathways but infection, undernourishment, psychological stress and environmental exposures have the potential to act on both pathways through intermediates of oxidative stress, inflammation, inadequate immune protection and placental dysfunction.

4. New knowledge about the mechanisms of pregnancy continues to emerge providing a better understanding of ways to support optimal fetal growth and duration of gestation targeted to those with the greatest ability to benefit, thus affording opportunities for comprehensive, personalised support for pregnant women globally.

Embedded in the United Nations' Sustainable Development Goals is a roadmap to break the cycle of poverty and disadvantage perpetuated by vulnerable childhood and adolescence giving rise to vulnerable pregnancy and infancy. In this series, we examine the vulnerability conferred by small size at birth resulting from growth restriction and/or preterm birth. We cover the prevalence, causes, consequences and possible routes to prevention, either by accelerating existing strategies or considering new approaches. Approximately one in four infants worldwide is born either preterm, small-for-gestational-age or both. Forty per cent of global neonatal mortality occurs in preterm infants and 28% occurs in small-for-gestational-age infants born at term.

Despite global attention and targets set for reducing the prevalence of the small vulnerable newborn, there has been little change over the last 10 years. The slow progress can be attributed in part to gaps in our common understanding of the mechanisms controlling fetal growth and gestational duration. Multiple, often interacting, risk factors contribute to poor health in women both before and during pregnancy (Panel 1). However, connecting risk factors to the biological processes leading to preterm birth and growth restriction remains a challenge. For some of the most prevalent risk factors, the relationship with causal mechanisms may be indirect. For example, maternal iron deficiency anaemia is the largest global population-attributable risk factor for spontaneous preterm and small-for-gestational-age births, 40,41 however iron supplementation (which reduces maternal anaemia by 70%) has not reduced he prevalence of these outcomes in most contexts. A similar conundrum is the global prevalence of bacterial vaginosis and its association with spontaneous preterm birth; 25 years of trials with antibiotics during pregnancy show that treatment can reduce the prevalence of bacterial vaginosis but not the risk of spontaneous preterm birth. 43,44

Within the series, this article reviews the pathway to the birth of a healthy thriving newborn in order to provide a framework to describe what can go wrong. Knowledge of these mechanisms is incomplete, however new information is constantly emerging, often from disciplines outside of mammalian reproduction and development. Novel concepts emerging from randomised controlled trials, animal models, observational studies and laboratory work that recapitulates mechanisms *in vitro* have enabled connections to be made with biological mechanisms in order to explain why some strategies for prevention are effective and some require new approaches. This article will demonstrate that it is useful to consider preterm birth and growth restriction together because many risk factors can contribute to both, albeit through different pathways. Context-specific, targeted and even personalised intervention strategies to prevent preterm and small-for-gestational-age births are possible and likely to bring better health to the next generation.

Born at the right size but how?

Factors influencing the growth and development of the fetus change over the course of pregnancy. The first critical period begins around the time of conception and ends at implantation. At this stage, the embryo can sense the concentrations of nutrients in the surrounding fluids and calibrate of metabolic processes to compensate for overabundance, in the case of maternal obesity, or paucity, in the case of undernutrition.⁴⁵ The subsequent adaptations in embryonic gene expression and regulation can become "fixed" in the form of heritable chromatin changes that can lead to dysregulated fetal growth and obesity and metabolic disease in adulthood.⁴⁶

The next critical period begins with implantation, which triggers a hormonal surge leading to changes in maternal physiology to support placental development and the increased metabolic demands of pregnancy. Fetal trophoblast cells invade the maternal endometrial

spiral arteries, displacing the vascular endothelium and directing larger, stronger versions to be rebuilt on the same tissue scaffold. 47 Proliferating trophoblasts elaborate the basic placental structure, which consists of finger-like villi that float in compartments of maternal blood (Figure 1). Peak placental growth occurs at the end of the first trimester but remodelling of the maternal vasculature continues for the duration of pregnancy (Figure 2). As the placenta develops, it takes over the production of hormones that maintain pregnancy and direct the production of growth factors (Figure 3). Thus, a physiological dialog ensues between the placenta and fetus, and the placenta and pregnant woman. For example, placentally produced hormones create a transient state of mild insulin resistance at the cellular level in the woman, presumably to free up more glucose for the infant.⁴⁸ Excess glucose is taken up and stored as glycogen by the placenta, possibly to buffer the effects of transient moderate undernourishment or to prepare for accelerated weight gain later in gestation.49 The second trimester is the critical period of peak fetal length gain, largely driven by insulinlike growth factors (IGFs) and regulated by growth hormone and a system of six binding proteins and their proteases.⁵⁰ IGF-1 is involved in bone elongation and skeletal growth.⁵¹ IGF-2 drives placental growth as well as the synthesis of other placentally-derived hormones.⁵² The last trimester sees peak fetal weight gain with the enlarging of muscle and laying down of fat under the skin and around the organs. Fat deposition is controlled and regulated by insulin, leptin, adiponectin and other adipokines.⁵³ Undernutrition during the third trimester leads to an infant that is too thin at birth whereas mid trimester undernutrition leads to an infant that is overall too small.⁵⁴ Due to resource allocation to head and brain development (so-called "brain sparing") head growth can follow a normal growth trajectory even when the growth of the fetal body is faltering.⁵⁵ Since maternal weight gain is steadier than that of the infant, it should be possible to identify women who are not gaining adequate

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

weight and intervene to support nutrient intake ahead of peak fetal weight gain in the third trimester.

Born at the right time

Pregnancy is maintained by progesterone-mediated suppression of the processes of labour and by an impenetrable cervix (Figure 2). Progesterone inhibits the production of components involved in receiving signals to prepare the uterus for labour such as the estrogen and oxytocin receptors. In most mammals, plasma progesterone concentrations decrease towards the end of pregnancy. In contrast, levels remain high throughout human pregnancy, even during labour. Activation of labour systems is brought about instead by the functional inhibition of progesterone, possibly by a soluble "A" form of the progesterone receptor (PR-A).⁵⁶

The uterine cervix remains long and closed for the duration of pregnancy due to its rigid structure bestowed by the high collagen content of the extracellular matrix. Compared with many other mammals, the human cervix needs to be strong enough to counteract the downward pressure of weight attributable to the growing fetus during the time the woman spends in the upright position.⁵⁷ Additionally, the cervix needs to be kept free of bacteria ascending from the vagina. Cervical mucus provides a scaffold for immunoglobulins and antimicrobial peptides as it accumulates and forms the mucus plug.^{58,59} The cellular defence of the cervix is mainly provided by neutrophils that populate the mucus having exited the maternal circulation.⁶⁰

Events leading to labour and birth of humans are not fully understood. However, there are pathways observed in other mammals that are likely to operate similarly in humans. A

common view is that signals from the infant indicating that key late developmental milestones have been achieved are also able to start the processes leading to labour and birth. For example, one of the final steps in lung development is the release of surfactant to the surface of the lung alveoli so that when they fill with air at birth, the surface tension will be kept low. Since the lungs are full of amniotic fluid and the infant is performing breathing movements in the womb, the surfactant diffuses throughout the amniotic fluid around the infant. In rodents, the accumulation of surfactant in amniotic fluid acts as a trigger to start the birth process. 61,62 A similar process occurs with fetal cortisol and corticosteroids. Towards the end of pregnancy, the fetal brain signals to increase production of corticotropin releasing hormone which leads to an increase in cortisol and corticosteroids in the fetal circulation (Figure 2).⁶³ As the main steroid involved in the stress response, cortisol directs the release of glucose into the fetal bloodstream and increases blood flow to the brain. It may have the dual function of bringing new alertness and awareness to the infant as well as signalling that the infant is ready for parturition to begin. The first committed step toward labour occurs when cortisol and corticosteroids in the fetal circulation reach the threshold for the activation of the production of the cyclooxygenase 2

164

165

166

167

168

169

170

171

172

173

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

circulation reach the threshold for the activation of the production of the cyclooxygenase 2 (COX2) in the fetal membranes (figure 2). COX2 converts long chain polyunsaturated fatty acids (LCPUFAs) into prostaglandins. The essential LCPUFA for labour is arachidonic acid, which selectively accumulates in the myometrium, cervix and fetal membranes over the course of pregnancy. COX2 converts arachidonic acid into prostaglandins E2 and F2 α , which trigger a gene and protein expression cascade, leading to the functional inhibition of progesterone, the production of contraction-associated proteins and the recruitment of monocytes and neutrophils to the uterus and cervix. These cells produce matrix

metalloproteinases which dissolve the extracellular collagen matrix of the myometrium and cervix causing the cervix to soften.⁶⁶ Tight gap junctions form between the cells of the myometrium, which then takes on the appearance and function of smooth muscle.

Omega-3 LCPUFAs are also substrates of COX2 and may act as competitive inhibitors of prostaglandin E2 and F2α production thus contributing to the maintenance of pregnancy and the inhibition of labour.⁶⁷ Women with lower circulating concentration of omega-3 LCPUFAs are at increased risk of preterm birth,⁶⁸ suggesting that these compounds, like progesterone, act to raise the threshold for the activation of labour processes. One of the unintended consequences of supplementation with omega-3 LCPUFAs is an increase in the rate of post term birth⁶⁹ suggesting that if the threshold is too high, signals from the fetus can't overcome the inhibitory mechanisms and the pregnancy is prolonged.

Good nutrition supports more than just growth

The impact of maternal nutrition before and during pregnancy is now understood to extend well beyond birth and childhood into the life courses of future generations. ^{45,70} Physiological changes in pregnancy enable women to meet the increased demand for energy, nutrients, and oxygen to supply to the growing fetus (Table 1). However, women who begin a pregnancy before having reached their own biological growth potential due to chronic undernourishment, young age, or both, are at increased risk of being unable to meet these demands. Among underweight women, partitioning of energy and nutrients may result in limited provision to the fetus in favour of maternal requirements for her survival. Thus, it is not surprising that underweight women, who may also have inadequate gestational weight gain, are at higher risk of delivering a small-for-gestational-age infant. ^{7,77}

Anaemia is a highly prevalent risk factor linked to a wide range of adverse pregnancy outcomes. 78 There are many causes of anaemia unrelated to nutrition including malaria and other infectious/inflammatory conditions. However, iron supplementation during pregnancy independently reduces the prevalence of anaemia, suggesting that iron deficiency is a key contributor. 42 Anaemia, as a measurable risk factor, may also identify women with a wider range of micronutrient deficiencies. Supplementation with a broad range of micronutrients is able to lower the risk of small-for-gestational age births, ^{79,80} particularly among underweight and anaemic women. 80 in comparison to iron and folic acid alone. This positive effect on growth without the provision of energy is likely conferred by the efficiency gained when multiple metabolic processes are supported simultaneously. Provision of micronutrients may also lower the risk of preterm birth in underweight women.⁸¹ There are many mechanisms that might contribute to this effect listed in Table 2. We will expand on the ability of good nutrition to enhance immune responses and reduce damage caused by oxidative stress. Damage to tissue caused by the accumulation of reactive oxygen species is both a threat to pregnancy and a natural consequence of oxygen regulation in the placenta. 85 Micronutrients with antioxidant properties including vitamins C and E, carotenoids and long-chain polyunsaturated fatty acids (LCPUFAs) can reduce oxidative stress. The body can dismantle reactive oxygen species using enzymes such as superoxide dismutase, glutathione reductase and various peroxidases that can catalyse their binding to antioxidant molecules. However, once an antioxidant is peroxidated, it is removed from tissue leading to increased turnover and reduced bioavailability. 86 The pathway to spontaneous preterm birth caused by oxidative stress may involve the increased turnover of LCPUFAs, particularly docosahexaenoic acid, which, as previously discussed, may act as a natural inhibitor of labour. People who smoke cigarettes carry a higher burden of oxidative damage compared with non-smokers, 87 and have lower levels of endogenous omega-3 LCPUFAs. 88 Thus, it is unsurprising that a trial

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

comparing omega-3 LCPUFA supplementation with placebo in pregnant women found spontaneous preterm birth reduced by almost one-half in smokers, whereas there was no benefit in non-smokers.⁸⁹ Zinc is an essential co-factor for superoxide dismutase and a wide range of enzymes and transcription factors, and its deficiency is associated with immune dysfunction and increased susceptibility to infection. 90 White blood cells require tenfold more zinc in comparison to red blood cells. 91 In a healthy pregnancy, there is an increase in white blood cell counts, largely due to the 50% increase in neutrophils.⁷⁴ As one of the first lines of defence against pathogens, neutrophils are ubiquitous at points of entry into the body. In pregnancy, they are crucial to defending the cervix against ascending infection. ⁶⁰ Recent evidence supports previously unknown roles for neutrophils in vascular and tissue remodelling. 92 The secretion of matrix metalloproteinases, for which zinc is a cofactor, by neutrophils is likely to be essential for this latter role. Blocking neutrophils, 93 knocking out matrix metalloproteinases, 94 and reducing bioavailable zinc,95 all have detrimental effects on placentation in mice leading to fetal demise. The roles of neutrophils and zinc in placentation and protection against pathways leading to preterm birth are only just beginning to be understood and represent a new frontier in reproductive biology.

240

241

242

243

244

245

246

247

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

Infectious threats to the fetus

Microbial infections in pregnant women are major contributors to preterm birth, growth restriction, stillbirth and infection in newborns. Screening for and treating infections in pregnant women has well-established positive effects and there is a need for wider coverage for syphilis, chlamydia, gonorrhoea, HIV, and malaria. However, even in parts of the world where the prevalence of these infections is low, the majority of spontaneous preterm birth – that is, preterm birth preceded by labour or preterm pre-labour rupture of membranes – is also

likely to be caused by microbial infection given the high prevalence of chorioamnionitis found in membrane and placenta tissue on histopathological examination. 96-98

Chorioamnionitis refers to infiltration of the fetal membranes by maternal neutrophils. It is usually asymptomatic during pregnancy and the diagnosis is made after the birth of the infant. Whilst it is presumed to be caused by colonisation by bacteria that ascended the cervix from the vagina, identification of microbes in these tissues is seldom undertaken. When molecular methods are used to detect microbes in fetal membranes, the most common species identified are members of the *Ureaplasma* genus of bacteria. 99-101 Some species of *Ureaplasma* are able to break down antimicrobial defences and exploit natural weaknesses in the immune system that are unmasked by pregnancy in some women. This may explain the association between spontaneous preterm birth and both periodontal disease and urinary tract infections. The mouth, the vagina, and the urinary tract are dependent on the same mechanisms (antibodies, antimicrobial peptides and neutrophils) to protect against microbial invasion.

There are three general pathways through which infection could lead to spontaneous preterm birth. First, there are likely unique features of certain bacterial species, as opposed to viruses or parasites, that trigger the expression of COX2 on their invasion of the placenta, fetal membranes or amniotic fluid. Injecting bacteria or bacterial products into the uteri of pregnant mice is the most widely-used method of modelling preterm birth. It could be that COX2 can be upregulated by signalling through molecules, such as toll-like receptors 2 and 4, that specifically recognize certain types of bacteria and bacterial products. Secondly, microbes that are able to ascend the cervix from the vagina could simply damage the fetal membranes causing rupture (Figure 5). In this scenario, there may not be inflammation or the

activation of mechanisms that lead to labour. In many cases of preterm pre-labour rupture of membranes, labour does not occur after a sufficient period of time and the infant must be delivered by labour induction or Caesarean section due to loss of amniotic fluid and the concerns regarding the potential for systemic spread of the infection. Finally, high levels of inflammatory cytokines in the placenta and may be able to activate COX2 and the pathways that culminate in labour. This may be an evolutionary adaptation to delivery the infant from an unfavourable environment where the mother's life is under threat.

Inflammation likely suppresses fetal growth by inhibiting the growth hormone/insulin-like growth factor (GH/IGF) axis (Figure 4). In a study comparing maternal plasma, placental, and cord blood levels of IGF-1 and its inhibitory binding proteins in pregnancies with and without placental malaria, IGF-1 levels were reduced by 28% in plasma samples from women with placental malaria and by 25% in their neonates compared with samples from uninfected women. The inhibitory IGF binding protein-1 was elevated in cord blood of neonates with placental malaria. The inhibitory IGF binding protein-1 was elevated in cord blood of neonates with

Clues to the molecular interactions between inflammation and growth factors come from the observation of poor growth in children with systemic inflammation, ¹⁰⁷ and elevated inflammation in children with poor growth. ¹⁰⁸ A surprising result of treating children with anti-tumour necrosis factor alpha and other anti-cytokine therapeutics for inflammatory conditions was the restoration of normal growth trajectory. ¹⁰⁷ Studies in mice indicate that interleukin-6, a key inflammatory cytokine that is elevated in response to infection, may have the ability to directly suppress IGF-1 and growth hormone. ¹⁰⁹ The slowing of growth in response to inflammation may be an evolutionary adaptation to promote successful vaginal

birth. As the mother's body prepares for labour, the increase in systemic inflammatory cytokines may contribute to the observed slowing of head growth at the end of pregnancy.

Cervical shortening and preterm birth

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

When a woman's cervix shortens in the course of pregnancy, there is an increased risk of preterm birth. It is not known why this occurs in some women, but it is associated with the premature expression of proteins involved in the recruitment of monocytes and neutrophils which could lead to the premature destruction of collagen and loss of integrity. 110 As a key hormone responsible for maintaining pregnancy, progesterone may be able to disrupt this process. Progesterone delivered directly to the cervix in soluble capsules, injected intramuscularly (IM) or taken as tablets has been tested in randomized controlled trials to determine its effect on preterm birth. A recent individual patient data meta-analysis revealed that both vaginal (9 trials, 3769 women) and oral (2 trials, 183 women) progesterone supplementation are effective at reducing preterm birth before 34 weeks of gestation in high risk women, namely those with a previous preterm birth or a short cervix (< 25 mm). 111 The evidence of benefit in reducing birth before 34 weeks is less certain for IM progesterone (5 trials, 3053 women). 111 Furthermore, there have been recent concerns about the maternal and neonatal safety of the synthetic version of progesterone (17-hydroxyprogesterone caproate) used for IM administration. 112 In light of this data, vaginal progesterone remains the most promising treatment to prolong gestation for women with a short cervix. Serial ultrasound surveillance of cervical length is required to reliably detect cervical shortening, which may preclude the use of cervical monitoring in resource-poor settings. Analysis of soluble factors in amniotic and vaginal fluids have identified macrophage chemoattractant protein as a biomarker with the strongest association with cervical shortening. 110,113,114 Macrophage chemoattractant protein 1 is easy to detect in mucus from

the vaginal end of the cervix and holds potential to report cervical shortening with minimal equipment.

Pre-eclampsia, fetal growth restriction and preterm birth

Major problems arising during implantation and early placental development result in miscarriage. However, minor issues often remain silent until around mid-gestation when the fetus overtakes the placenta in size. At this time, minor inadequacies in placental size, patterning or maternal blood supply can result in an inability to meet the requirements for the growth and development of the fetus. For reasons that are not completely understood, one of the most common signs that there are supply-and-demand issues with a pregnancy is the elevation of the pregnant woman's blood pressure. The clinical definition of pre-eclampsia has recently been expanded to include the development of high blood pressure during pregnancy along with any related problem, not only elevated protein in the urine. Five percent of pregnancies worldwide are affected by pre-eclampsia with 76,000 attributable maternal deaths per year, second only to post-partum haemorrhage as a cause of maternal death. Around 500,000 fetal and newborn deaths each year are attributed to pre-eclampsia and eclampsia. Approximately 9% of all preterm birth is by induction of labour or Caesarean section to treat severe pre-eclampsia and eclampsia.

Pre-existing maternal cardiovascular vulnerability and poor cardiovascular adaptation to pregnancy are increasingly recognised as important to the development of pre-eclampsia. 116

Pregnancy has even been described as a stress-test that reveals women who have poor cardiovascular reserve or dysfunction. 117 It is therefore unsurprising that well-established treatments for cardiovascular disease such as low-dose aspirin, when given during pregnancy, also reduce the risk of preterm pre-eclampsia, 118 and new treatments (statins) are under investigation. 119

A calcium-rich diet or calcium supplementation during pregnancy are also able to reduce the risk of pre-eclampsia and associated morbidity and mortality in the newborn.⁴ It is likely that both aspirin and calcium are able to prevent the establishment of a systemic vasoconstrictive environment. In chronic, sustained high blood pressure, the ratio of the vasoconstrictive thromboxane to the vasodilator prostacyclin is skewed towards vasoconstriction. Both molecules are synthesized by cyclooxygenases 1 and 2 (COX1/2). At low doses, aspirin appears to be able selectively and irreversibly to inactivate COX1 in platelets, thus reducing thromboxane production and restoring this ratio to normotensive levels.¹²⁰ However, aspirin has been shown to be most effective at preventing preterm pre-eclampsia when commenced early in pregnancy (< 16 weeks) suggesting a supportive effect on early placentation.¹²¹

Changing social and environmental contexts

Some subgroups of pregnant women, such as smokers, primi- and secundigravidae, teenagers, and women with low body mass index scores, tend to respond more favourably to nutrient supplementation or preventive treatment of infections, reducing the risk of delivering small and vulnerable newborns. However, this does not justify the exclusive use of these interventions strategies to reduce the prevalence of small vulnerable newborns. Increased antenatal contacts afford opportunities to address the wellbeing of pregnant women in a more holistic way. Depression, anxiety, lack of agency, chronic illness, physical workload and intimate partner abuse can all be exacerbated by pregnancy. High levels of psychological and physical stress during pregnancy are associated with growth restriction and shorter pregnancy duration. 122-124 Cortisol entering the placenta from the fetal circulation is an important step in the preparation of mother and child for birth. Although increases in cortisol and corticotropin releasing hormone in the mother's circulation are normal during pregnancy, it is possible that

prolonged elevated or acute bursts of cortisol may be able to trigger preterm labour. Furthermore, elevated cortisol has also been associated with higher concentrations of proinflammatory cytokines, 125,126 that can negatively affect fetal growth as previously described (Figure 4). Creation of energy from oxygen combined with glucose and other monosaccharides is the final step in the pathway that powers fetal growth. The pathway starts with clean air that is free of pollutants that interfere with oxygen binding by maternal hemoglobin. In addition to increasing the burden of oxidative stress, smoking and cooking over biomass fuels can limit oxygen delivery to the placenta (Figure 4). 127 Exposure to air pollution and living at high altitude have also been linked to fetal growth restriction. 128,129 Interventions that help women to quit or reduce smoking during pregnancy reduce the risk of giving birth to a small infant. 130 Countries that have banned smoking in indoor public spaces have experienced a dramatic reduction in the prevalence of preterm and low birth weight newborns. 131-133 Lowand middle-income countries have higher outdoor pollution levels and indoor pollution due to a reliance on solid biomass (usually wood) fuels and chimneyless stoves for cooking and heating. 134 Because women are more exposed to indoor pollution from cookstoves and heating due to a greater amount of time spent in the home, the World Health Organization considers indoor pollution as a "silent killer" of women in low-resource settings. 135 Trials of liquid fuel cookstoves have so far failed to demonstrate their ability to lower the risk of low

393

394

395

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

New evidence is emerging on the effect extra heat on pregnancy outcomes, with a 5% (95% CI 3% - 7%) increase in the odds of having a preterm birth every one degrees above seasonal

birth weight, preterm birth or small-for-gestational-age births, possibly because they are

unable to sufficiently reduce airborne particulate matter to have an observable effect. 136,137

average.^{38,138} Further epidemiological evidence suggests that conception and early first trimester are particularly vulnerable to heat stress, increasing the risk of stillbirth and preterm birth.¹³⁹ In animals, transient elevated temperatures lead to reduced feeding and overall food intake resulting in growth restriction in the fetus.¹⁴⁰ However, the damage may run deeper with loss of intestinal barrier function, changes to intestinal epithelial morphology.¹⁴¹

Food and water-borne pollutants are also likely to contribute to the prevalence of small vulnerable newborns. Components of Aspergillus fungal spores collectively known as aflatoxins are common contaminants of food production in under-resourced settings. ¹⁴² High concentrations of aflatoxins in maternal and cord blood are associated with low birthweight, likely mediated through growth restriction, although the exact mechanism is not known. ³⁴ In addition to known teratogenic and carcinogenic effects of aflatoxins, they may also interfere with hormone secretion and signaling and thus are part of a wider group of both natural and artificial toxicants known as endocrine disruptors, which include bisphenol A, phthalates, pesticides, polychlorinated biphenyls, polybrominated diethyl ethers and dioxins. ³⁵ Of particular concern is the high levels of phthalate metabolites that contaminate food and water globally. In keeping with their role in modulating estrogen levels, different phthalate compounds can increase or reduce gestational length and are therefore associated with both pre- and post-term birth. ¹⁴³ Governments have sought to ban the use of phthalates in plastics production, however the toxicity of potential replacements is uncertain. ³⁵

What can be done? The foreground and the horizon

Knowledge of the mechanisms that lead to the birth of a small vulnerable newborn continues to grow as well as our understanding of how to intervene to reduce or prevent this outcome.

In the short term, increasing the quantity and quality of antenatal contacts with healthcare

providers affords the opportunity to monitor and support physical (weight gain, fetal growth, prevention and treatment of pregnancy complications) and psychological (mental health, agency) wellbeing. Reductions in preterm birth and growth restriction can be achieved with broader implementation of proven antenatal interventions, including multiple micronutrient supplements, balanced protein energy supplements, aspirin, treatment of syphilis, education for smoking cessation, prevention of malaria in pregnancy, treatment of asymptomatic bacteriuria, and progesterone provided vaginally as presented with this series. ¹⁴⁴ In addition, the specific vulnerability of those *in utero* to poor air quality, heat waves and toxins in food and water should contribute the urgency of global efforts to reduce harmful environmental exposures and the impact of climate change.

In the longer term, new knowledge can be used to improve our understanding of the molecular and cellular biology underlying risk factors that inform interventions for populations with the greatest ability to benefit. Risk stratification tools and algorithms that incorporate individual risk profiles, together with biomarkers, can identify individuals who might benefit from pre-emptive care and early pathway-specific interventions. For example, a test that predicts future cervical shortening would identify women who are most likely to benefit from progesterone supplementation without the need for serial ultrasound monitoring. Progesterone supplementation itself is also evolving with new analogues that are resistant to inhibition by the mechanisms that lead to labour. Tests that can be performed and interpreted in the timescale of an antenatal care visit (point-of-care tests) will improve uptake of treatment for infections; treatment can be issued on the same day removing the need to return to clinic for follow-up. Point-of-care tests should fulfil the WHO ASSURED (Affordable, Sensitive, Specific, User-friendly, Rapid, and Equipment-free, and Deliverable) criteria for use in low resource settings. Tests that informs interventions for such algorithms that incorporate interventions are used to such a such as a

Placental histopathology is underutilized as a means to diagnose chorioamnionitis and other placental conditions leading to birth of small vulnerable newborns. In cases of preterm prelabour rupture of membranes, the rupture site is the "scene of the crime" and should be fully investigated. If *Ureaplasma* species are the leading cause of spontaneous preterm birth, prevalence and virulence factors need to be resolved at the level of species. It will be important to demonstrate a causal relationship between species and spontaneous labour and membrane rupture so that antibiotics that can "cure" the individual and prevent these outcomes are not overused.

There are also new opportunities to understand placental health *in situ*. A particularly promising development is the discovery of extracellular vesicles which are small particles consisting of a lipid bilayer containing the proteins, metabolites, RNA, and DNA that have budded off from a parent cell. In pregnancy, extracellular vesicles in the maternal circulation mainly come from fetal trophoblasts of the placenta. Extracellular vesicles in a peripheral blood may reveal key aspects of the placental environment including oxygen tension, glucose concentration, inflammation, and vascular dysfunction. In abnormal states such as gestational diabetes and pre-eclampsia, numbers of extracellular vesicles are elevated and contain molecular signatures of these conditions. ¹⁴⁸

Every woman's journey through pregnancy and childbirth is unique and the ultimate goal should be individually tailored care for all with an eye towards optimizing both mother and infant health and wellbeing. Personalized antenatal care does not need to be complex or expensive but the barriers may be higher in low- and middle income settings in comparison with a pragmatic public health approach. Interventions can span from the bedside (e.g., better

gestational age assessment) to the clinic (e.g. pre-eclampsia screening) to the operating room (e.g. safer anaesthesia for Caesarean sections) and to society generally (e.g. limiting tobacco or pollution exposure). A more precise deployment of the existing toolkit of interventions is likely to be more cost effective. However, many aspects of even healthy pregnancy remain poorly understood, and it is only with continuous discovery that we move forward.

Authors' contributions

PA and NK, in collaboration with other members of the Lancet SVN steering committee, designed the study. NK and PJH verified the underlying data and PJH conducted the analyses. All authors participated in the conceptualisation and drafting of the original manuscript, reviewed and edited subsequent drafts, and approved the final version of the manuscript.

Funding

The production of this manuscript was funded by a grant from the Children's Investment Fund Foundation to PA (grant number 2004-04635).

The funder had no role in the production of the manuscript or the decision to submit it for publication.

Declarations of competing interests

PA reports grants from Children's Investment Fund Foundation (CIFF) and the Bill & Melinda Gates Foundation concurrent with production of the manuscript. AD is an unpaid Scientific Trustee of Tommy's Charity. CD reports grant support from the WHO, USAID and NIH (P30 DK040561). He receives royalties from UpToDate. His institution receives grant support from Takeda and the American Society for Nutrition in work unrelated to this paper. UR receives honoraria from NIH for educational activities. Her institution receives funding from NIH. All other authors declare no competing interests.

Author collaborative groups

Lancet Small Vulnerable Newborn Steering Committee: Per Ashorn, Robert E Black, Joy E Lawn, Ulla Ashorn, Nigel Klein, G Justus Hofmeyr, Marleen Temmerman, Sufia Askari

References

- 1. Lawn JE, Ohuma EO, Bradley E et al. Small babies, big risks: Global estimates of prevalence and mortality for vulnerable newborns to accelerate change and improve counting. Lancet SVN series paper 2.
- 2. Rahman MM, Abe SK, Rahman MS, et al. Maternal anemia and risk of adverse birth and health outcomes in low- and middle-income countries: systematic review and meta-analysis. Am J Clin Nutr. 2016 Feb;103(2):495-504. doi: 10.3945/ajcn.115.107896. Epub 2016 Jan 6. PMID: 26739036.
- 3. Carducci B, Keats EC, Bhutta ZA. Zinc supplementation for improving pregnancy and infant outcome. Cochrane Database Syst Rev. 2021 Mar 16;3(3):CD000230. doi: 10.1002/14651858.CD000230.pub6. PMID: 33724446; PMCID: PMC8094617.
- 4. Hofmeyr GJ, Lawrie TA, Atallah ÁN, Torloni MR. Calcium supplementation during pregnancy for preventing hypertensive disorders and related problems. Cochrane Database Syst Rev. 2018 Oct 1;10(10):CD001059. doi: 10.1002/14651858.CD001059.pub5. PMID: 30277579; PMCID: PMC6517256.
- 5. Kozuki N, Katz J, Lee AC, et al. Short Maternal Stature Increases Risk of Smallfor-Gestational-Age and Preterm Births in Low- and Middle-Income Countries: Individual Participant Data Meta-Analysis and Population Attributable Fraction. J Nutr. 2015 Nov;145(11):2542-50. doi: 10.3945/jn.115.216374. Epub 2015 Sep 30. PMID: 26423738; PMCID: PMC6457093.
- 6. Han Z, Mulla S, Beyene J, Liao G, McDonald SD. Maternal underweight and the risk of preterm birth and low birth weight: a systematic review and meta-analyses. Int J Epidemiol. 2011 Feb;40(1):65-101. doi: 10.1093/ije/dyq195. Epub 2010 Nov 22. PMID: 21097954.
- 7. Goldstein RF, Abell SK, Ranasinha S, Misso ML, Boyle JA, Harrison CL, Black MH, Li N, Hu G, Corrado F, Hegaard H, Kim YJ, Haugen M, Song WO, Kim MH, Bogaerts A, Devlieger R, Chung JH, Teede HJ. Gestational weight gain across continents and ethnicity: systematic review and meta-analysis of maternal and infant outcomes in more than one million women. BMC Med. 2018 Aug 31;16(1):153. doi: 10.1186/s12916-018-1128-1. PMID: 30165842; PMCID: PMC6117916.
- 8. Xiao PL, Zhou YB, Chen Y, et al. Association between maternal HIV infection and low birth weight and prematurity: a meta-analysis of cohort studies. BMC Pregnancy Childbirth. 2015 Oct 8;15:246. doi: 10.1186/s12884-015-0684-z. PMID: 26450602; PMCID: PMC4599647.

- 9. Steketee RW, Nahlen BL, Parise ME, Menendez C. The burden of malaria in pregnancy in malaria-endemic areas. Am J Trop Med Hyg. 2001 Jan-Feb;64(1-2 Suppl):28-35. doi: 10.4269/ajtmh.2001.64.28. PMID: 11425175.
- 10. Qin J, Yang T, Xiao S, Tan H, Feng T, Fu H. Reported estimates of adverse pregnancy outcomes among women with and without syphilis: a systematic review and meta-analysis. PLoS One. 2014 Jul 15;9(7):e102203. doi: 10.1371/journal.pone.0102203. PMID: 25025232; PMCID: PMC4099012.
- 11. Olson-Chen C, Balaram K, Hackney DN. Chlamydia trachomatis and Adverse Pregnancy Outcomes: Meta-analysis of Patients With and Without Infection. Matern Child Health J. 2018 Jun;22(6):812-821. doi: 10.1007/s10995-018-2451-z. PMID: 29417367.
- 12. Vallely LM, Egli-Gany D, Wand H, et al. Adverse pregnancy and neonatal outcomes associated with *Neisseria gonorrhoeae:* systematic review and meta-analysis. Sex Transm Infect. 2021 Mar;97(2):104-111. doi: 10.1136/sextrans-2020-054653. Epub 2021 Jan 12. PMID: 33436505; PMCID: PMC7892372.
- 13. Ansaldi Y, Martinez de Tejada Weber B. Urinary tract infections in pregnancy. Clin Microbiol Infect. 2022 Aug 27:S1198-743X(22)00431-1. doi: 10.1016/j.cmi.2022.08.015. Epub ahead of print. PMID: 36031053.
- 14. Leitich H, Kiss H. Asymptomatic bacterial vaginosis and intermediate flora as risk factors for adverse pregnancy outcome. Best Pract Res Clin Obstet Gynaecol. 2007 Jun;21(3):375-90. doi: 10.1016/j.bpobgyn.2006.12.005. Epub 2007 Jan 22. PMID: 17241817.
- 15. Cotch MF, Pastorek JG 2nd, Nugent RP, et al. Trichomonas vaginalis associated with low birth weight and preterm delivery. The Vaginal Infections and Prematurity Study Group. Sex Transm Dis. 1997 Jul;24(6):353-60. doi: 10.1097/00007435-199707000-00008. PMID: 9243743.
- 16. Bianchi-Jassir F, Seale AC, Kohli-Lynch M, et al. Preterm Birth Associated With Group B Streptococcus Maternal Colonization Worldwide: Systematic Review and Meta-analyses. Clin Infect Dis. 2017 Nov 6;65(suppl_2):S133-S142. doi: 10.1093/cid/cix661. PMID: 29117329; PMCID: PMC5850429.
- 17. Kozuki N, Lee AC, Silveira MF, et al. The associations of parity and maternal age with small-for-gestational-age, preterm, and neonatal and infant mortality: a meta-analysis. BMC Public Health. 2013;13 Suppl 3(Suppl 3):S2. doi: 10.1186/1471-2458-13-S3-S2. Epub 2013 Sep 17. PMID: 24564800; PMCID: PMC3847520.
- 18. Kozuki N, Lee AC, Silveira MF, et al. The associations of birth intervals with small-for-gestational-age, preterm, and neonatal and infant mortality: a meta-analysis. BMC Public Health. 2013;13 Suppl 3(Suppl 3):S3. doi: 10.1186/1471-2458-13-S3-S3. Epub 2013 Sep 17. PMID: 24564484; PMCID: PMC3847557.

- 19. Barros FC, Papageorghiou AT, Victora CG, et al. The distribution of clinical phenotypes of preterm birth syndrome: implications for prevention. JAMA Pediatr. 2015 Mar;169(3):220-9. doi: 10.1001/jamapediatrics.2014.3040. PMID: 25561016.
- 20. Chisholm KM, Folkins AK. Placental and Clinical Characteristics of Term Small-for-Gestational-Age Neonates: A Case-Control Study. Pediatr Dev Pathol. 2016 Jan-Feb;19(1):37-46. doi: 10.2350/15-04-1621-OA.1. Epub 2015 Sep 14. PMID: 26368794.
- 21. Hedderson MM, Ferrara A, Sacks DA. Gestational diabetes mellitus and lesser degrees of pregnancy hyperglycemia: association with increased risk of spontaneous preterm birth. Obstet Gynecol. 2003 Oct;102(4):850-6. doi: 10.1016/s0029-7844(03)00661-6. PMID: 14551018.
- 22. Derakhshan A, Peeters RP, Taylor PN, et al. Association of maternal thyroid function with birthweight: a systematic review and individual-participant data meta-analysis. Lancet Diabetes Endocrinol. 2020 Jun;8(6):501-510. doi: 10.1016/S2213-8587(20)30061-9. PMID: 32445737; PMCID: PMC8168324.
- 23. Iams JD, Goldenberg RL, Meis PJ et al. The length of the cervix and the risk of spontaneous premature delivery. National Institute of Child Health and Human Development Maternal Fetal Medicine Unit Network. N Engl J Med. 1996 Feb 29;334(9):567-72. doi: 10.1056/NEJM199602293340904. PMID: 8569824.
- 24. Panagiotopoulos M, Tseke P, Michala L. Obstetric Complications in Women With Congenital Uterine Anomalies According to the 2013 European Society of Human Reproduction and Embryology and the European Society for Gynaecological Endoscopy Classification: A Systematic Review and Meta-analysis. Obstet Gynecol. 2022 Jan 1;139(1):138-148. doi: 10.1097/AOG.00000000000004627. PMID: 34856567.
- 25. Bruun MR, Arendt LH, Forman A, Ramlau-Hansen CH. Endometriosis and adenomyosis are associated with increased risk of preterm delivery and a small-for-gestational-age child: a systematic review and meta-analysis. Acta Obstet Gynecol Scand. 2018 Sep;97(9):1073-1090. doi: 10.1111/aogs.13364. Epub 2018 May 29. PMID: 29753309.
- 26. Vogel JP, Torloni MR, Seuc A, et al. Maternal and perinatal outcomes of twin pregnancy in 23 low- and middle-income countries. PLoS One. 2013 Aug 1;8(8):e70549. doi: 10.1371/journal.pone.0070549. PMID: 23936446; PMCID: PMC3731264.
- 27. Nelson HD, Darney BG, Ahrens K, et al. Associations of Unintended Pregnancy With Maternal and Infant Health Outcomes: A Systematic Review and Meta-analysis. JAMA. 2022 Nov 1;328(17):1714-1729. doi: 10.1001/jama.2022.19097. PMID: 36318133; PMCID: PMC9627416.

- 28. Donovan BM, Spracklen CN, Schweizer ML, Ryckman KK, Saftlas AF. Intimate partner violence during pregnancy and the risk for adverse infant outcomes: a systematic review and meta-analysis. BJOG. 2016 Jul;123(8):1289-99. doi: 10.1111/1471-0528.13928. Epub 2016 Mar 9. PMID: 26956568.
- 29. Thomson K, Moffat M, Arisa O, et al. Socioeconomic inequalities and adverse pregnancy outcomes in the UK and Republic of Ireland: a systematic review and meta-analysis. BMJ Open. 2021 Mar 15;11(3):e042753. doi: 10.1136/bmjopen-2020-042753. PMID: 33722867; PMCID: PMC7959237.
- 30. Ghimire U, Papabathini SS, Kawuki J, Obore N, Musa TH. Depression during pregnancy and the risk of low birth weight, preterm birth and intrauterine growth restriction- an updated meta-analysis. Early Hum Dev. 2021 Jan;152:105243. doi: 10.1016/j.earlhumdev.2020.105243. Epub 2020 Oct 24. PMID: 33190020.
- 31. Shah NR, Bracken MB. A systematic review and meta-analysis of prospective studies on the association between maternal cigarette smoking and preterm delivery. Am J Obstet Gynecol. 2000 Feb;182(2):465-72. doi: 10.1016/s0002-9378(00)70240-7. PMID: 10694353; PMCID: PMC2706697.
- 32. Patra J, Bakker R, Irving H, Jaddoe VW, Malini S, Rehm J. Dose-response relationship between alcohol consumption before and during pregnancy and the risks of low birthweight, preterm birth and small for gestational age (SGA)-a systematic review and meta-analyses. BJOG. 2011 Nov;118(12):1411-21. doi: 10.1111/j.1471-0528.2011.03050.x. Epub 2011 Jul 6. PMID: 21729235; PMCID: PMC3394156.
- 33. Gouin K, Murphy K, Shah PS, et al. Effects of cocaine use during pregnancy on low birthweight and preterm birth: systematic review and metaanalyses. Am J Obstet Gynecol. 2011 Apr;204(4):340.e1-12. doi: 10.1016/j.ajog.2010.11.013. Epub 2011 Jan 22. PMID: 21257143.
- 34. da Silva JVB, de Oliveira CAF, Ramalho LNZ. Effects of Prenatal Exposure to Aflatoxin B1: A Review. Molecules. 2021 Dec 2;26(23):7312. doi: 10.3390/molecules26237312. PMID: 34885894; PMCID: PMC8659025.
- 35. Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, Prins GS, Toppari J, Zoeller RT. Executive Summary to EDC-2: The Endocrine Society's Second Scientific Statement on Endocrine-Disrupting Chemicals. Endocr Rev. 2015 Dec;36(6):593-602. doi: 10.1210/er.2015-1093. Epub 2015 Sep 28. PMID: 26414233; PMCID: PMC4702495.
- 36. Amegah AK, Quansah R, Jaakkola JJ. Household air pollution from solid fuel use and risk of adverse pregnancy outcomes: a systematic review and meta-analysis of the empirical evidence. PLoS One. 2014 Dec 2;9(12):e113920. doi: 10.1371/journal.pone.0113920. PMID: 25463771; PMCID: PMC4252082.

- 37. Fleischer NL, Merialdi M, van Donkelaar A, et al. Outdoor air pollution, preterm birth, and low birth weight: analysis of the world health organization global survey on maternal and perinatal health. Environ Health Perspect. 2014 Apr;122(4):425-30. doi: 10.1289/ehp.1306837. Epub 2014 Feb 7. Erratum in: Environ Health Perspect. 2014 Jun;122(6):A151. PMID: 24508912; PMCID: PMC3984219.
- 38. Chersich MF, Pham MD, Areal A, et al. Associations between high temperatures in pregnancy and risk of preterm birth, low birth weight, and stillbirths: systematic review and meta-analysis. BMJ. 2020 Nov 4;371:m3811. doi: 10.1136/bmj.m3811. PMID: 33148618; PMCID: PMC7610201.
- 39. Jensen GM, Moore LG. The effect of high altitude and other risk factors on birthweight: independent or interactive effects? Am J Public Health. 1997 Jun;87(6):1003-7. doi: 10.2105/ajph.87.6.1003. PMID: 9224184; PMCID: PMC1380938.
- 40. Bryce E, Gurung S, Tong H, et al. Population attributable fractions for risk factors for spontaneous preterm births in 81 low- and middle-income countries: A systematic analysis. J Glob Health. 2022 Mar 26;12:04013. doi: 10.7189/jogh.12.04013. PMID: 35356651; PMCID: PMC8959104.
- 41. Gurung S, Tong HH, Bryce E, et al. A systematic review on estimating population attributable fraction for risk factors for small-for-gestational-age births in 81 low- and middle-income countries. J Glob Health. 2022 Mar 26;12:04024. doi: 10.7189/jogh.12.04024. PMID: 35356650; PMCID: PMC8942297.
- 42. Peña-Rosas JP, De-Regil LM, Garcia-Casal MN, Dowswell T. Daily oral iron supplementation during pregnancy. Cochrane Database Syst Rev. 2015 Jul 22;2015(7):CD004736. doi: 10.1002/14651858.CD004736.pub5. PMID: 26198451; PMCID: PMC8918165.
- 43. Brocklehurst P, Gordon A, Heatley E, Milan SJ. Antibiotics for treating bacterial vaginosis in pregnancy. Cochrane Database Syst Rev. 2013 Jan 31;(1):CD000262. doi: 10.1002/14651858.CD000262.pub4. PMID: 23440777.
- 44. Subtil D, Brabant G, Tilloy E, et al. Early clindamycin for bacterial vaginosis in pregnancy (PREMEVA): a multicentre, double-blind, randomised controlled trial. Lancet. 2018 Nov 17;392(10160):2171-2179. doi: 10.1016/S0140-6736(18)31617-9. Epub 2018 Oct 12. PMID: 30322724.
- 45. Fleming TP, Watkins AJ, Velazquez MA, et al. Origins of lifetime health around the time of conception: causes and consequences. Lancet. 2018 May 5;391(10132):1842-1852. doi: 10.1016/S0140-6736(18)30312-X. Epub 2018 Apr 16. PMID: 29673874; PMCID: PMC5975952.
- 46. Tobi EW, Goeman JJ, Monajemi R, et al. DNA methylation signatures link prenatal famine exposure to growth and metabolism. Nat Commun. 2014 Nov

- 26;5:5592. doi: 10.1038/ncomms6592. Erratum in: Nat Commun. 2015;6:7740. PMID: 25424739; PMCID: PMC4246417.
- 47. Maltepe E, Fisher SJ. Placenta: the forgotten organ. Annu Rev Cell Dev Biol. 2015;31:523-52. doi: 10.1146/annurev-cellbio-100814-125620. Epub 2015 Oct 5. PMID: 26443191.
- 48. Butte NF. Carbohydrate and lipid metabolism in pregnancy: normal compared with gestational diabetes mellitus. Am J Clin Nutr. 2000 May;71(5 Suppl):1256S-61S. doi: 10.1093/ajcn/71.5.1256s. PMID: 10799399.
- 49. Tunster SJ, Watson ED, Fowden AL, Burton GJ. Placental glycogen stores and fetal growth: insights from genetic mouse models. Reproduction. 2020 Jun;159(6):R213-R235. doi: 10.1530/REP-20-0007. PMID: 32191912.
- 50. Kaur H, Muhlhausler BS, Roberts CT, Gatford KL. The growth hormone-insulin like growth factor axis in pregnancy. J Endocrinol. 2021 Sep 1:JOE-21-0087.R1. doi: 10.1530/JOE-21-0087. PMID: 34479185.
- 51. Giustina A, Mazziotti G, Canalis E. Growth hormone, insulin-like growth factors, and the skeleton. Endocr Rev. 2008 Aug;29(5):535-59. doi: 10.1210/er.2007-0036. Epub 2008 Apr 24. PMID: 18436706; PMCID: PMC2726838.
- 52. Sferruzzi-Perri AN, Sandovici I, Constancia M, Fowden AL. Placental phenotype and the insulin-like growth factors: resource allocation to fetal growth. J Physiol. 2017 Aug 1;595(15):5057-5093. doi: 10.1113/JP273330. Epub 2017 May 23. PMID: 28337745; PMCID: PMC5538190.
- 53. Fasshauer M, Blüher M, Stumvoll M. Adipokines in gestational diabetes. Lancet Diabetes Endocrinol. 2014 Jun;2(6):488-99. doi: 10.1016/S2213-8587(13)70176-1. Epub 2013 Dec 30. PMID: 24731659.
- 54. Smits I, Hoftiezer L, van Dillen J, Hogeveen M. Neonatal hypoglycaemia and body proportionality in small for gestational age newborns: a retrospective cohort study. Eur J Pediatr. 2022 Oct;181(10):3655-3662. doi: 10.1007/s00431-022-04592-8. Epub 2022 Aug 18. PMID: 35980543; PMCID: PMC9508048.
- 55. Crane JP, Kopta MM. Comparative newborn anthropometric data in symmetric versus asymmetric intrauterine growth retardation. Am J Obstet Gynecol. 1980 Nov 1;138(5):518-22. doi: 10.1016/0002-9378(80)90279-3. PMID: 7191639.
- 56. Mesiano S, Chan EC, Fitter JT, Kwek K, Yeo G, Smith R. Progesterone withdrawal and estrogen activation in human parturition are coordinated by progesterone receptor A expression in the myometrium. J Clin Endocrinol Metab. 2002 Jun;87(6):2924-30. doi: 10.1210/jcem.87.6.8609. PMID: 12050275.

- 57. Myers KM, Socrate S, Paskaleva A, House M. A study of the anisotropy and tension/compression behavior of human cervical tissue. J Biomech Eng. 2010 Feb;132(2):021003. doi: 10.1115/1.3197847. PMID: 20370240.
- 58. Amabebe E, Anumba DOC. Mechanistic Insights into Immune Suppression and Evasion in Bacterial Vaginosis. Curr Microbiol. 2022 Feb 7;79(3):84. doi: 10.1007/s00284-022-02771-2. PMID: 35128579; PMCID: PMC8818625.
- 59. Cole AM. Innate host defense of human vaginal and cervical mucosae. Curr Top Microbiol Immunol. 2006;306:199-230. PMID: 16909923.
- 60. Hunter PJ, Sheikh S, David AL, Peebles DM, Klein N. Cervical leukocytes and spontaneous preterm birth. J Reprod Immunol. 2016 Feb;113:42-9. doi: 10.1016/j.jri.2015.11.002. Epub 2015 Nov 21. PMID: 26637953; PMCID: PMC4764650.
- 61. Condon JC, Jeyasuria P, Faust JM, Mendelson CR. Surfactant protein secreted by the maturing mouse fetal lung acts as a hormone that signals the initiation of parturition. Proc Natl Acad Sci U S A. 2004 Apr 6;101(14):4978-83. doi: 10.1073/pnas.0401124101. Epub 2004 Mar 25. PMID: 15044702; PMCID: PMC387359.
- 62. Garcia-Verdugo I, Leiber D, Robin P, Billon-Denis E, Chaby R, Tanfin Z. Direct interaction of surfactant protein A with myometrial binding sites: signaling and modulation by bacterial lipopolysaccharide. Biol Reprod. 2007 Apr;76(4):681-91. doi: 10.1095/biolreprod.106.058131. Epub 2007 Jan 3. PMID: 17202387.
- 63. Vannuccini S, Bocchi C, Severi FM, Challis JR, Petraglia F. Endocrinology of human parturition. Ann Endocrinol (Paris). 2016 Jun;77(2):105-13. doi: 10.1016/j.ando.2016.04.025. Epub 2016 May 5. PMID: 27155774.
- 64. Challis JR, Lye SJ, Gibb W. Prostaglandins and parturition. Ann N Y Acad Sci. 1997 Sep 26;828:254-67. doi: 10.1111/j.1749-6632.1997.tb48546.x. PMID: 9329846.
- 65. Sennström MB, Brauner A, Byström B, Malmström A, Ekman G. Matrix metalloproteinase-8 correlates with the cervical ripening process in humans. Acta Obstet Gynecol Scand. 2003 Oct;82(10):904-11. doi: 10.1080/j.1600-0412.2003.00249.x. PMID: 12956839.
- 66. Winkler M, Rath W. Changes in the cervical extracellular matrix during pregnancy and parturition. J Perinat Med. 1999;27(1):45-60. doi: 10.1515/JPM.1999.006. PMID: 10343934.
- 67. Saccone G, Saccone I, Berghella V. Omega-3 long-chain polyunsaturated fatty acids and fish oil supplementation during pregnancy: which evidence? J Matern Fetal Neonatal Med. 2016;29(15):2389-97. doi: 10.3109/14767058.2015.1086742. PMID: 26382010

- 68. Simmonds LA, Sullivan TR, Skubisz M, Middleton PF, Best KP, Yelland LN, Quinlivan J, Zhou SJ, Liu G, McPhee AJ, Gibson RA, Makrides M. Omega-3 fatty acid supplementation in pregnancy-baseline omega-3 status and early preterm birth: exploratory analysis of a randomised controlled trial. BJOG. 2020 Jul;127(8):975-981. doi: 10.1111/1471-0528.16168. Epub 2020 Mar 3. PMID: 32034969.
- 69. Middleton P, Gomersall JC, Gould JF, Shepherd E, Olsen SF, Makrides M. Omega-3 fatty acid addition during pregnancy. Cochrane Database Syst Rev. 2018 Nov 15;11(11):CD003402. doi: 10.1002/14651858.CD003402.pub3. PMID: 30480773; PMCID: PMC6516961.
- 70. Hanson MA, Bardsley A, De-Regil LM, et al. The International Federation of Gynecology and Obstetrics (FIGO) recommendations on adolescent, preconception, and maternal nutrition: "Think Nutrition First". Int J Gynaecol Obstet. 2015 Oct;131 Suppl 4:S213-53. doi: 10.1016/S0020-7292(15)30034-5. PMID: 26433230.
- 71. Sanghavi M, Rutherford JD. Cardiovascular physiology of pregnancy. Circulation. 2014 Sep 16;130(12):1003-8. doi: 10.1161/CIRCULATIONAHA.114.009029. PMID: 25223771.
- 72. McAuliffe F, Kametas N, Costello J, Rafferty GF, Greenough A, Nicolaides K. Respiratory function in singleton and twin pregnancy. BJOG. 2002 Jul;109(7):765-9. doi: 10.1111/j.1471-0528.2002.01515.x. PMID: 12135212.
- 73. Retief FP, Brink AJ. A study of pregnancy anaemia: blood volume changes correlated with other parameters of haemopoietic efficiency. J Obstet Gynaecol Br Commonw. 1967 Oct;74(5):683-93. doi: 10.1111/j.1471-0528.1967.tb03781.x. PMID: 6058531.
- 74. Dockree S, Shine B, Pavord S, Impey L, Vatish M. White blood cells in pregnancy: reference intervals for before and after delivery. EBioMedicine. 2021 Dec;74:103715. doi: 10.1016/j.ebiom.2021.103715. Epub 2021 Nov 23. PMID: 34826802; PMCID: PMC8626574.
- 75. Nakamura N, Miyazaki K, Kitano Y, Fujisaki S, Okamura H. Suppression of cytotoxic T-lymphocyte activity during human pregnancy. J Reprod Immunol. 1993 Mar;23(2):119-30. doi: 10.1016/0165-0378(93)90002-y. PMID: 8389871.
- 76. Wald A, Van Thiel DH, Hoechstetter L, et al. Effect of pregnancy on gastrointestinal transit. Dig Dis Sci. 1982 Nov;27(11):1015-8. doi: 10.1007/BF01391748. PMID: 7140485.
- 77. Kac G, Arnold CD, Matias SL, Mridha MK, Dewey KG. Gestational weight gain and newborn anthropometric outcomes in rural Bangladesh. Matern Child Nutr. 2019 Oct;15(4):e12816. doi: 10.1111/mcn.12816. Epub 2019 Apr 24. PMID: 30903801; PMCID: PMC6859981.

- 78. Shi H, Chen L, Wang Y, et al. Severity of Anemia During Pregnancy and Adverse Maternal and Fetal Outcomes. JAMA Netw Open. 2022 Feb 1;5(2):e2147046. doi: 10.1001/jamanetworkopen.2021.47046. PMID: 35113162; PMCID: PMC8814908.
- 79. Keats EC, Haider BA, Tam E, Bhutta ZA. Multiple-micronutrient supplementation for women during pregnancy. Cochrane Database Syst Rev. 2019 Mar 14;3(3):CD004905. doi: 10.1002/14651858.CD004905.pub6. PMID: 30873598; PMCID: PMC6418471.
- 80. Smith ER, Shankar AH, Wu LS, et al. Modifiers of the effect of maternal multiple micronutrient supplementation on stillbirth, birth outcomes, and infant mortality: a meta-analysis of individual patient data from 17 randomised trials in low-income and middle-income countries. Lancet Glob Health. 2017 Nov;5(11):e1090-e1100. doi: 10.1016/S2214-109X(17)30371-6. PMID: 29025632.
- 81. Lassi ZS, Padhani ZA, Rabbani A, et al. Impact of Dietary Interventions during Pregnancy on Maternal, Neonatal, and Child Outcomes in Low- and Middle-Income Countries. Nutrients. 2020 Feb 19;12(2):531. doi: 10.3390/nu12020531. PMID: 32092933; PMCID: PMC7071393.
- 82. Fakhraei R, Denize K, Simon A, et al. Predictors of Adverse Pregnancy Outcomes in Pregnant Women Living with Obesity: A Systematic Review. Int J Environ Res Public Health. 2022 Feb 12;19(4):2063. doi: 10.3390/ijerph19042063. PMID: 35206265; PMCID: PMC8872310.
- 83. Han Z, Lutsiv O, Mulla S, McDonald SD; Knowledge Synthesis Group. Maternal height and the risk of preterm birth and low birth weight: a systematic review and meta-analyses. J Obstet Gynaecol Can. 2012 Aug;34(8):721-746. doi: 10.1016/S1701-2163(16)35337-3. PMID: 22947405.
- 84. Colleoni F, Lattuada D, Garretto A, et al. Maternal blood mitochondrial DNA content during normal and intrauterine growth restricted (IUGR) pregnancy. Am J Obstet Gynecol. 2010 Oct;203(4):365.e1-6. doi: 10.1016/j.ajog.2010.05.027. Epub 2010 Jul 8. PMID: 20619387.
- 85. Burton GJ, Jauniaux E. Oxidative stress. Best Pract Res Clin Obstet Gynaecol. 2011 Jun;25(3):287-99. doi: 10.1016/j.bpobgyn.2010.10.016. PMID: 21130690; PMCID: PMC3101336.
- 86. Moore TA, Ahmad IM, Zimmerman MC. Oxidative Stress and Preterm Birth: An Integrative Review. Biol Res Nurs. 2018 Oct;20(5):497-512. doi: 10.1177/1099800418791028. PMID: 30068228; PMCID: PMC6346316.
- 87. Stone WL, Bailey B, Khraisha N. The pathophysiology of smoking during pregnancy: a systems biology approach. Front Biosci (Elite Ed). 2014 Jun 1;6(2):318-28. doi: 10.2741/e708. PMID: 24896208.

- 88. Pawlosky RJ, Hibbeln JR, Salem N Jr. Compartmental analyses of plasma n-3 essential fatty acids among male and female smokers and nonsmokers. J Lipid Res. 2007 Apr;48(4):935-43. doi: 10.1194/jlr.M600310-JLR200. PMID: 17234605.
- 89. Kuper SG, Abramovici AR, Jauk VC, Harper LM, Biggio JR, Tita AT. The effect of omega-3 supplementation on pregnancy outcomes by smoking status. Am J Obstet Gynecol. 2017 Oct;217(4):476.e1-476.e6. doi: 10.1016/j.ajog.2017.05.033. Epub 2017 May 23. PMID: 28549983; PMCID: PMC5614814.
- 90. Wessels I, Maywald M, Rink L. Zinc as a Gatekeeper of Immune Function. Nutrients. 2017 Nov 25;9(12):1286. doi: 10.3390/nu9121286. PMID: 29186856; PMCID: PMC5748737.
- 91. Naber TH, van den Hamer CJ, van den Broek WJ, Roelofs H. Zinc exchange by blood cells in nearly physiologic standard conditions. Biol Trace Elem Res. 1994 Oct-Nov;46(1-2):29-50. doi: 10.1007/BF02790066. PMID: 7888283.
- 92. Turner TC, Sok MCP, Hymel LA, et al. Harnessing lipid signaling pathways to target specialized pro-angiogenic neutrophil subsets for regenerative immunotherapy. Sci Adv. 2020 Oct 30;6(44):eaba7702. doi: 10.1126/sciadv.aba7702. PMID: 33127670; PMCID: PMC7608810.
- 93. Nadkarni S, Smith J, Sferruzzi-Perri AN, et al. Neutrophils induce proangiogenic T cells with a regulatory phenotype in pregnancy. Proc Natl Acad Sci U S A. 2016 Dec 27;113(52):E8415-E8424. doi: 10.1073/pnas.1611944114. Epub 2016 Dec 12. PMID: 27956610; PMCID: PMC5206541.
- 94. Yamamoto H, Flannery ML, Kupriyanov S, et al. Defective trophoblast function in mice with a targeted mutation of Ets2. Genes Dev. 1998 May 1;12(9):1315-26. doi: 10.1101/gad.12.9.1315. PMID: 9573048; PMCID: PMC316781.
- 95. Wilson RL, Leemaqz SY, Goh Z, et al. Zinc is a critical regulator of placental morphogenesis and maternal hemodynamics during pregnancy in mice. Sci Rep. 2017 Nov 9;7(1):15137. doi: 10.1038/s41598-017-15085-2. PMID: 29123159; PMCID: PMC5680205.
- 96. Chellam VG, Rushton DI. Chorioamnionitis and funiculitis in the placentas of 200 births weighing less than 2.5 kg. Br J Obstet Gynaecol. 1985 Aug;92(8):808-14. doi: 10.1111/j.1471-0528.1985.tb03050.x. PMID: 4027203.
- 97. Doyle RM, Harris K, Kamiza S, et al. Bacterial communities found in placental tissues are associated with severe chorioamnionitis and adverse birth outcomes. PLoS One. 2017 Jul 12;12(7):e0180167. doi: 10.1371/journal.pone.0180167. PMID: 28700642; PMCID: PMC5507499.
- 98. Al-Adnani M, Sebire NJ. The role of perinatal pathological examination in subclinical infection in obstetrics. Best Pract Res Clin Obstet Gynaecol. 2007

- Jun;21(3):505-21. doi: 10.1016/j.bpobgyn.2007.02.001. Epub 2007 Apr 19. PMID: 17448728.
- 99. Sprong KE, Mabenge M, Wright CA, Govender S. *Ureaplasma* species and preterm birth: current perspectives. Crit Rev Microbiol. 2020 Mar;46(2):169-181. doi: 10.1080/1040841X.2020.1736986. Epub 2020 Mar 6. PMID: 32141797.
- 100. Sweeney EL, Dando SJ, Kallapur SG, Knox CL. The Human Ureaplasma Species as Causative Agents of Chorioamnionitis. Clin Microbiol Rev. 2016 Dec 14;30(1):349-379. doi: 10.1128/CMR.00091-16. PMID: 27974410; PMCID: PMC5217797.
- de Goffau MC, Lager S, Sovio U, et al. Human placenta has no microbiome but can contain potential pathogens. Nature. 2019 Aug;572(7769):329-334. doi: 10.1038/s41586-019-1451-5. Epub 2019 Jul 31. Erratum in: Nature. 2019 Oct;574(7778):E15. PMID: 31367035; PMCID: PMC6697540.
- 102. Kranz A, Feierabend N, Sliwka D, Wiesegart A, Abele H, Graf J. Assessment of the Association of Periodontal Diseases in Pregnant Women and the Efficacy of Periodontal Treatment in the Context of Premature Births and Pregnancy Complications a Narrative Review. Geburtshilfe Frauenheilkd. 2022 Aug 10;82(8):831-841. doi: 10.1055/a-1868-4693. PMID: 35967744; PMCID: PMC9365472.
- 103. Gilman-Sachs A, Dambaeva S, Salazar Garcia MD, Hussein Y, Kwak-Kim J, Beaman K. Inflammation induced preterm labor and birth. J Reprod Immunol. 2018 Sep;129:53-58. doi: 10.1016/j.jri.2018.06.029. Epub 2018 Jun 30. PMID: 30025845.
- 104. Salvi V, Vaira X, Gianello V, et al.. TLR Signalling Pathways Diverge in Their Ability to Induce PGE2. Mediators Inflamm. 2016;2016:5678046. doi: 10.1155/2016/5678046. Epub 2016 Aug 18. PMID: 27630451; PMCID: PMC5007370.
- 105. Kyathanahalli C, Snedden M, Hirsch E. Is Human Labor at Term an Inflammatory Condition?†. Biol Reprod. 2022 Sep 29:ioac182. doi: 10.1093/biolre/ioac182. Epub ahead of print. PMID: 36173900.
- 106. Umbers AJ, Boeuf P, Clapham C, et al. Placental malaria-associated inflammation disturbs the insulin-like growth factor axis of fetal growth regulation. J Infect Dis. 2011 Feb 15;203(4):561-9. doi: 10.1093/infdis/jiq080. Epub 2011 Jan 7. PMID: 21216864; PMCID: PMC3071224.
- 107. Walters TD, Griffiths AM. Mechanisms of growth impairment in pediatric Crohn's disease. Nat Rev Gastroenterol Hepatol. 2009 Sep;6(9):513-23. doi: 10.1038/nrgastro.2009.124. PMID: 19713986.

- 108. Prendergast AJ, Rukobo S, Chasekwa B, et al. Stunting is characterized by chronic inflammation in Zimbabwean infants. PLoS One. 2014 Feb 18;9(2):e86928. doi: 10.1371/journal.pone.0086928. PMID: 24558364; PMCID: PMC3928146.
- 109. Hsiao EY, Patterson PH. Activation of the maternal immune system induces endocrine changes in the placenta via IL-6. Brain Behav Immun. 2011 May;25(4):604-15. doi: 10.1016/j.bbi.2010.12.017. Epub 2010 Dec 30. PMID: 21195166; PMCID: PMC3081363.
- 110. Keeler SM, Kiefer DG, Rust OA, et al. Comprehensive amniotic fluid cytokine profile evaluation in women with a short cervix: which cytokine(s) correlates best with outcome? Am J Obstet Gynecol. 2009 Sep;201(3):276.e1-6. doi: 10.1016/j.ajog.2009.05.045. PMID: 19733278.
- 111. EPPPIC Group. Evaluating Progestogens for Preventing Preterm birth International Collaborative (EPPPIC): meta-analysis of individual participant data from randomised controlled trials. Lancet. 2021 Mar 27;397(10280):1183-1194. doi: 10.1016/S0140-6736(21)00217-8. Erratum in: Lancet. 2021 Apr 17;397(10283):1446. PMID: 33773630.
- 112. Sibai B, Saade GR, Das AF, Gudeman J. Safety review of hydroxy-progesterone caproate in women with a history of spontaneous preterm birth. J Perinatol. 2021 Apr;41(4):718-725. doi: 10.1038/s41372-020-00849-y. Epub 2020 Oct 14. PMID: 33057132; PMCID: PMC8049867.
- 113. Chandiramani M, Seed PT, Orsi NM, et al. Limited relationship between cervico-vaginal fluid cytokine profiles and cervical shortening in women at high risk of spontaneous preterm birth. PLoS One. 2012;7(12):e52412. doi: 10.1371/journal.pone.0052412. Epub 2012 Dec 26. PMID: 23300664; PMCID: PMC3530581.
- 114. Tarca AL, Fitzgerald W, Chaemsaithong P, et al. The cytokine network in women with an asymptomatic short cervix and the risk of preterm delivery. Am J Reprod Immunol. 2017 Sep;78(3):e12686. doi: 10.1111/aji.12686. Epub 2017 Jun 6. PMID: 28585708; PMCID: PMC5575567.
- 115. Poon LC, Shennan A, Hyett JA, et al. The International Federation of Gynecology and Obstetrics (FIGO) initiative on pre-eclampsia: A pragmatic guide for first-trimester screening and prevention. Int J Gynaecol Obstet. 2019 May;145 Suppl 1(Suppl 1):1-33. doi: 10.1002/ijgo.12802. Erratum in: Int J Gynaecol Obstet. 2019 Sep;146(3):390-391. PMID: 31111484; PMCID: PMC6944283.
- 116. Thilaganathan B, Kalafat E. Cardiovascular System in Preeclampsia and Beyond. Hypertension. 2019 Mar;73(3):522-531. doi: 10.1161/HYPERTENSIONAHA.118.11191. PMID: 30712425; PMCID: PMC6380450.

- 117. Craici I, Wagner S, Garovic VD. Preeclampsia and future cardiovascular risk: formal risk factor or failed stress test? Ther Adv Cardiovasc Dis. 2008 Aug;2(4):249-59. doi: 10.1177/1753944708094227. PMID: 19124425; PMCID: PMC2674507.
- 118. Duley L, Meher S, Hunter KE, Seidler AL, Askie LM. Antiplatelet agents for preventing pre-eclampsia and its complications. Cochrane Database Syst Rev. 2019 Oct 30;2019(10):CD004659. doi: 10.1002/14651858.CD004659.pub3. PMID: 31684684; PMCID: PMC6820858
- 119. Smith DD, Costantine MM. The role of statins in the prevention of preeclampsia. Am J Obstet Gynecol. 2022 Feb;226(2S):S1171-S1181. doi: 10.1016/j.ajog.2020.08.040. Epub 2020 Aug 17. PMID: 32818477; PMCID: PMC8237152.
- 120. Masotti G, Galanti G, Poggesi L, Abbate R, Neri Serneri GG. Differential inhibition of prostacyclin production and platelet aggregation by aspirin. Lancet. 1979 Dec 8;2(8154):1213-7. doi: 10.1016/s0140-6736(79)92334-1. PMID: 92623.
- 121. Rolnik DL, Nicolaides KH, Poon LC. Prevention of preeclampsia with aspirin. Am J Obstet Gynecol. 2022 Feb;226(2S):S1108-S1119. doi: 10.1016/j.ajog.2020.08.045. Epub 2020 Aug 21. PMID: 32835720.
- 122. Stewart CP, Oaks BM, Laugero KD, et al. Maternal cortisol and stress are associated with birth outcomes, but are not affected by lipid-based nutrient supplements during pregnancy: an analysis of data from a randomized controlled trial in rural Malawi. BMC Pregnancy Childbirth. 2015 Dec 22;15:346. doi: 10.1186/s12884-015-0793-8. PMID: 26694646; PMCID: PMC4688934.
- 123. Shapiro GD, Fraser WD, Frasch MG, Séguin JR. Psychosocial stress in pregnancy and preterm birth: associations and mechanisms. J Perinat Med. 2013 Nov;41(6):631-45. doi: 10.1515/jpm-2012-0295. PMID: 24216160; PMCID: PMC5179252.
- 124. Christian LM, Glaser R, Porter K, Iams JD. Stress-induced inflammatory responses in women: effects of race and pregnancy. Psychosom Med. 2013 Sep;75(7):658-69. doi: 10.1097/PSY.0b013e31829bbc89. Epub 2013 Jul 19. PMID: 23873713; PMCID: PMC3788648.
- 125. Coussons-Read ME, Okun ML, Schmitt MP, Giese S. Prenatal stress alters cytokine levels in a manner that may endanger human pregnancy. Psychosom Med. 2005 Jul-Aug;67(4):625-31. doi: 10.1097/01.psy.0000170331.74960.ad. PMID: 16046378.
- 126. Coussons-Read ME, Okun ML, Nettles CD. Psychosocial stress increases inflammatory markers and alters cytokine production across pregnancy. Brain Behav Immun. 2007 Mar;21(3):343-50. doi: 10.1016/j.bbi.2006.08.006. Epub 2006 Oct 6. PMID: 17029703.

- 127. Zdravkovic T, Genbacev O, McMaster MT, Fisher SJ. The adverse effects of maternal smoking on the human placenta: a review. Placenta. 2005 Apr;26 Suppl A:S81-6. doi: 10.1016/j.placenta.2005.02.003. PMID: 15837073.
- 128. Siddiqui AR, Gold EB, Yang X, Lee K, Brown KH, Bhutta ZA. Prenatal exposure to wood fuel smoke and low birth weight. Environ Health Perspect. 2008 Apr;116(4):543-9. doi: 10.1289/ehp.10782. PMID: 18414641; PMCID: PMC2290983.
- 129. Moore LG. Fetal growth restriction and maternal oxygen transport during high altitude pregnancy. High Alt Med Biol. 2003 Summer;4(2):141-56. doi: 10.1089/152702903322022767. PMID: 12855048.
- 130. Chamberlain C, O'Mara-Eves A, Porter J, et al. Psychosocial interventions for supporting women to stop smoking in pregnancy. Cochrane Database Syst Rev. 2017 Feb 14;2(2):CD001055. doi: 10.1002/14651858.CD001055.pub5. PMID: 28196405; PMCID: PMC6472671.
- 131. Kabir Z, Clarke V, Conroy R, McNamee E, Daly S, Clancy L. Low birthweight and preterm birth rates 1 year before and after the Irish workplace smoking ban. BJOG. 2009 Dec;116(13):1782-7. doi: 10.1111/j.1471-0528.2009.02374.x.
- 132. Faber T, Kumar A, Mackenbach JP, Millett C, Basu S, Sheikh A, Been JV. Effect of tobacco control policies on perinatal and child health: a systematic review and meta-analysis. Lancet Public Health. 2017 Sep 5;2(9):e420-e437. doi: 10.1016/S2468-2667(17)30144-5. PMID: 28944313; PMCID: PMC5592249.
- 133. Díez-Izquierdo A, Balaguer A, Lidón-Moyano C, et al. Correlation between tobacco control policies and preterm births and low birth weight in Europe. Environ Res. 2018 Jan;160:547-553. doi: 10.1016/j.envres.2017.10.033. Epub 2017 Oct 28. PMID: 29089104.
- 134. World Health Organization Regional Office for Europe. Air quality guidelines: global update 2005: particulate matter, ozone, nitrogen dioxide and sulfur dioxide. World Health Organization. Regional Office for Europe; 2006.
- 135. Austin KF, Mejia MT. Household air pollution as a silent killer: women's status and solid fuel use in developing nations. Popul Environ. 2017 Sep;39(1):1–25.
- 136. Katz J, Tielsch JM, Khatry SK, et al. Impact of Improved Biomass and Liquid Petroleum Gas Stoves on Birth Outcomes in Rural Nepal: Results of 2 Randomized Trials. Glob Health Sci Pract. 2020 Sep 30;8(3):372–82.
- 137. Clasen TF, Chang HH, Thompson LM, et al. Liquefied Petroleum Gas or Biomass for Cooking and Effects on Birth Weight. N Engl J Med. 2022 Nov 10;387(19):1735-1746. doi: 10.1056/NEJMoa2206734. Epub 2022 Oct 10. PMID: 36214599.

- 138. Kuehn L, McCormick S. Heat Exposure and Maternal Health in the Face of Climate Change. Int J Environ Res Public Health. 2017 Jul 29;14(8):853. doi: 10.3390/ijerph14080853. PMID: 28758917; PMCID: PMC5580557.
- 139. Wang J, Tong S, Williams G, Pan X. Exposure to Heat Wave During Pregnancy and Adverse Birth Outcomes: An Exploration of Susceptible Windows. Epidemiology. 2019 Jul;30 Suppl 1:S115-S121. doi: 10.1097/EDE.0000000000000995. PMID: 31181014.
- 140. Olivier K, Reinders LA, Clarke MW, Crew RC, Pereira G, Maloney SK, Wyrwoll CS. Maternal, Placental, and Fetal Responses to Intermittent Heat Exposure During Late Gestation in Mice. Reprod Sci. 2021 Feb;28(2):416-425. doi: 10.1007/s43032-020-00291-7. Epub 2020 Aug 17. PMID: 32804351.
- 141. Ghulam Mohyuddin S, Khan I, Zada A, et al. Influence of Heat Stress on Intestinal Epithelial Barrier Function, Tight Junction Protein, and Immune and Reproductive Physiology. Biomed Res Int. 2022 Sep 1;2022:8547379. doi: 10.1155/2022/8547379. PMID: 36093404; PMCID: PMC9458360.
- 142. Wan J, Chen B, Rao J. Occurrence and preventive strategies to control mycotoxins in cereal-based food. Compr Rev Food Sci Food Saf. 2020 May;19(3):928-953. doi: 10.1111/1541-4337.12546. Epub 2020 Mar 4. PMID: 33331688.
- 143. Wu Y, Wang J, Wei Y, et al. Maternal exposure to endocrine disrupting chemicals (EDCs) and preterm birth: A systematic review, meta-analysis, and meta-regression analysis. Environ Pollut. 2022 Jan 1;292(Pt A):118264. doi: 10.1016/j.envpol.2021.118264. Epub 2021 Oct 1. PMID: 34606968.
- 144. Hofmeyr GJ, Black RE, Rogozińska E, et al. Evidence-based antenatal interventions to reduce the incidence of small vulnerable newborns and their associated poor outcomes. Lancet SVN series paper 4.
- 145. Shynlova O, Nadeem L, Dorogin A, Mesiano S, Lye SJ. The selective progesterone receptor modulator-promegestone-delays term parturition and prevents systemic inflammation-mediated preterm birth in mice. Am J Obstet Gynecol. 2022 Feb;226(2):249.e1-249.e21. doi: 10.1016/j.ajog.2021.08.013. Epub 2021 Aug 19. PMID: 34418351; PMCID: PMC8810746.
- 146. Peeling RW, Mabey D, Herring A, Hook EW 3rd. Why do we need quality-assured diagnostic tests for sexually transmitted infections? Nat Rev Microbiol. 2006 Dec;4(12):909-21. doi: 10.1038/nrmicro1555. PMID: 17109030.
- 147. Nakahara A, Nair S, Ormazabal V, et al. Circulating Placental Extracellular Vesicles and Their Potential Roles During Pregnancy. Ochsner J. 2020 Winter;20(4):439-445. doi: 10.31486/toj.20.0049. PMID: 33408584

148. Awoyemi T, Tannetta D, Zhang W, et al. Glycosylated Siglec-6 expression in syncytiotrophoblast-derived extracellular vesicles from preeclampsia placentas. Biochem Biophys Res Commun. 2020 Dec 17;533(4):838-844. doi: 10.1016/j.bbrc.2020.09.081. PMID: 32998819.

Table 1. Changes to organ systems in women during pregnancy.

Organ system	Change		
Heart	Cardiac output increases by 50%. ⁷¹		
Lungs	Ventilation (volume/minute) increases by 50%. ⁷²		
Vasculature	Vascular resistance decreases by 30 – 50%. ⁷¹		
Red blood cells	Early 10% decrease in RBC and hemoglobin per volume		
	due to increase in plasma volume. ⁷³		
White blood cells	Circulating neutrophil counts increase by 50%. ⁷⁴		
	T cells become less responsive to antigenic stimulation. ⁷⁵		
Gastro-intestinal tract	Transit time slows down, possibly to allow longer time		
	for absorption of nutrients. ⁷⁶		
Pancreas	Small increase in insulin production in response to mild		
	insulin resistance in maternal tissues. ⁴⁸		

Table 2. Nutritional factors related to the small vulnerable newborn

Maternal nutritional factor	Potential mechanistic pathways	Outcomes
Nutrient supply (energy and	Energy and nutrient delivery to the	Growth
macronutrients:	placenta and fetus. ⁸¹	restriction
carbohydrates, proteins,		
lipids)		
Body composition	Underweight or low GWG: low energy	Growth
(underweight, overweight);	supply. ⁸¹	restriction
gestational weight gain	Overweight or excess GWG: metabolic	
(GWG)	and hormonal dysregulation, gestational	
	diabetes, hypertension, inflammation. ⁸²	
Dietary quality	Metabolic and hormonal dysregulation,	Growth
	gestational diabetes, hypertension,	restriction,
	inflammation, oxidative stress.	preterm birth
Stature	Small "container effect" on uterine and	Growth
	placental size. ⁸³	restriction
Micronutrients related to	Oxygen supply to placenta and fetus.	Growth
cardiac function, anaemia		restriction,
and oxygen supply (e.g.,		preterm birth
iron, riboflavin, folic acid,		
vitamin B12, vitamin C)		
Nutrients that support	Ability to fight infection and control	Fetal growth
immune function (e.g., zinc,	inflammation.	restriction,
fatty acids, vitamin D, iron)		preterm birth
Antioxidants and cofactors	Ability to reduce and repair damage	Fetal growth
of antioxidant enzymes (e.g.,	caused by oxidative stress.	restriction,
vitamins C, E, carotenoids,		preterm birth
copper, zinc, fatty acids)		
Nutrients related to cortisol	Control of inflammation, prevention of	Fetal growth
metabolism (e.g., fatty acids,	preterm COX2 activation and	restriction,
zinc, magnesium)	prostaglandin production.	preterm birth
Nutrients related to	Mitochondrial efficiency and protection	Fetal growth
mitochondrial function (e.g.,	against oxidative stress. ⁸⁴	restriction
vitamins C and E, zinc,		
copper, iodine, selenium)		
Nutrients related to	Omega-3 fatty acids: competitive	Preterm birth
production of prostaglandins	inhibition of preterm production of	
(e.g., long chain poly-	prostaglandins E2 and F2α from	
unsaturated fatty acids)	arachidonic acid. ⁶⁷	

Panel 1. Risk factors for the birth of a small vulnerable newborn

Undernutrition	Infection	Characteristics of woman and	Environmental exposures
Anaemia ²	HIV ⁸	pregnancy	and psychosocial stress
Zinc deficiency ³ Calcium deficiency ⁴ Short stature ⁵ Low BMI ⁶ Inadequate weight gain ⁷	Malaria ⁹ Syphilis ¹⁰ Chlamydia ¹¹ Gonorrhoea ¹² Urinary tract infection ¹³ Bacterial vaginosis ¹⁴ Trichomonas vaginalis ¹⁵ Group B Streptococcus ¹⁶	First pregnancy ¹⁷ Adolescent pregnancy ¹⁷ Short interpregnancy interval ¹⁸ Extreme parity ¹⁷ Older age ¹⁷ Preeclampsia ¹⁹ Placental dysfunction ²⁰ Gestational diabetes ²¹ Hypothyroidism ²² Cervical weakness ²³ Uterine malformations ²⁴ Endometriosis ²⁵ Multiple pregnancy ²⁶	Unwanted pregnancy ²⁷ Intimate partner abuse ²⁸ Lack of support or agency ²⁵ Mental illness ³⁰ Smoking ³¹ Alcohol abuse ³² Drug abuse ³³ Toxins ³⁴ Endocrine disruptors ³⁵ Indoor air pollution ³⁶ Outdoor air pollution ³⁷ Heat waves ³⁸ High altitude ³⁹

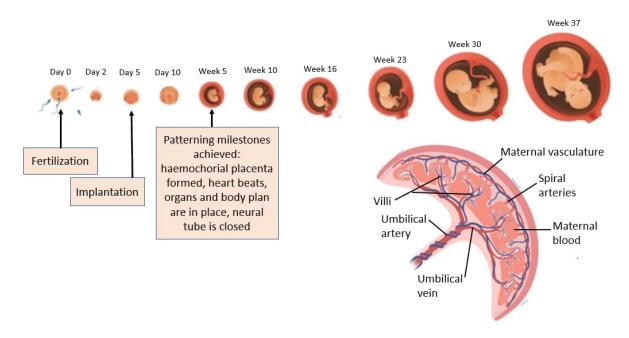


Figure 1. Developing fetus and fully developed placenta. The basic body plan with rudimentary organs are in place by 5 weeks post fertilization. The umbilical artery carries deoxygenated, waste-replete, nutrient-depleted fetal blood to the placental villi where waste is exchanged for nutrients and carbon dioxide is exchanged for oxygen from maternal blood.

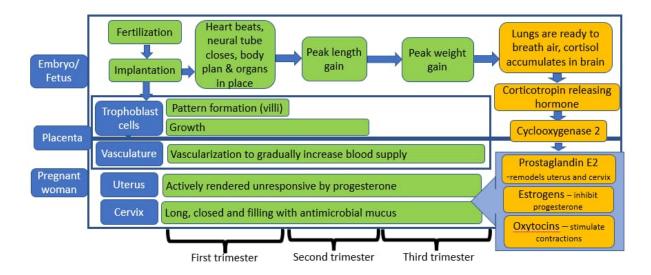


Figure 2. Conceptual model of key determinants of gestational length. When the fetus is ready to be born, cortisol enters the placenta and circulation and activates cyclooxygenase-2 to generate prostaglandin E2, which directs cervical and uterine remodelling. Estrogens override the suppressive effects of progesterone and oxytocins trigger uterine contractions. CRH – corticotropin releasing hormone.

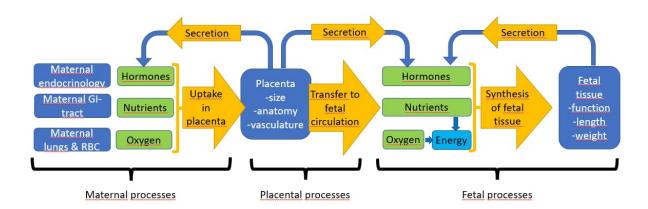


Figure 3. Conceptual model of key determinants of fetal growth. Hormones, nutrients and oxygen from the mother are taken up by the placenta and transferred to the fetal circulation to support synthesis of fetal tissue.

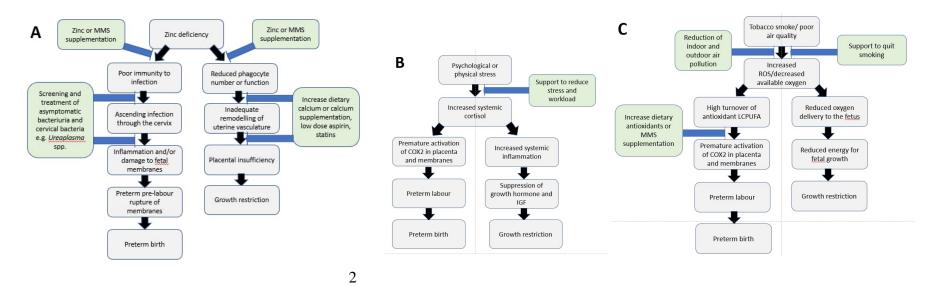


Figure 4. Examples of exposures that are able to contribute to both preterm birth and growth restriction via different pathways and ways to intervene toward prevention. Zinc deficiency (A), psychological and physical stress (B) and poor air quality/tobacco smoke (C) contribute to the birth of a small vulnerable newborn. MMS – multiple micronutrient supplements, COX2 - cyclooxygenase 2, LCPUFA – long chain polyunsaturated fatty acids, IGF – insulin-like growth factor, ROS – reactive oxygen species.

7

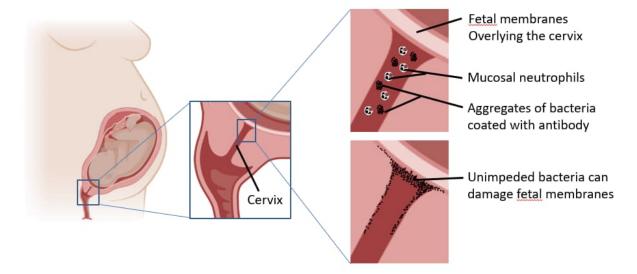


Figure 5. Immune defence of the cervix. The cervix remains long and closed for the duration of pregnancy. It is defended by antimicrobial chemicals including peptides, antibodies and enzymes. Neutrophils are also present in the mucus and are able to destroy invading microbes. In the absence of adequate immune defence, bacteria are able to colonize and damage the membranes leading to rupture or chorioamnionitis.